

**Taxonomy, phylogeny and biogeography of Central
European *Kybos* (Insecta: Hemiptera: Cicadellidae)**

Inauguraldissertation

zur
Erlangung der Würde eines Doktors der Philosophie
vorgelegt der
Philosophisch-Naturwissenschaftlichen Fakultät
der Universität Basel

von

Roland Mühlethaler
aus Basel (Basel-Stadt)

Basel, 2008

Genehmigt von der Philosophisch-Naturwissenschaftlichen Fakultät

auf Antrag von

Prof. Dr. Peter Nagel
PD Dr. Daniel Burckhardt
Prof. Dr. Hannelore Hoch

Basel, den 27. März 2007

Dekan
Prof. Dr. Hans-Peter Hauri

Contents

1 Introduction.....	3
2 Material and methods	7
3. Results.....	10
3.1 Taxonomy	10
3.1.1 Morphological character assessment	10
3.1.2 Morphological species concept	11
3.1.3 Species descriptions.....	12
3.1.4 Key to <i>Kybos</i> species	34
3.1.5 Molecular evidence.....	35
3.1.6 Bioacoustics	40
3.2 Phylogeny	42
3.2.1 Morphological analysis.....	39
3.2.2 Molecular analyses	46
3.3 Distribution and host associations	49
4 Discussion	51
4.1 Taxonomy	51
4.2 Phylogeny	53
4.3 Biogeography	53
4.4 Host plants.....	54
5 Conclusions.....	56
6 Acknowledgments	58
7 References.....	59
Appendix 1 Material examined.....	64
Appendix 2 COI sequences.....	74
Appendix 3 16S sequences	82
Appendix 4 Distribution and host plant information of the known <i>Kybos</i> species.....	86

List of figures

1. Male genitalia of <i>Kybos strigilifer</i>	21
2. Male aedeagi (<i>abstrusus</i> , <i>butleri</i> , <i>limpidus</i> , <i>mesasiaticus</i> , <i>populi</i> , <i>rufescens</i> , <i>sordidulus</i> , <i>aetnicola</i> , <i>austriacus</i> , <i>calyculus</i> , <i>candelabricus</i> , <i>digitatus</i> , <i>ludus</i> , <i>lindbergi</i> , <i>mucronatus</i>).....	22
3. Male aedeagi (<i>strigilifer</i> , <i>strobli</i> , <i>smaragdulus</i> , <i>virgator</i>).....	23
4. Male anal collars of <i>Kybos</i> spp.	24
5. Male pygofers of <i>Kybos</i> spp.....	25
6. Male apodemes (<i>abstrusus</i> , <i>aetnicola</i> , <i>butleri</i> , <i>calyculus</i> , <i>candelabricus</i> , <i>digitatus</i> , <i>limpidus</i> , <i>lindbergi</i> , <i>mesasiaticus</i> , <i>mucronatus</i> , <i>populi</i> , <i>rufescens</i>).....	26
7. Male apodemes (<i>smaragdulus</i> , <i>sordidulus</i> , <i>strigilifer</i> , <i>virgator</i>).....	27
8. Female ovipositors (<i>abstrusus</i> , <i>butleri</i> , <i>limpidus</i> , <i>populi</i> , <i>rufescens</i> , <i>austriacus</i>).....	28
9. Female ovipositors (<i>lindbergi</i> , <i>smaragdulus</i> , <i>strobli</i> , <i>digitatus</i> , <i>strigilifer</i> , <i>virgator</i>).....	29
10. Female ovipositors (<i>mucronatus</i> , <i>smaragdulus</i>).....	30
11. Female valvifers (<i>Kyboasca bipunctata</i> , <i>Kybos butleri</i> , <i>lindbergi</i> , <i>limpidus</i> , <i>mesasiaticus</i> , <i>populi</i> , <i>rufescens</i> , <i>smaragdulus</i>).....	31
12. Female valvifers (<i>strigilifer</i> , <i>strobli</i> , <i>virgator</i>).....	32
13. Coloration patterns of forewings and nymphs	33
14. COI sequences of the two genotypes of <i>Kybos butleri</i>	37
15. 16S sequences of the two genotypes of <i>Kybos butleri</i>	38
16. COI sequences of all individuals of <i>Kybos rufescens</i>	39
17. Sonograms and Spectrograms	41
18. Cladogram (morphology compared to host plant information)	45
19. Cladogram (COI).....	47
20. Cladogram (16S)	48
21. Distribution of <i>K. lindbergi</i> and <i>K. ludus</i> in Europe	50

List of tables

1. Morphological characters used for cladistic analysis.....	43
2. Matrix of the morphological characters used for the cladistic analysis	44

1 Introduction

Groups of phytophagous insects are usually more species-rich than their non-phytophagous sister clades. Host shifts (= new ecological niches) as well as coevolution with their host plants have been advocated as underlying mechanism for this richness (Futuyma & Mitter, 1996; Price, 1997). Another independent factor triggering speciation is geographic vicariance and dispersal (Burckhardt & Ouvrard, 2007). For investigating these evolutionary models detailed knowledge of the taxonomic and phylogenetic relationships is required. Traditionally taxonomy was based on morphology which provides in most cases good characters for diagnosing species. In examples where morphology currently fails this is most often due to poor work rather than genuine absence of distinctive characters. In some cases diagnostic characters are known for one sex or one stage only, and association of females and nymphs with identified males is difficult. For some time now molecular techniques are in use, which potentially help defining species and associating different sexes and instars of the same species. A relatively new practice is the so called “DNA barcoding” which is using the mitochondrial COI sequence. The initial euphorism, however, was dampened by recent studies who demonstrated the unsuitability as the universal gene for discrimination of animal species (Tautz *et al.*, 2003; Wheeler, 2005; Meier, 2006).

Within exopterygote insects, the Hemiptera constitutes with about 80,000-100,000 described species the most species-rich order. They belong to the “Big Five” (orders with around 100,000 and more species) of the global biodiversity. They are characterised by the unique sucking mouth parts for extracting liquids from plants, animals or fungi. They occupy a wide range of habitats although a majority is terrestrial and phytophagous. Only the suborder Heteroptera contains aquatic and carnivorous taxa. Unsurprisingly Hemiptera form a morphologically and biologically extremely diverse group (Carver, Gross & Woodward, 1991; Grimaldi & Engel, 2005; Hodkinson & Casson, 1991; Kristensen, 1991).

Traditionally the order Hemiptera comprises the suborders Sternorrhyncha, Auchenorrhyncha, Coleorrhyncha and Heteroptera (Dolling, 1991). The probably paraphyletic Auchenorrhyncha with some 40,000 described species worldwide is composed of the two well supported monophyletic Fulgoromorpha and Cicadomorpha. Recent analyses of hemipteran phylogeny are still controversial (Cryan, 2005; Yoshizawa & Saigusa, 2001), though the Sternorrhyncha, Fulgoromorpha, Cicadomorpha, Coleorrhyncha and Heteroptera are considered monophyletic by most authors (Bourgoin & Campbell, 2002; Dietrich, 2002; Nielson, 1985). Sorensen *et al.* (1995) proposed the formal classification with the suborders

Sternorrhyncha, Clypeorrhyncha (for extent Cicadomorpha), Archaeorrhyncha (for Fulgoromorpha) and Prosorrhyncha (for Coleorrhyncha and Heteroptera).

Cicadomorpha can be distinguished from other Hemiptera by following characters: postclypeus enlarged; antennal pedicel small, without conspicuous sensilla, flagellum aristiform; tegulae absent; anal veins of forewing usually separated from base to apex; mesocoxae small and narrowly separated. According to Dietrich (2002) the Cicadomorpha comprises the cicadas (Cicadoidea), spittlebugs and froghoppers (Cercopoidea), as well as leafhoppers, sharpshooters and treehoppers (Membracoidea s.l.). He defines the superfamilies mainly with morphological characters, and estimates that about 6-10% of plant-feeding insects belong to the Cicadomorpha. To date, 30,000 species have been described in over 5,000 genera and 13 families. They feed either on phloem (Membracidae), on xylem (Cicadoidea, Cercopoidea and many Cicadellinae) or on parenchyma (Typhlocybinae). Many taxa of the superfamily Membracoidea are of economic importance. They transmit phytopathogenic organisms causing plant diseases (Bourgoin & Campbell, 2002; Carver *et al.*, 1991; Dietrich, 2005; Kristensen, 1991; Nielson, 1985).

Cicadellidae is the largest family of Membracoidea with 50 subfamilies and 17,000-20,000 described species (Dolling, 1991; Hodkinson & Casson, 1991). Leafhoppers (Cicadellidae) live in all biogeographical regions and feed on a wide range of host plants, though individual species have often trophically and geographically restricted ranges. Eight subfamilies of Cicadellidae occur almost exclusively in the Australian Region, five in the Neotropics, five in the Palaearctics, one in the Nearctics and the rest is present in two or more biogeographical regions; thirteen are cosmopolitan (Dolling, 1991; Nielson, 1985).

Despite the economic importance, there are surprisingly many gaps in the knowledge on the taxonomy, phylogeny, life history and biology of cicadellids. Generally the species identification of Cicadomorpha is difficult due to their homogenous morphology, the huge number of species and the paucity of identification keys (Dietrich, 2005; Nielson, 1985).

Within the Cicadellidae one of the most advanced subfamilies is the Typhlocybinae with respect to morphology and biology. This subfamily is cosmopolitan and the morphology is remarkably uniform compared to other subfamilies of leafhoppers. The body size is small (2.0-5.5 mm), adults are always macropterous with reduced forewing venation lacking forks or cross-veins. Most species feed on leaf mesophyll cells of Dicotyledones. This is an evolutionary derived and specialised method of plant feeding causing white spots on leaves. The spots are in fact empty air-filled cells which can be seen on a variety of herbaceous plants. Exceptions are members of the tribe Empoascini feeding both on parenchyma and

phloem, and *Empoasca vitis* and *Jacobiasca libyca* on phloem only. Unlike other Cicadomorpha and Fulgoromorpha, Typhlocybinae lack secondary microbial endosymbionts. Parenchyma cells seem to contain all necessary nutrients (Dolling, 1991; Nickel, 2003; Nielson, 1985).

The eggs are laid with the ovipositor in slits in plant tissue. As most Hemiptera the Typhlocybinae have five nymphal instars. In cool or temperate regions the species hibernate as egg, nymph or adult. Depending on the climatic environment they produce one or more generations per year (Dolling, 1991; Nielson, 1985).

Mostly morphological characters of adults have been used for the classification of leafhoppers, e.g. the type, form and venation of the wings, the structure of the head, the position of the ocelli and the male genitalia. On tribal and generic levels the venation of the fore and hind wings are used (DeLong, 1971). Male genitalia and sound producing organs (apodemes) have been considered the only reliable characters for species separation. Identification keys are based on these characters (e.g. Ribaut, 1936; Le Quesne & Payne, 1981; Ossiannilsson, 1981). Closely related sympatric species are often difficult to separate by these characters for following reasons: the male genitalia are subjected to intraspecific variation which has, however, not been thoroughly examined in all species and some species have been described from a few males based on subtle genital differences only; the shape of the apodemes depends on the age of an individual which was not taken into consideration by some authors when describing new species. Data on host plant, behaviour and life history are still unknown for most species (Blocker & Triplehorn, 1985; Günthart, 1977; Nielson, 1985).

In the last decades bioacoustic techniques became increasingly important in insect taxonomy (Claridge, 2006). All leafhoppers and planthoppers produce substrate transmitted acoustic signals in contrast to the loud and prominent airborne songs of cicadas. Still little is known about the mechanisms of sound production in leafhoppers. Until the pioneer work of Ossiannilsson (1946, 1949) only the airborne calls of cicadas were known. With simple techniques he demonstrated that many species of leaf- and planthoppers produce sounds. Their calls consist usually of pulses with characteristic patterns of amplitude modulation. Male calls are normally more sophisticated than those of females. The vibrations are transmitted to the plant substrate through the tarsi and sometimes through the inserted stylets of the mouth. One of the main purposes of these so called songs is to attract and find mates for reproduction. Closely related species tend to differ clearly in their songs (Claridge, 2006; Cryan, 2005; Drosopoulos, 2006). In several different genera of Cicadellidae the taxonomy of

sibling species was clarified by song differences (Shaw, Vargo & Carlson, 1974; Claridge, 1985).

A good example to illustrate the taxonomic problems typical for Typhlocybinae is the genus *Empoasca* Walsh with some 500 described species. The genus is currently divided into three subgenera: *Empoasca* s.str., *Kybos* Fieber and *Kyboasca* Zachvatkin. The Holarctic subgenus *Kybos* comprises some 110 species, 42 of which are restricted to the Palaearctic Region. *Kybos* species are usually monophagous on *Salix*, *Populus*, *Alnus* or *Betula* spp. The taxonomy, phylogeny and biogeography of *Kybos* are mostly unknown. Many species have been described after single individuals and differences between some species are subtle. Only two papers (Ross, 1963; Dworakowska, 1976) deal with the taxonomy and phylogeny of *Kybos*. The monophyly of the genus *Empoasca* nor of any of its subgenera has been tested so far.

Here I revise the taxonomy and phylogeny of the Central European *Kybos* species. I investigate standard morphological (male genitalia, apodemes), molecular (the protein coding cytochrome c oxidase subunit I – COI – of the mitochondrial DNA and the ribosomal subunit rRNA – 16S) and bioacoustic methods for their taxonomic suitability. I examine females and last instar nymphs for taxonomically relevant characters. The improved taxonomic base is used for investigating host plant and biogeographical patterns.

2 Material and methods

2.1 Material examined

A total of 1098 specimens and 22 European *Kybos* species were examined (Appendix 1). Additionally six specimens of five Central Asian species and one specimen of an unidentified Nearctic *Kybos* species were added to the molecular data set. The closely related *E. vitis* (Göthe, 1875) and *K. bipunctata* (Oshanin, 1871) were used for outgroup comparison.

Seventeen *Kybos* species were collected and examined for the molecular studies. Totally 56 specimens and 23 species from different localities were examined (Appendix 1). Most of the insects were collected using a sweep net and an aspirator, stored in 96 % pure ethanol and identified by R.M. The samples of H. Nickel were collected in malaise traps and stored in 70% alcohol. As control and outgroup for the molecular analyses we chose the spittle bug *Philaenus spumarius* (L.) (Hemiptera, Cercopidae) taking the sequence data from GenBank.

2.2 Morphological studies

For the morphological studies dry mounted specimens from museum collections and freshly collected individuals were examined. For the observations of the genital structures with the light microscope the abdomen of fresh or dried specimens were removed and cleared in a solution of hot 10% KOH. After cleaning in distilled water and 70% alcohol the genitalia were stained with chlorazol black if necessary. Drawings and digital photographs of the genitalia were made in glycerin or glycerine-gelatine using a Leica DMLB light microscope. A camera lucida was used to produce drawings. The software Automontage (Syncroscopy®) was used to produce perfectly focused pictures with high resolution. The female abdomens used for SEM photographs were detached from the dried body and the abdominal sternite 7 was removed to render the base of the ovipositor visible. All SEM pictures were made at the Zentrum für Elektronenmikroskopie (ZBM, University Basel) with a scanning electron microscope Philips XL30 FEG.

The species descriptions and the identification key were made using the software DELTA (DEscription Language for TAxonomy), version 1.04 (Dallwitz, 1980; Paine & Zurcher, 1999). The morphological terminology follows Ossiannilsson (1978) and Davis (1975).

2.3 DNA sequences

We extracted mitochondrial DNA from specimens preserved in 96 % pure ethanol using a modification of the NaCl-extraction technique of Medrano, Aasen & Sharrow (1990). A 769bp sequence of the cytochrome oxidase I gene (COI), corresponding to positions 2265-3033 of the *Drosophila yakuba* mtDNA (Clary & Wolstenholme, 1985), was amplified using primers C1-J-1718 and C1-N-2191 (Nancy). A 388bp sequence of the 16S rRNA gene corresponding to positions 12295-12683 of the *D. yakuba* mtDNA (Clary & Wolstenholme, 1985), was amplified using primers N1-J-12261 and LR-N-12945 (N116S). The specimens sequenced for each of the mtDNA regions are listed in Appendix 1. The PCR reactions were performed in a MJ Research PTC-100 thermal cycler. The optimized temperature profile for the COI fragment consisted of 35 cycles of 30 s denaturation at 95°C, 1 min annealing at 40°C, a further 30 s annealing at 50°C and 2 min extension at 72°C. For the 16S rRNA–tRNA^{Ala}(CUN)–ND1 fragment the temperature profile consisted of 35 cycles of 30 s denaturation at 95°C, 30 s annealing at 47°C and 90 s extension at 72°C. In both cases there was an initial denaturation step of 2 min at 95°C. The PCR products were checked for size and lack of multiple bands by agarose gel electrophoresis and purified with the Wizard PCR Preps DNA Purification System (Promega) according to the manufacturer's protocols. Sequences were determined directly, in both the 5' and 3' directions, using the same primers as above. Sequencing was either manual, using the Sequenase Version 2.0 DNA Sequencing Kit (United States Biochemical/ Amersham International plc.) and 35S dATP (Du Pont NEN) with autoradiographic visualization, or automated, using the ABI PRISM Dye Terminator Cycle Sequencing Ready Reaction Kit with AmpliTaq DNA Polymerase, FS (Perkin Elmer) with separation on an ABI PRISM 377 DNA Sequencer (Perkin Elmer). Sequences were aligned using Sequencher (Gene Codes Corporation Inc., Ann Arbor, Michigan), CLUSTAL X version 1.83 (Thompson *et al.*, 1997) and DNAMAN version 6 (Lynnon Corporation, Quebec). Comparison with the *D. yakuba* mtDNA from the GenBank suggested that the sequence fragments were from functional COI and 16S rRNA genes and no missense mutations or stop codons were detected. All sequences will be available at GenBank after publication.

2.4 Bioacoustic analyses

All acoustic studies were made at the National Institute of Biology (NIB) in Ljubljana (Slovenia). For song recording the animals were placed in the laboratory on small branches of

their fresh host plants. A small reflector was glued on the stem and the vibratory signals were recorded with a commercial laser vibrometer (Polytec system with the OFV-303 sensor head, OFV-300 modular vibrometer controller, OVD-02 velocity and OVD-20 displacement decoders; Polytech GmbH, Waldbrün, Germany) and stored directly on a notebook with an appropriate soundcard. Simultaneously the behaviour was observed. Further analyses of the songs have been made using the software Cool Edit Pro version 2.0 (Adobe Systems Inc., San Jose, CA) and GoldWave version 5.04 (GoldWave Inc., St. John's, Canada).

2.5 Phylogenetic analyses

For the morphological cladistic analysis a data matrix was produced (Table 2) with 19 species of *Kybos* and one species each of the outgroups *Kyboasca* and *Empoasca* respectively. Sixteen characters were chosen (Table 1). Character states were marked as dashes (–) if inapplicable and as question marks (?) if ambiguous or missing (Table 2). Three characters relate to the wings, seven to the male genitalia, five to the female genitalia and one to the nymphs. The morphological cladistic analysis was performed with NoNa (Goloboff, 1999) using the interface WinClada version 1.00.08 (Nixon, 2002). Multistate characters were treated as unordered (= non additive), all characters were assigned equal weights. A heuristic search was performed with following settings: maximum trees to keep = 10000; number of replications = 5; starting trees per replication = 5; search strategy = multiple tbr + tbr. The Nelsen command was used to calculate a consensus tree. For mapping the characters onto the consensus cladogram (Fig. 18), the fast character optimization was used.

PAUP*4.10 (Swofford, 1998) was used to perform the molecular cladistic analyses. *Philaenus spumarius* (Cercopidae) was added as an outgroup (GB accession number AY630340). In order to investigate genetic divergences and possible saturation effects that could interfere with later phylogenetic analysis, an initial neighbor joining analysis using uncorrected (p) and corrected (K2P) genetic distances, which corrects for differences in transition and transversion ratios, was performed. The commonly used Kimura two-parameter model of substitution (K2P) was employed on the basis that more sophisticated models do not necessarily result in better phylogenetic inference (Nei & Kumar, 2000). Branch support was assessed by 1000 nonparametric bootstrap replicates (Felsenstein, 1985).

3 Results

3.1 Taxonomy

3.1.1 Morphological character assesment

Coloration. Adult: Face usually greenish, sometimes ranging to yellowish as in *K. abstrusus* and *K. populi*, or rarely fuscous as in *K. sordidulus*. Pronotum and scutellum greenish or brownish, bearing often a few white spots or a white longitudinal band in the middle (e.g. *K. lindbergi*, *K. populi*, *K. smaragdula* or *K. virgator*). Fore wing generally greenish; commisural border sometimes concolorous with remainder of wing (e.g. *K. populi*) or with a brown band (e.g. *K. rufescens*, *K. butleri*); colour of corioclaval suture either concolorous with remainder of wing (e.g. *K. populi*) or bearing a black spot or fuscous (e.g. *K. smaragdula*). Abdomen greenish, in some species with darker and stronger pigmented tergites. Teneral specimens pale and weakly pigmented. With a few exceptions (*K. rufescens*, *K. sordidulus*) the coloration does not diagnose *Kybos* species. – Nymph: Body colour varying from green, with hardly any markings, to brownish. Posterior margins of abdominal tergites often brown.

Sexual characters. Male (Fig. 1): The aedeagus is, by far, the most important structure for diagnosing *Kybos* species. Appendages absent (e.g. *K. rufescens*, *K. populi*) or present (e.g. *K. lindbergi*, *K. virgator*); where present, divergent from (e.g. *K. virgator*) or parallel to the main stem (e.g. *K. lindbergi*); length varying from shorter than (e.g. *K. lindbergi*) to as long as main stem (e.g. *K. austriacus*); base of appendages broad and widely separated in the middle (e.g. *K. lindbergi*) or almost contiguous (e.g. *K. virgator*). There are some species where the appendages of some individuals may bear irregular spines or teeth (prominent e.g. in *K. strigilifer*); this character is very variable and not diagnostic. Pygofer process apically slender in all Central European species except for *K. populi* where it is broadened. Processes of anal collar either short and stout (e.g. *K. lindbergi*, *K. smaragdula*) or long and slender (e.g. *K. rufescens*, *K. virgator*). A further taxonomically important character is the shape of the male singing organs, the apodemes. This structure is only fully developed after a few days of hatching and caution is required with teneral specimens where the apodemes are usually shorter. Length of apodemes of sternite II varying from shorter than broad to twice as long as broad; length of phragma lobes of tergite III varying from shorter than broad to longer than broad. – Female: Sternite VII in all European species bearing prolonged lobe, except for *K. populi* where it is equally rounded. Three types of shape of the ovipositor base can be distinguished: 1. wider than long (e.g. *K. smaragdula*), 2. as long as broad as in *K. lindbergi*

or 3. longer than wide as in *K. populi*; distal edge straight (e.g. *K. mucronatus*), equally rounded as in *K. smaragdula* or with an incision (e.g. *K. populi*).

3.1.2 Morphological species concept

As shown in the character assessment the aedeagus is the single most important structure for diagnosing species. The second good diagnoser is the shape of the sound apodemes, teneral specimens excluded.

Kybos virgator (Figs. 7D-E) and *volgensis* (Figs. 7F-G) are morphologically similar and differ only in the shape of the apodemes. The two taxa are sympatric and are both associated with willows. In the surroundings of Basel (Switzerland) and Moravia (Czech Republic) individuals of *K. volgensis* were observed only during a few days in June and August respectively, whereas those of *K. virgator* were registered during a few weeks. A few specimens with intermediate apodeme shape were collected. These observations are more consistent with *K. volgensis* representing a juvenile apodeme form of *K. virgator* than the occurrence of two biologically separated species. *K. volgensis* is, therefore, synonymised here with *K. virgator*.

K. perplexus (Figs. 3C-D) was separated from *strigilifer* (Figs. 3A-B) by the aedeagal appendages bearing spines. Günthart (1974) showed with breeding experiments that the presence of spines is highly variable within *strigilifer*. Dworakowska (1976), interpreting Günthart's data slightly differently, concluded that the two are separate species. In the material at hand from several Central and Northern European countries (Appendix 1) we observed a significant variability in this structure. The two forms share the same distribution and occur together. In the surroundings of Basel (Switzerland) the two were collected together on the same tree. These observations are in accord with Günthart's conclusions can be best interpreted as intraspecific variation with respect to aedeagal appendages. For this reason *K. strigilifer* and *perplexus* are here synonymised.

A third type of taxonomic problem is illustrated by *K. lindbergi* and *K. ludus* (Figs. 7S-U), a species pair associated with birch. They are morphologically similar but differ subtly in the shape of the aedeagal appendages. *K. ludus* occurs in Western Europe, *K. lindbergi* in Northern and Eastern Europe. In Germany and Switzerland the two species are sympatric and intermediate morphological forms are frequent. Based on the available evidence we conclude that *K. lindbergi* and *K. ludus* are distinct, geographically separated species which are sympatric in a relatively restricted area in Central Europe, where hybridisation occurs. The name *K. ludus* is used here according to Hamilton (1983) who suggested that this is a senior

synonym of *Empoasca* (*Kybos*) *betulicola*. This was followed by della Giustina (1989). Based on the original description and illustration of *K. ludus* by Davidson & DeLong (1938) Nickel (2003) and other European workers rejected this synonymy. We have examined the holotype of *K. ludus* which clearly shows that it is conspecific with *K. betulicola*. We agree with Hamilton and reject the use of the name *K. betulicola*.

3.1.3 Species descriptions

***Kybos* Fieber**

Kybos Fieber, 1866: 508; type species *Cicada smaragdula* Fallén, 1806, by monotypy; Nast, 1972; Le Quesne, 1981.

Empoasca Walsh, 1862: 149, p.p.; Horváth, 1897, with *Kybos* as junior synonym.

Empoasca (*Kybos*) DeLong, 1931: 14; Metcalf, 1968; Dworakowska, 1976; Ossiannilsson, 1981.

Host plant range. Betulaceae (*Alnus*, *Betula*) and Salicaceae (*Populus*, *Salix*).

Distribution. Holarctic; Eurasia 45 species, North America 78 species.

***Kybos abstrusus* (Linnavuori) (Figs. 2A-B, 5A, 6A, 8A)**

Empoasca abstrusa Linnavuori, 1949: 148.

Kybos topoli Zachvatkin, 1953a: 209; synonymised by Dworakowska, 1976.

Body length. Male 4.2-4.7 mm; female 4.5-4.9 mm.

Coloration. Adult: Face greenish or yellowish; commisural border concolorous with remainder of fore wing; corioclaval suture concolorous with forewing.

Sexual characters. Male: Aedeagus without processes. Pygofer process slender apically. Process of anal collar long and slender. Apodemes of sternite II twice as long as broad; phragma lobes of tergite 3 shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view narrow and angular, without a median ridge, hook-shaped distally.

Host plant. *Populus nigra*.

Distribution. Austria, Bulgaria, Finland, France (mainland), Germany, Hungary, Lithuania, Poland, Romania, central part of European Russia, Sweden, Switzerland, Ukraine, former Yugoslavia.

***Kybos aetnicola* Wagner, 1959 (Figs. 2K, 4A, 5B, 6B)**

Kybos aetnicola Wagner, 1959: 81.

Body length. Male: 4.4-4.5 mm; female: 4.5-4.8 mm; nymph: 2.8 mm.

Coloration. Adult: Face greenish; commisural border concolorous with fore wing; corioclaval suture black or fuscous. – Nymph: Green, almost with hardly any markings, posterior margins of abdominal tergites brown.

Sexual characters. Male: Aedeagus with divergent processes, in lateral view, ventral to main stem; length of processes as long as stem; basis of processes close to each other. Pygofer process in male slender apically. Process of anal collar short and stout. – Female: Sternite VII with prolonged lobe.

Host plant. *Betula aetnensis*.

Distribution. Italy (Mt. Etna, Sicily).

***Kybos austriacus* (Wagner, 1949)** (Figs. 2L-M, 8F)

Empoasca austriaca Wagner, 1949: 43.

Body length. Male: 4.2-4.5 mm; female: 4.7-4.8 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture black or fuscous.

Sexual characters. Male: Aedeagus with parallel sited processes, in lateral view parallel to main stem; length of processes as long as stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar short and stout. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded, with a median ridge, distally straight.

Host plant. *Betula* spp.

Distribution. Austria, Switzerland.

***Kybos butleri* (Edwards, 1908)** (Figs. 2C, 4B, 5C, 6C, 8B, 11B)

Empoasca butleri Edwards, 1908: 82.

Kybos oshanini occidentalis Zachvatkin, 1953a: 208; synonymised by Dworakowska, 1976.

Empoasca ovalis Ross, 1963: 217; synonymised by Dworakowska, 1976.

Body length. Male 3.9-4.3 mm; female 4.2-4.7 mm; nymph 3.5 mm.

Coloration. Adult: Face greenish; commisural border of fore wing with a brown band; corioclaval suture concolorous with forewing. – Nymph: brown pattern on pro-, meso- and metanotum, tergite 2 with two dark spots, posterior margins of abdominal tergites brown.

Sexual characters. Male: Aedeagus without processes. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad; phragma lobes of tergite III as long as broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view narrow and angular; without a median ridge; distally hook-shaped; apex of median valvula bearing 6-10 teeth; structure of teeth regular triangular; ventral edge of valvula apically evenly curved.

Nymph. Length of tibia 3 distinctly shorter than half of body length (<0.4).

Host plant. *Salix aurita*, *S. cinerea*, *S. repens*, *S. triandra*, *S. caprea*, *S. myrsinifolia*.

Distribution. Belgium, Denmark (mainland), Estonia, Finland, France (mainland), Germany, Great Britain, Greece (mainland), Hungary, Italy (mainland), Latvia, Lithuania, Netherlands, Poland, North part of European Russia, Slovakia, Slovenia, Sweden, Switzerland, Ukraine, former Yugoslavia.

***Kybos calyculus* (Cerutti, 1939)** (Figs. 2N-O, 4C, 5D, 6D)

Empoasca calycula Cerutti, 1939: 92.

Kybos studzinskii Dworakowska, 1973: 239; synonymised by Dworakowska, 1976.

Body length. Male 4.1-4.3 mm; female 4.2-4.4 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture black or fuscous.

Sexual characters. Male: Aedeagus with divergent processes, in lateral view ventrally of main stem, length of processes shorter than stem, basis of processes close to each other.

Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III as long as broad or shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded; with a median ridge; distally straight.

Host plant. *Betula pubescens*.

Distribution. Czech Republic, Germany, Great Britain, Poland, Switzerland.

***Kybos candelabricus* Dlabola, 1958** (Figs 2P, 4D, 5E, 6E)

Kybos candelabricus Dlabola, 1958: 331.

Body length. Male: 4.7-5.0 mm; female: 4.8-5.1 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous.

Sexual characters. Male: Aedeagus with divergent processes, in lateral view parallel to main stem, length of processes as long as stem; basis of processes widely apart from each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe.

Host plant. *Salix* spp.

Distribution. Bulgaria, Georgia, Turkey (Anatolia).

***Kybos digitatus* (Ribaut, 1936)** (Figs. 2Q-R, 4E, 5F, 6F, 9D)

Empoasca digitata Ribaut, 1936: 178.

Body length. Male 3.7-3.9 mm; female 3.9-4.4 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous.

Sexual characters. Male: Aedeagus with parallel sited processes, in lateral view dorsally of main stem; length of processes shorter than stem; basis of processes widely apart from each other. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad; phragma lobes of tergite III as long as broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded; without a median ridge; distally rounded.

Host plant. *Salix elaeagnos*.

Distribution. France (mainland), Germany, Switzerland.

***Kybos limpidus* (Wagner, 1955)** (Figs. 2D-E, 4F, 5G, 6G, 8C, 11D)

Empoasca limpida Wagner, 1955: 176.

Body length. Male 4.2-4.4 mm; female 4.8-5.1 mm; nymph 2.9-3.2 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture concolorous with forewing. – Nymph: Light green, almost without any markings.

Sexual characters. Male: Aedeagus without processes. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view narrow and angular; without a median ridge; distally hook-shaped; apex of median valvula bearing 6-10 teeth; structure of teeth irregular coarse; ventral edge of valvula apically evenly curved.

Nymph. Length of tibia 3 distinctly shorter than half of body length (<0.4).

Host plant. *Salix viminalis*, *S. triandra*, possibly *S. pentandra*.

Distribution. Czech Republic, Germany, Hungary, Netherlands, Poland, Ukraine.

***Kybos lindbergi* (Linnavuori, 1951)** (Figs. 2T-U, 4G, 5H, 6H, 9A, 11C)

Empoasca lindbergi Linnavuori, 1951: 60.

Empoasca borealis Lindberg, 1952: 144; synonymised by Linnavuori, 1953.

Kybos betulae (sic!) Zachvatkin, 1953a: 206; synonymised by Nast, 1972

Empoasca austriaca Ossiannilsson, 1955: 131 (nec Wagner, 1949).

Body length. Male 4.2-4.4 mm; female 4.6-4.8 mm; nymph 2.9 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture black or fuscous. – Nymph: Light green, almost without any markings, only posterior margins of abdominal tergites brown.

Sexual characters. Male: Aedeagus with parallel sited processes, in lateral view parallel to main stem; length of processes shorter than stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad or as long as broad; phragma lobes of tergite III as long as broad or shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded with a median ridge, distally straight; apex of median valvula bearing 6-10 teeth; structure of teeth irregular coarse; ventral edge of valvula apically strongly curved.

Nymph. Length of tibia 3 almost half of body length.

Host plant. *Betula pendula*, *B. pubescens*, *B. nana*.

Distribution. Czech Republic, Estonia, Finland, Germany, Kazakhstan, Latvia, Norway (mainland), Poland, central part of European Russia, Sweden, Switzerland, Ukraine.

***Kybos ludus* (Davidson & DeLong, 1938)** (Figure 2S)

Empoasca luda Davidson & DeLong, 1938: 94.

Empoasca betulicola Wagner, 1955: 178; synonymised by Hamilton, 1983.

Body length. Male 4.2-4.4 mm; female 4.6-4.8 mm; nymph 2.9 mm.

Coloration. Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture black or fuscous. – Nymph: Green, almost without any markings, only posterior margins of abdominal tergites brown.

Sexual characters. Male: Aedeagus with parallel sited processes, in lateral view parallel to main stem, length of processes shorter than stem, basis of processes widely apart from each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad, or as long as broad, phragma lobes of tergite III as long as broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded, with a median ridge, distally straight; apex of median valvula bearing 6-10 teeth, structure of teeth irregular coarse, ventral edge of valvula apically strongly curved.

Nymph. Length of tibia 3 almost half of body length.

Host plant. *Betula pendula*, *B. pubescens*.

Distribution. Germany, Great Britain, Mongolia, Netherlands, south part of European Russia, Switzerland.

Kybos mesasiaticus* (Zachvatkin, 1953)** (Figs. 2F, 4H, 5J, 6J, 11E)*Kybos oshanini mesasiaticus* Zachvatkin, 1953a: 208.*Kybos mesasiaticus* Zachvatkin, 1953b: 237.*Body length.* Male: 3.8-4.2 mm; female: 3.8-4.2 mm*Coloration.* Adult: Face greenish; commissural border of fore wing concolorous; corioclaval suture concolorous with forewing.*Sexual characters.* Male: Aedeagus without processes. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad; phragma lobes of tergite III longer than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view narrow and angular, without a median ridge, distally hook-shaped; apex of median valvula bearing 6-10 teeth; structure of teeth irregular coarse; ventral edge of valvula apically evenly curved.*Host plant.* *Salix songorica*.*Distribution.* Dagestan, Mongolia, Kazakhstan, Kirghizia, south part of European Russia, Uzbekistan.Kybos mucronatus* (Ribaut, 1933)** (Figs. 2V-W, 4K, 5K, 6K, 10A)*Empoasca mucronata* Ribaut, 1933: 151.*Kybos mucronatus verbae* Zachvatkin, 1953a: 207.*Kybos cracoviensis* Dworakowska, 1973: 239; synonymised by Dworakowska, 1976.*Body length.* Male 4.6-4.7 mm; female 5.0 mm.*Coloration.* Adult: Face greenish; commissural border of fore wing concolorous.*Sexual characters.* Male: Aedeagus with divergent processes, in lateral view ventrally of main stem; length of processes shorter than stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad or as long as broad, phragma lobes of tergite III as long as broad or shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded with a median ridge; distally straight.*Host plant.* *Alnus glutinosa*, reported also from *A. incana* and *Salix* spp.*Distribution.* Czech Republic, France (mainland), Germany, Latvia, Poland, central part of European Russia, Slovenia, Switzerland, former Yugoslavia.*Comment.* Nickel (2003) discussed the synonymy of *verbae* with *mucronatus*. Here we follow Nickel.***Kybos paraltaicus* Orosz, 1996***Empoasca paraltaica* Orosz, 1996: 153.*Body length.* Male 4.3 mm.*Coloration.* Adult: Face greenish; commissural border of fore wing concolorous.*Sexual characters.* Male: Aedeagus with divergent processes, length of processes shorter than stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad; phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe.*Host plant.* Unknown.*Distribution.* Hungary.

Comment. *K. paraltaicus* is known from a single male from Hungary only. Additional material is necessary for elucidating its identity.

***Kybos populi* (Edwards, 1908)** (Figs. 2G, 4K, 5L, 6L, 8D, 11F)

Empoasca populi Edwards, 1908: 81.

Kybos populi tremulae Zachvatkin, 1953a: 208; synonymised by Dworakowska, 1976.

Kybos zaisanensis Mitjaev, 1968: 635; synonymised by Dworakowska, 1976.

Body length. Male 3.8-4.5 mm; female 4.0-4.7 mm; nymph 3.2.

Coloration. Adult: Face greenish, or yellowish; commisural border of fore wing concolorous; corioclaval suture concolorous with forewing. – Nymph: Light green, almost without any markings.

Sexual characters. Male: Aedeagus without processes. Pygofer process in male apically broadend. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III shorter than broad. – Female: Sternite VII without prolonged lobe, equally rounded; base of first valvifer in ventral view narrow and angular without a median ridge, distally straight; apex of median valvula bearing 6-10 teeth; structure of teeth irregular coarse; ventral edge of valvula apically evenly curved.

Nymph. Length of tibia III distinctly shorter than half of body length (<0.4).

Host plant. *Populus alba*, *P. nigra*, *P. suaveolens*, *P. tremula*.

Distribution. Austria, Belgium, Bulgaria, Czech Republic, Danish mainland, Estonia, Finland, France (mainland), Germany, Great Britain, Hungary, Italy (mainland and Sicily), Kazakhstan, Latvia, Republic of Moldova, Morocco, Netherlands, Norway (mainland), Poland, south part of European Russia, central part of Siberia, Slovakia, Slovenia, Sweden, Switzerland, Ukraine, former Yugoslavia.

***Kybos rufescens* Melichar, 1896** (Figs. 2H, 6M, 8E, 11G, 13B)

Kybos smaragdula (sic!) *rufescens* Melichar, 1896: 180.

Body length. Male 3.9-4.5 mm; female 4.1-4.6 mm.

Coloration. Adult: Face greenish; commisural border of fore wing brown band; corioclaval suture concolorous with forewing.

Sexual characters. Male: Aedeagus without processes. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view narrow and angular without a median ridge, distally hook-shaped; apex of median valvula bearing 6-10 teeth, structure of teeth irregular coarse; ventral edge of valvula apically evenly curved.

Host plant. *Salix purpurea*, possibly *S. caesia*.

Distribution. Austria, Belgium, Bulgaria, Czech Republic, Danish mainland, French mainland, Germany, Great Britain, Hungary, Italy (mainland and Sicily), . Kazakhstan, Netherlands, Mongolia, Poland, Romania, south part of European Russia, Sardinia, Slovakia, Slovenia, Sweden, Switzerland, Ukraine, former Yugoslavia.

Kybos smaragdula* (Fallén, 1806)** (Figs. 3H, 4L, 5M, 7A, 9B, 10B, 11H, 13E)*Cicada smaragdula* Fallén, 1806: 37.*Empoasca smaragdula*; Horváth, 1897: 45.*Kybos smaragdulus*; Fieber, 1866: 508.*Body length.* Male 3.8-4.3 mm; female 4.1-4.6 mm; nymph 3.6 mm.*Coloration.* Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture black or fuscous. – Nymph: Light green, almost without any markings, only posterior margins of abdominal tergites brown.*Sexual characters.* Male: Aedeagus with parallel sited processes, in lateral view parallel to main stem or dorsally of main stem; length of processes shorter than stem; basis of processes widely apart from each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II shorter than broad; phragma lobes of tergite III as long as broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded with a median ridge, distally straight; apex of median valvula bearing 6-10 teeth; structure of teeth regular triangular; ventral edge of valvula apically strongly curved.*Nymph.* Length of tibia 3 almost half of body length.*Host plant.* *Alnus glutinosa*, *A. incana*, possibly *A. alnobetula*.*Distribution.* Altai Mts., Armenia, Austria, Belgium, Bulgaria, Croatia, Czech Republic, Denmark (mainland), Estonia, Finland, France (mainland), Germany, Great Britain, Hungary, Ireland, Italy (mainland), Kazakhstan, Latvia, Lithuania, Netherlands, Norway (mainland), Poland, Portugal (mainland), Romania, central and north parts of European Russia, Sakhalin, Siberia, Slovakia, Slovenia, Spain (mainland), Sweden, Switzerland, Tadzhikistan, Ukraine, Uzbekistan, former Yugoslavia.Kybos sordidulus* (Ossiannilsson, 1941)** (Figs. 2J, 4M, 5N, 7B)*Empoasca sordidula* Ossiannilsson, 1941: 69 (male); Ossiannilsson, 1942: 114 (female).*Empoasca* (*Kybos*) *alaskana* Ross, 1963: 219; synonymised by Dworakowska, 1976.*Body length.* Male 4.0-4.7 mm; female 4.0-4.7 mm.*Coloration.* Adult: Face fuscous; commisural border of fore wing concolorous; corioclaval suture concolorous with forewing.*Sexual characters.* Male: Aedeagus without processes. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III longer than broad. – Female: Sternite VII with prolonged lobe.*Host plant.* *Salix caprea*, *S. myrsinifolia*, *S. purpurea*, *S. phylicifolia*.*Distribution.* Finland, Norway (mainland), central part of European Russia, Sweden.***Kybos strigilifer* (Ossiannilsson, 1941)** (Figs. 1A-F, 3A-D, 4N, 5O, 7C, 9E, 12A, 13C)*Empoasca strigilifera* Ossiannilsson, 1941: 198.*Empoasca perplexa* Ribaut, 1952: 459; syn. n.*Kybos altaicus* Mitjaev, 1963: 66; synonymised with *Empoasca perplexa* by Dworakowska, 1976.*Body length.* Male 3.9-4.3 mm; female 4.2-4.5 mm.*Coloration.* Adult: Face greenish; commisural border of fore wing concolorous; corioclaval suture black or fuscous.

Sexual characters. Male: Aedeagus with divergent processes, in lateral view parallel to main stem; length of processes shorter than stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded with a median ridge, distally rounded; apex of median valvula bearing 6-10 teeth; structure of teeth irregular coarse; ventral edge of valvula apically strongly curved.

Host plant. *Salix caprea*, *S. cinerea*, *S. myrsinifolia*.

Distribution. Austria, Belgium, Czech Republic, Danish mainland, Estonia, Finland, France (mainland), Germany, Great Britain, Ireland, Latvia, Lithuania, Norway (mainland), Poland, central and northern part of European Russia, Slovakia, Slovenia, Sweden, Switzerland, Ukraine, former Yugoslavia.

Comment. *K. perplexus* (Figs. 3C-D) is a younger synonym of *K. strigilifer*. The two species differ only in details of the aedeagal appendages. *K. perplexus* is reported from Belgium, France (mainland), Germany, Poland, Switzerland and Ukraine. *K. strigilifer* is not recorded for Belgium.

***Kybos strobli* (Wagner, 1949)** (Figs. 3E-G, 4O, 9C, 12B)

Empoasca strobli Wagner, 1949: 44.

Body length. Male 4.0-4.6 mm; female 4.0-4.6 mm.

Coloration. Adult: Face greenish; commissural border of fore wing concolorous; corioclaval suture black or fuscous.

Sexual characters. Male: Aedeagus with divergent processes, in lateral view dorsally of main stem; length of processes shorter than stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar short and stout. Length of apodemes of sternite II shorter than broad; phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded with a median ridge, distally straight; apex of median valvula bearing 6-10 teeth; structure of teeth regular triangular; ventral edge of valvula apically strongly curved.

Host plant. *A. incana*, possibly on other *Alnus* spp.

Distribution. Austria, Czech Republic, Germany, Hungary, Italy (mainland), Poland, Slovenia, Switzerland, Ukraine.

***Kybos virgator* (Ribaut, 1933)** (Figs. 3J-O, 4P, 5P, 7D-G, 9F, 12C, 13F)

Empoasca virgator Ribaut, 1933: 152.

Empoasca virgator v. *saageri* Wagner, 1935: 28.

Kybos volgensis Vilbaste, 1961: 318; syn. n.

Body length. Male 4.0-4.5 mm; female 4.2-4.8 mm.

Coloration. Adult: Face greenish; commissural border of fore wing concolorous; corioclaval suture black or fuscous.

Sexual characters. Male: Aedeagus with divergent processes, in lateral view parallel to main stem; length of processes shorter than stem; basis of processes close to each other. Pygofer process in male apically slender. Process of anal collar long and slender. Length of apodemes of sternite II twice as long as broad, phragma lobes of tergite III shorter than broad. – Female: Sternite VII with prolonged lobe; base of first valvifer in ventral view broad and irregularly rounded with a median ridge; distally rounded; apex of median valvula bearing 6-10 teeth; structure of teeth irregular coarse; ventral edge of valvula apically evenly curved.

Host plant. *Salix* spp.; in Central Europe mainly on *Salix alba* and *S. fragilis*.

Distribution. Austria, Belgium, Bulgaria, Czech Republic, Danish mainland, Finland, France (mainland), Germany, Great Britain, Greece (mainland), Hungary, Italy (mainland), Kazakhstan, Latvia, Lithuania, Republic of Moldova, Mongolia, Netherlands, Norway (mainland), Poland, Romania, central part of European Russia, Sardinia, Slovakia, Sweden, Switzerland, Ukraine, former Yugoslavia.

Comment: Here we synonymise *K. volgensis* and *K. virgator*. The differences of the length of the apodemes are a result of teneral specimens (Figs. 7F-G). *K. volgensis* is reported from the South part of European Russia, Sweden and Switzerland.

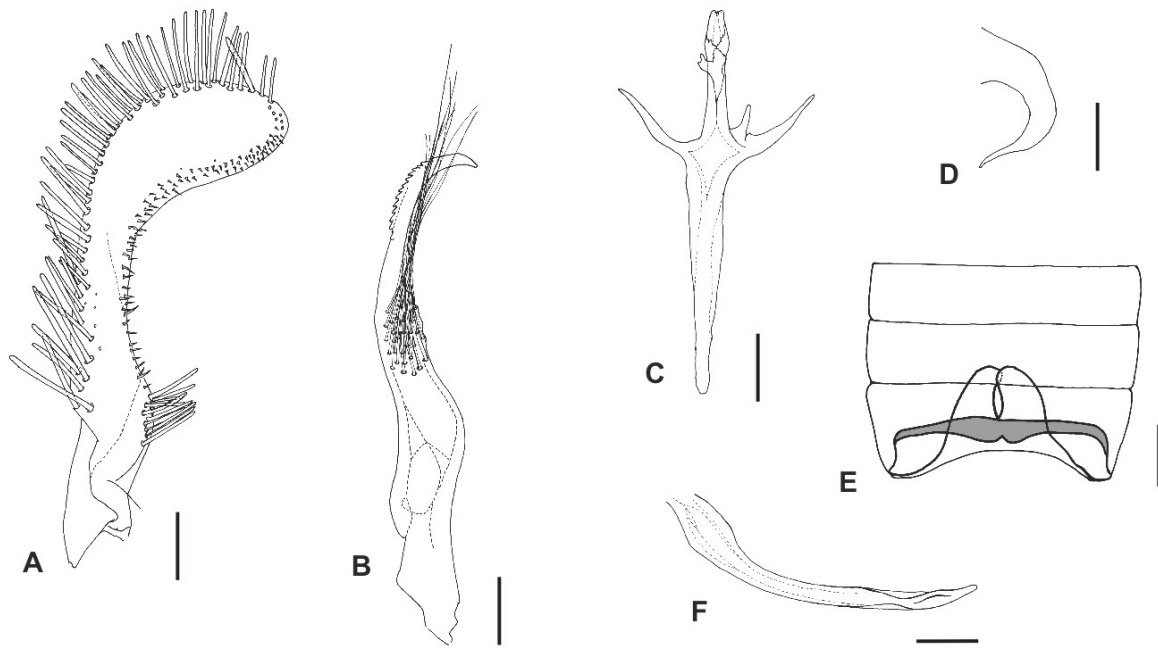


Figure 1. A-F, male genitalia of *Kybos strigilifer*. A, genital plate; B, stylus; C, aedeagus, ventral view; D, anal collar; E, apodemes (Aaodemes of tergite II in grey); F, pygofer. Scale bars: A-D, F = 0.1 mm; E = 0.2 mm.

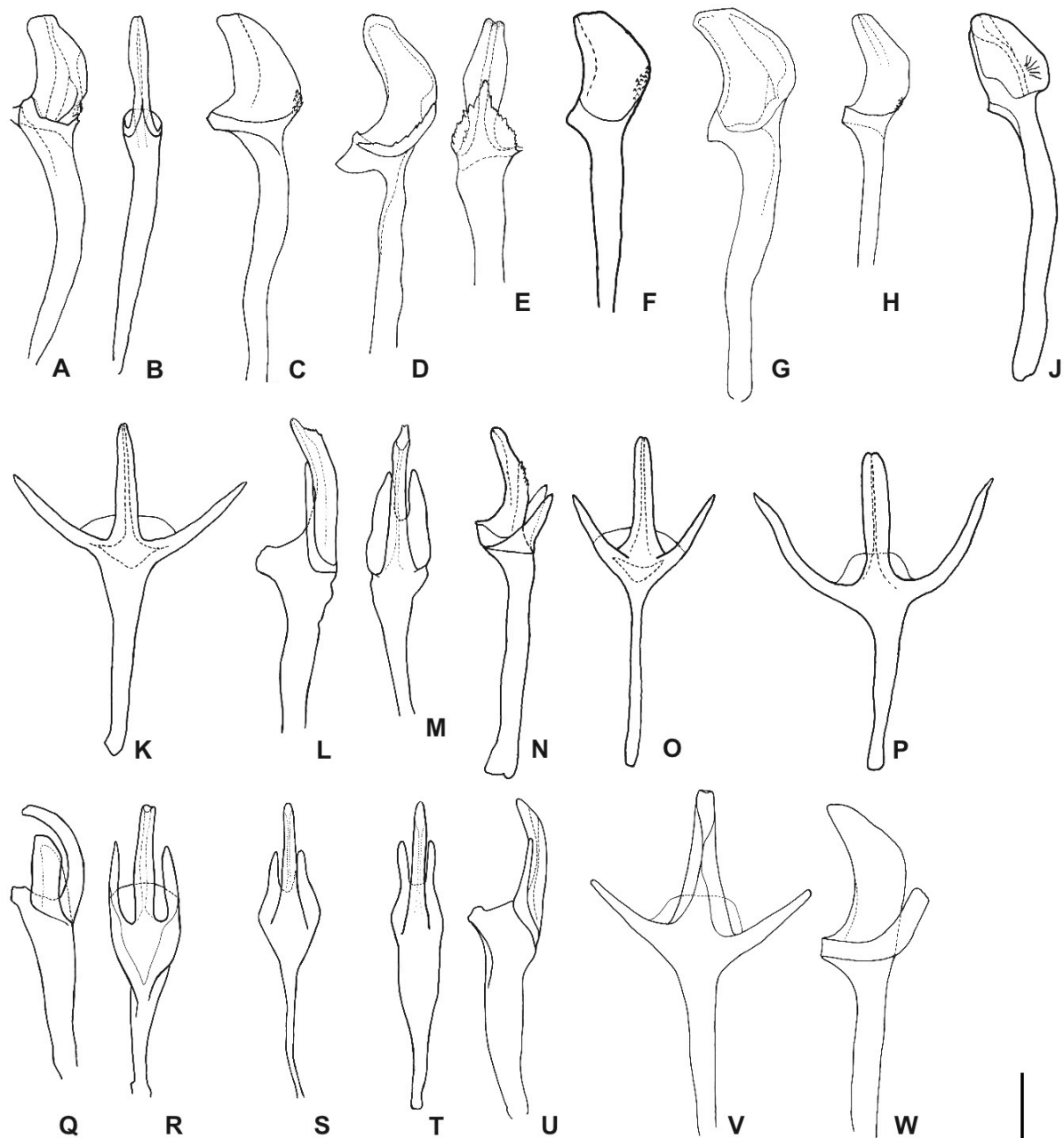


Figure 2. A-W, aedeagi of *Kybos* spp. A, *Kybos abstrusus*, lateral view; B, *K. abstrusus*, ventral view; C, *K. butleri*, lateral view; D, *K. limpidus*, lateral view; E, *K. limpidus*, ventral view; F, *K. mesasiaticus*, lateral view; G, *K. populi*, lateral view; H, *K. rufescens*, lateral view; J, *K. sordidulus*, lateral view; K, *K. aetnicola*, ventral view; L, *K. austriacus*, lateral view; M, *K. austriacus*, ventral view; N, *K. calyculus*, lateral view; O, *K. calyculus*, ventral view; P, *K. candelabricus*, ventral view; Q, *K. digitatus*, lateral view; R, *K. digitatus*, ventral view; S, *K. ludus*, ventral view; T, *K. lindbergi*, ventral view; U, *K. lindbergi*, lateral view; V, *K. mucronatus*, ventral view; W, *K. mucronatus*, lateral view. Scale bar = 0.1 mm.

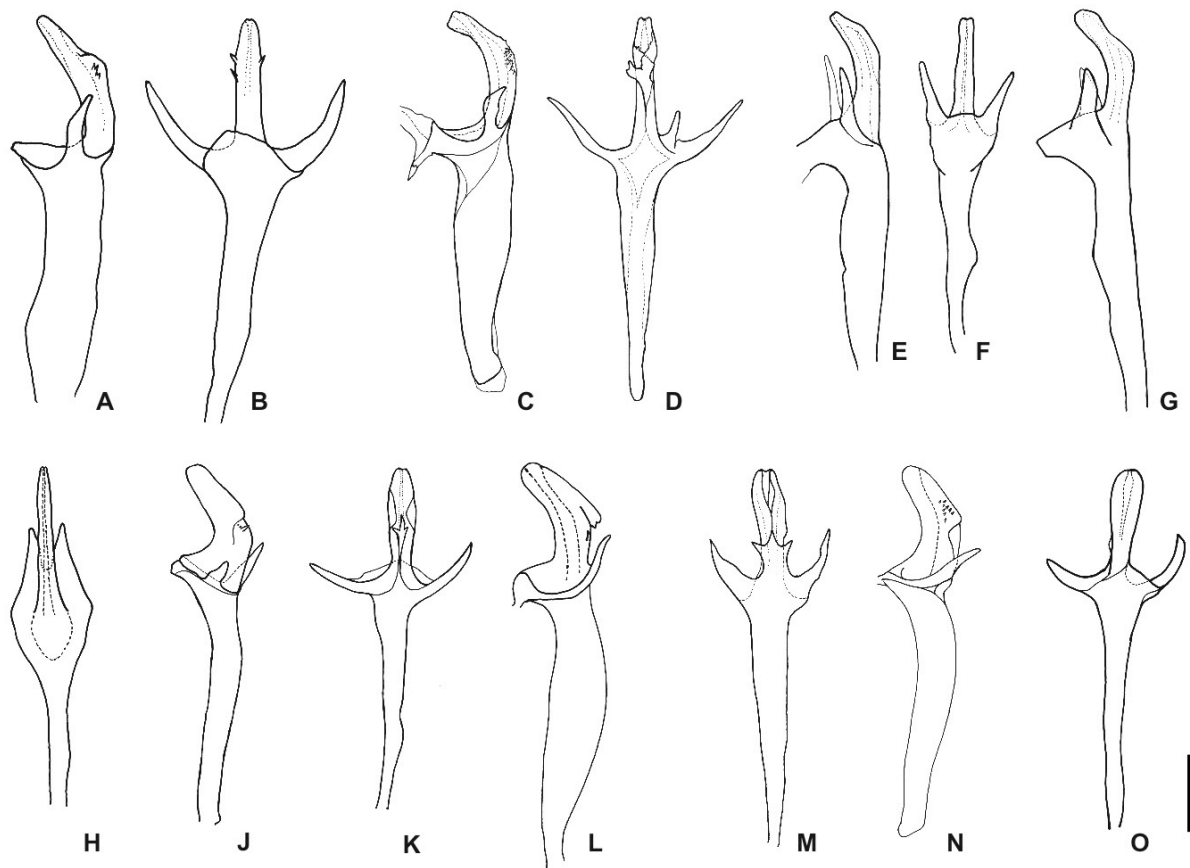


Figure 3. A-O, aedeagi of *Kybos* spp. A, *Kybos strigilifer*, lateral view; B, *K. strigilifer*, ventral view; C, *K. strigilifer* ("perplexus"), lateral view; D, *K. strigilifer* ("perplexus"), ventral view; E, *K. strobli*, lateral view; F, *K. strobli*, ventral view; G, *K. strobli*, lateral view; H, *K. smaragdulus*, ventral view; J, *K. virgator*, lateral view; K, *K. virgator*, ventral view; L, *K. virgator*, specimen from Slovakia, lateral view; M, *K. virgator*, specimen from Slovakia with some aberrations, ventral view; N, *K. virgator*, same specimens as in M, lateral view; O, *K. virgator* ("volgensis"). Scale bar = 0.1 mm.

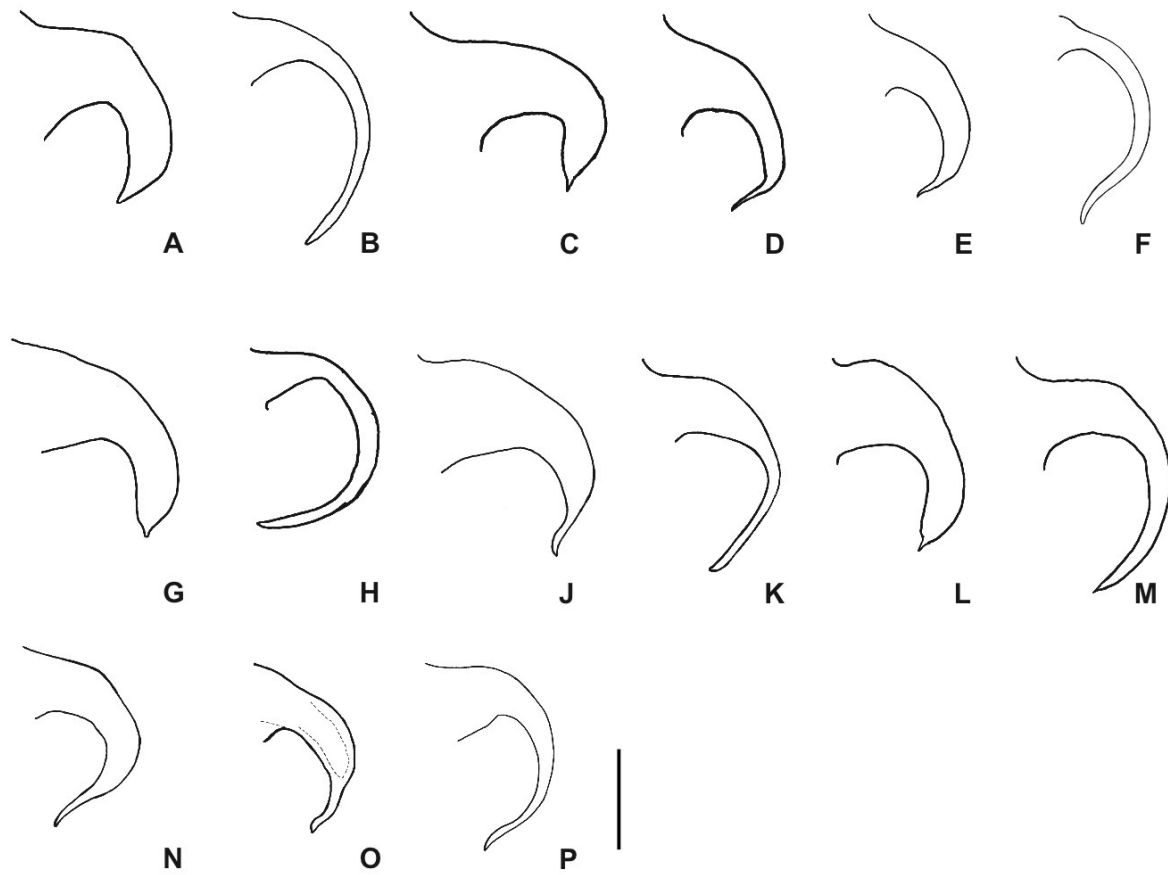


Figure 4. A-P, anal collar of *Kybos* spp. A, *Kybos aetnicola*; B, *K. butleri*; C, *K. calyculus*; D, *K. candelabricus*; E, *K. digitatus*; F, *K. limpidus*; G, *K. lindbergi*; H, *K. mesasiaticus*; K, *K. mucronatus*; K, *K. populi*; L, *K. smaragdulus*; M, *K. sordidulus*; N, *K. strigilifer*; O, *K. strobli*; P, *K. virgator*. Scale bar = 0.1 mm.

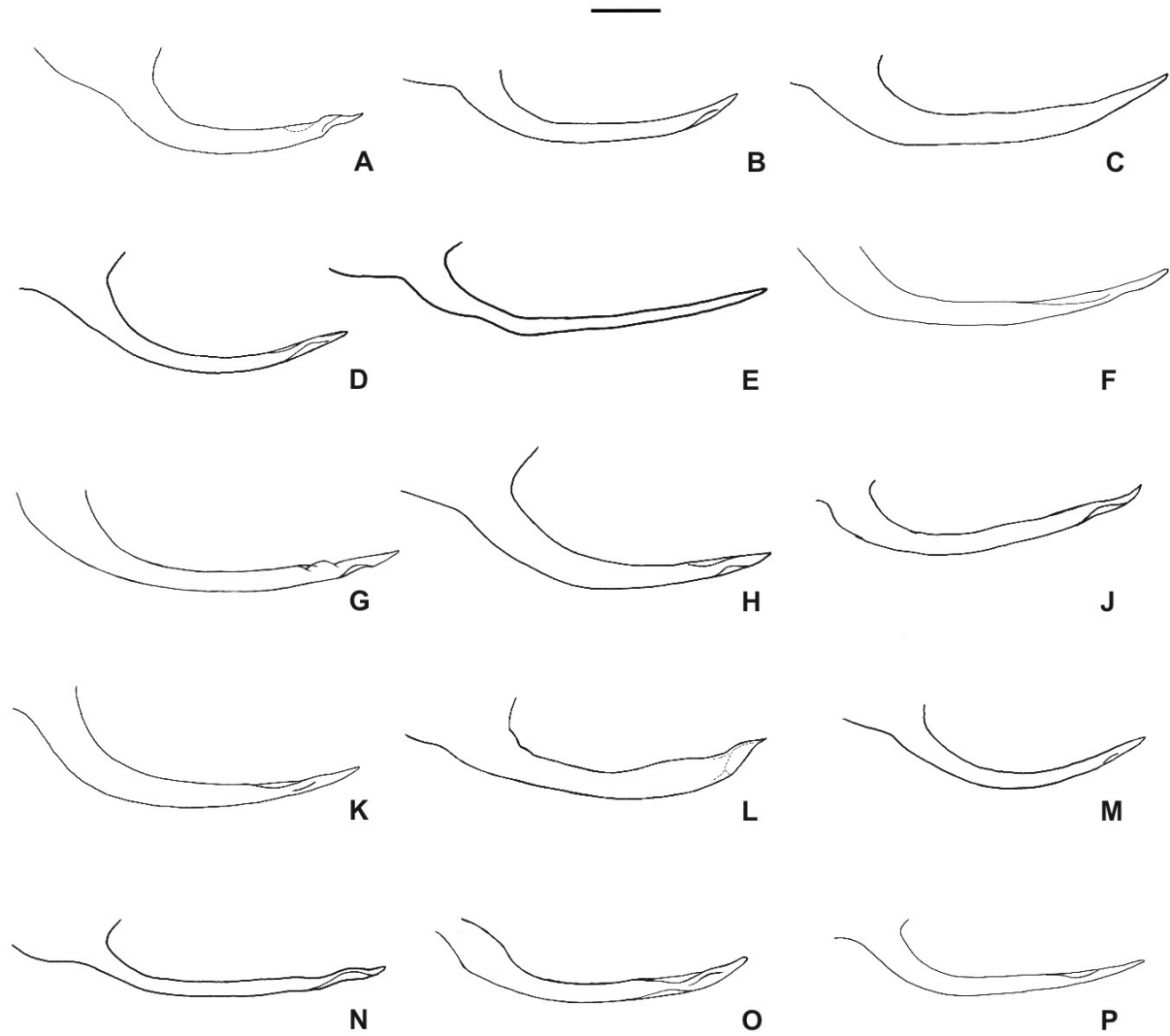


Figure 5. A-P, pygofer of *Kybos* spp. A, *Kybos abstrusus*; B, *K. aetnicola*; C, *K. butleri*; D, *K. calyculus*; E, *K. candelabricus*; F, *K. digitatus*; G, *K. limpidus*; H, *K. lindbergi*; J, *K. mesasiaticus*; K, *K. mucronatus*; L, *K. populi*; M, *K. smaragdulus*; N, *K. sordidulus*; O, *K. strigilifer*; P, *K. virgator*. Scale bar = 0.1 mm.

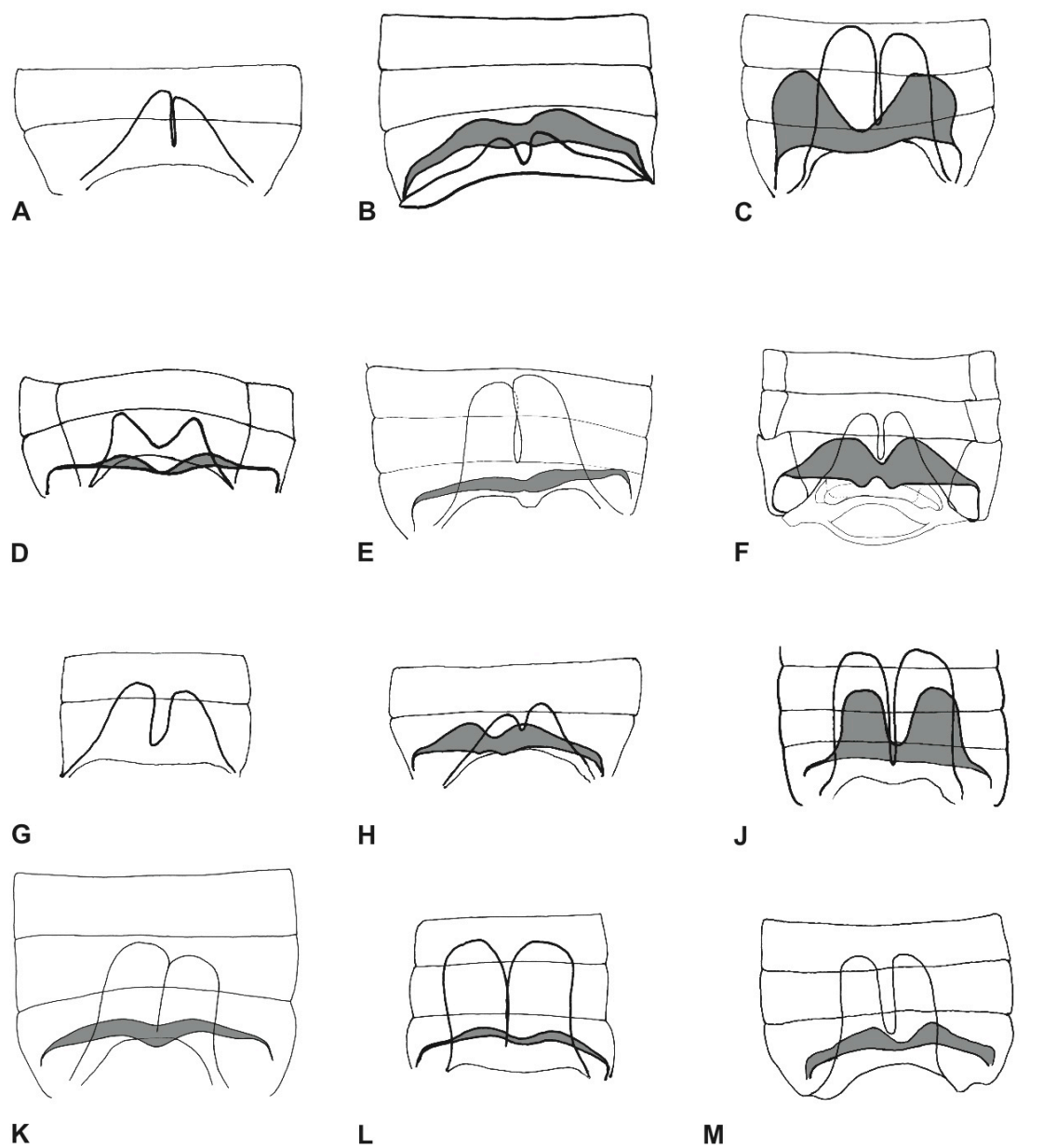


Figure 6. A-M, apodemes of *Kybos* spp. A, *Kybos abstrusus*; B, *K. aetnicola*; C, *K. butleri*; D, *K. calyculus*; E, *K. candelabricus*; F, *K. digitatus*; G, *K. limpidus*; H, *K. lindbergi*; J, *K. mesasiaticus*; K, *K. mucronatus*; L, *K. populi*; M, *K. rufescens*. Scale bar = 0.2 mm.

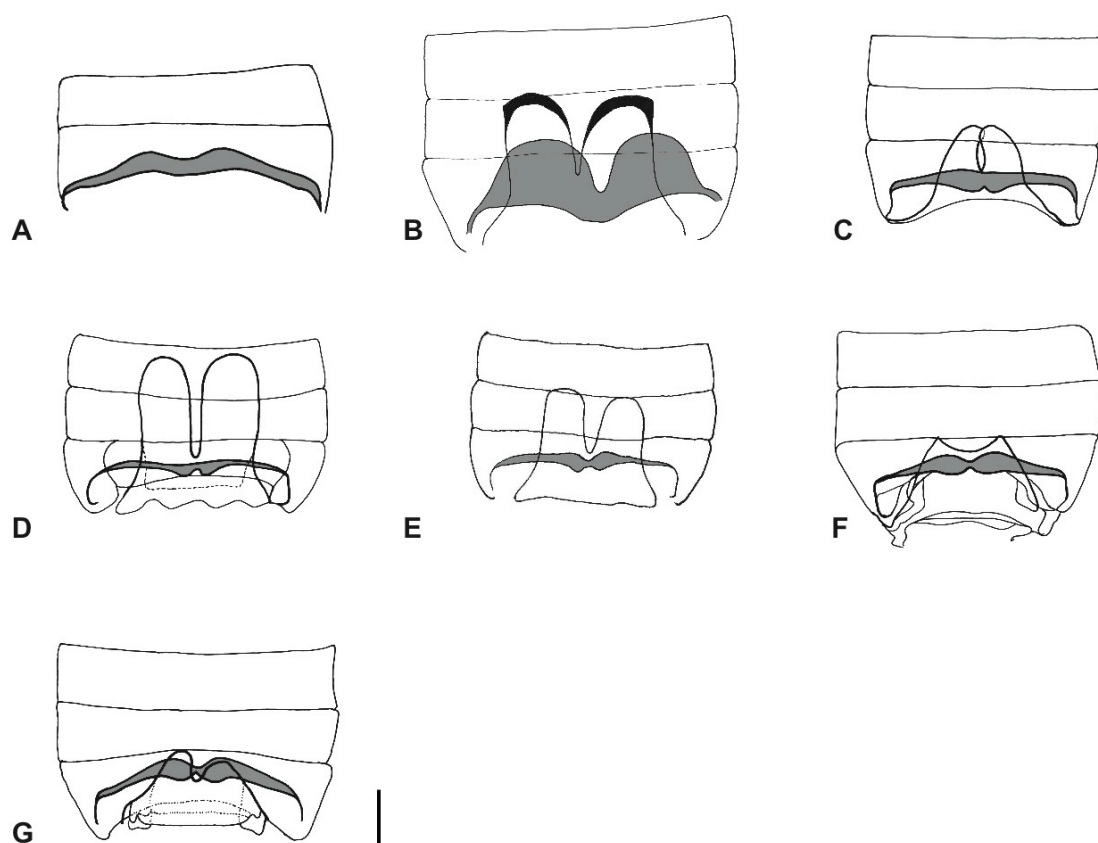


Figure 7. A-E, apodemes of *Kybos* spp. A, *K. smaragdulus*; B, *K. sordidulus*; C, *K. strigilifer*; D, *K. virgator*; E, *K. virgator*, intermediate specimen; F, *K. virgator*, teneral specimen; G, *K. virgator*, teneral specimen. Scale bar = 0.2 mm.

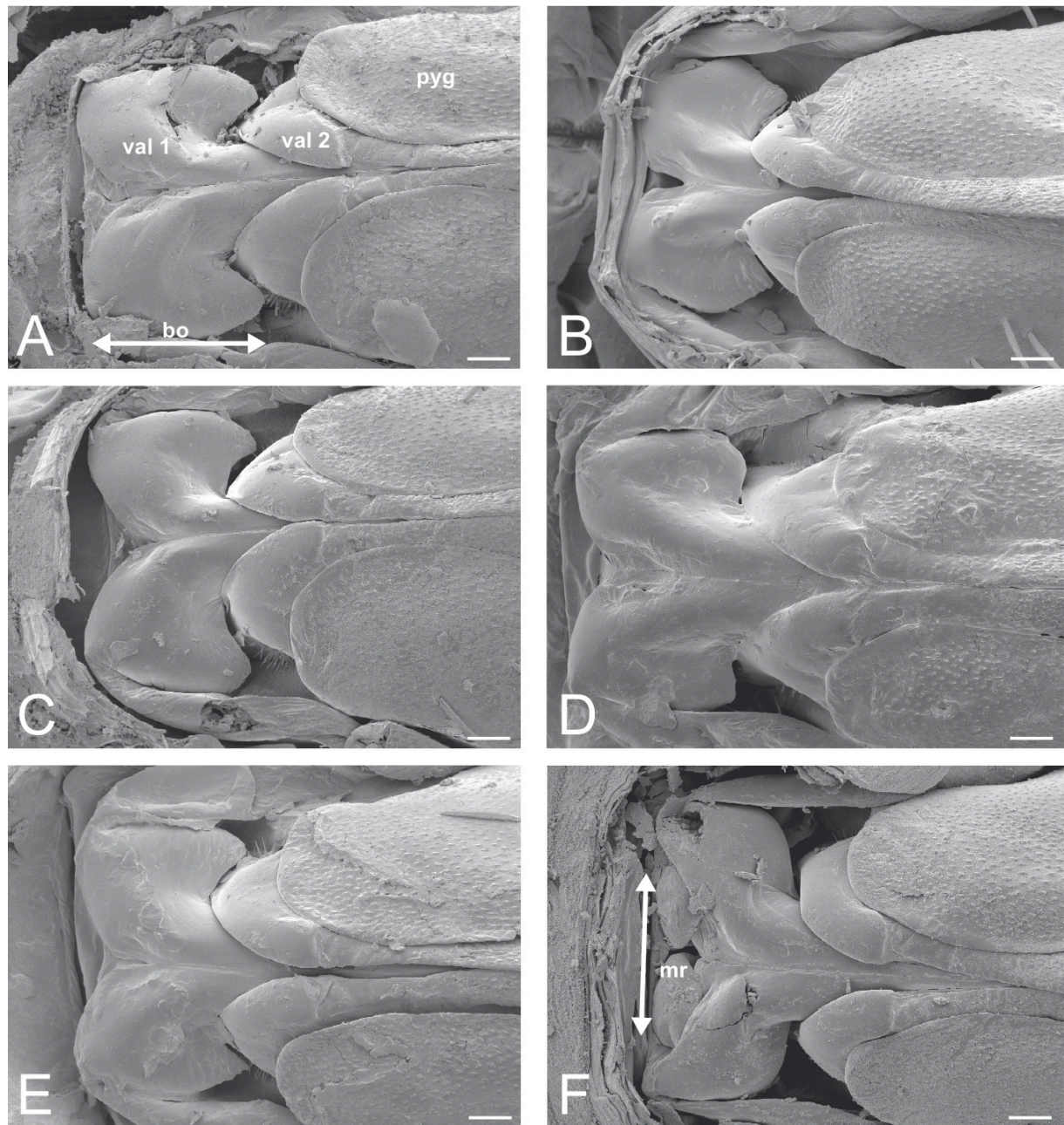


Figure 8. A-F, base of ovipositors of *Kybos* spp. (ventral view). A, *Kybos abstrusus*; B, *K. butleri*; C, *K. limpidus*; D, *K. populi*; E, *K. rufescens*; F, *K. austriacus*. Abbreviations: bo, base of ovipositor; mr, median ridge; pyg, pygofer; val 1, first valvifer; val 2, second valvifer. Scale bars = 50 μ m.

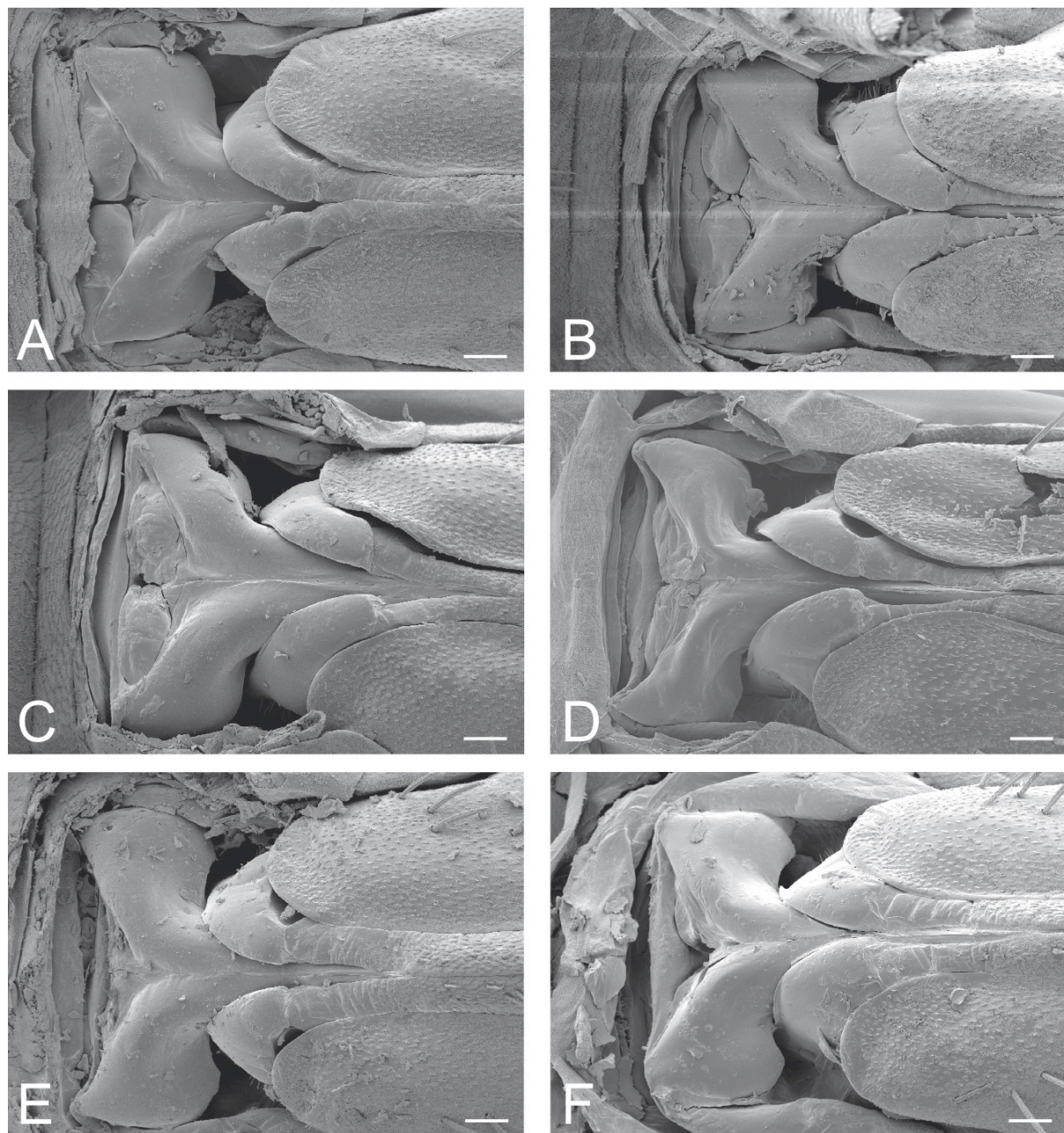


Figure 9. A-F, base of ovipositors of *Kybos* spp. (ventral view). A, *Kybos lindbergi*; B, *K. smaragdula*; C, *K. strobli*; D, *K. digitatus*; E, *K. strigilifer*; F, *K. virgator*. Scale bars = 50 μ m.

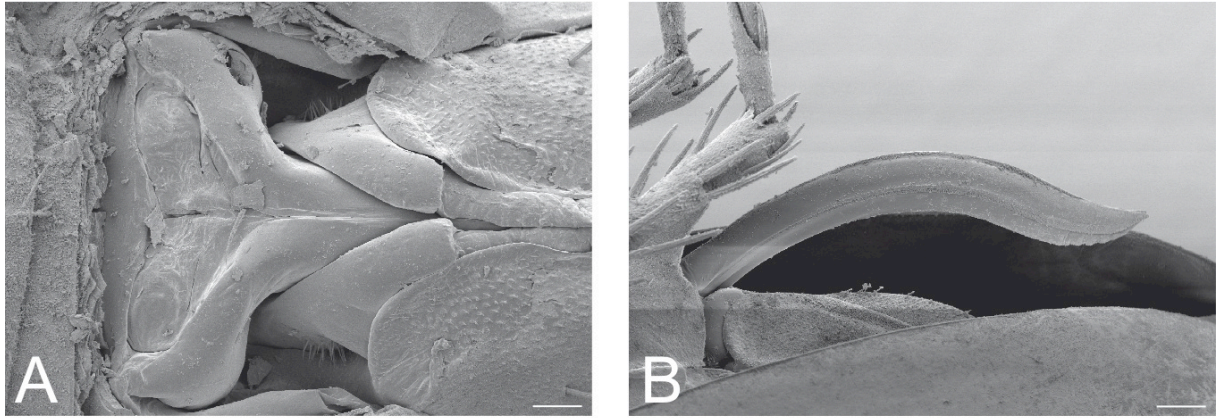


Figure 10. A, *Kybos mucronatus*, base of ovipositors (ventral view); B, *K. smaragdula*, ovipositor (lateral view). Scale bars: A = 50 μm , B = 100 μm .

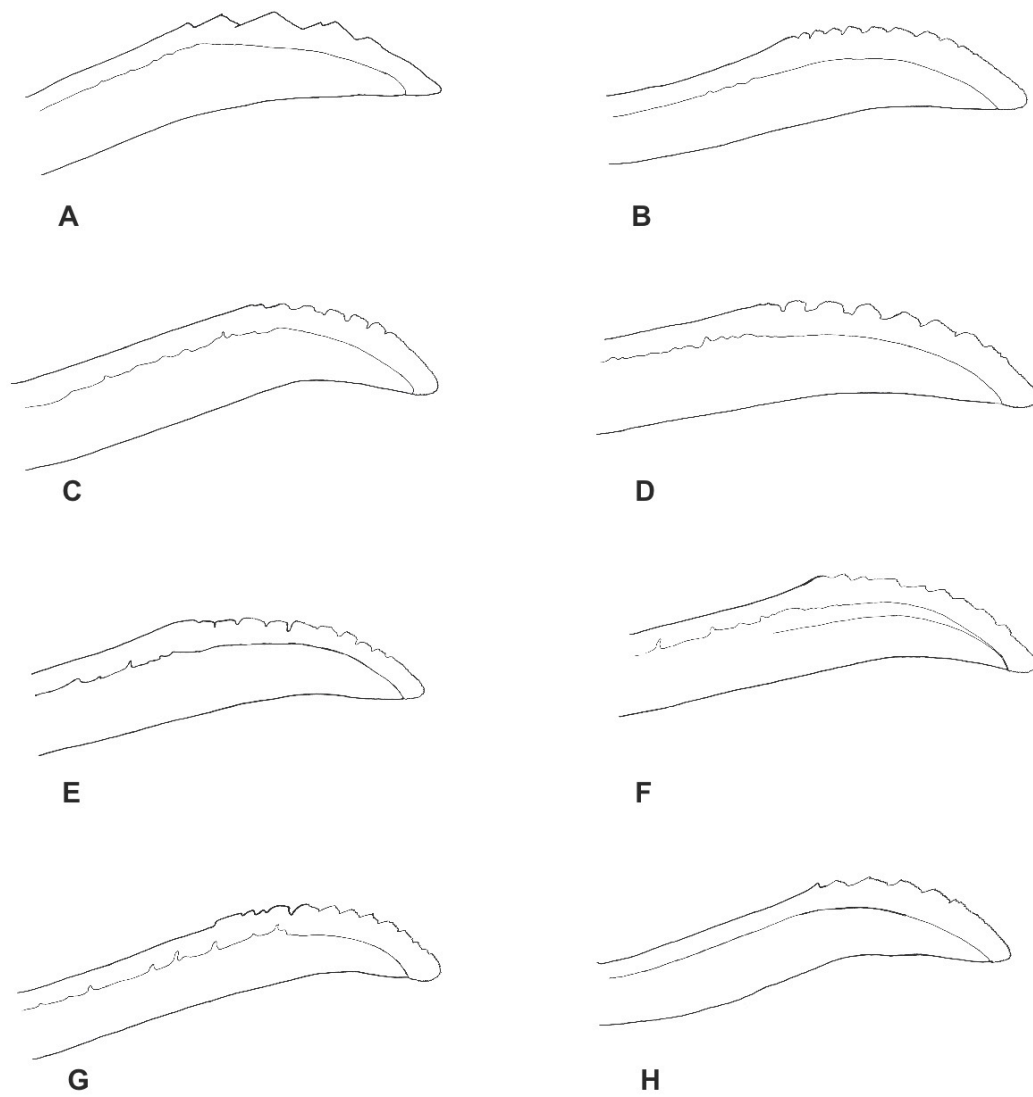


Figure 11. A-H, apex of valvifer (lateral view). A, *Kyboasca bipunctata*; B, *Kybos butleri*; C, *Kybos lindbergi*; D, *Kybos limpidus*; E, *Kybos mesasiaticus*; F, *Kybos populi*; G, *Kybos rufescens*; H, *Kybos smaragdulus*. Scale bar = 0.1 mm.

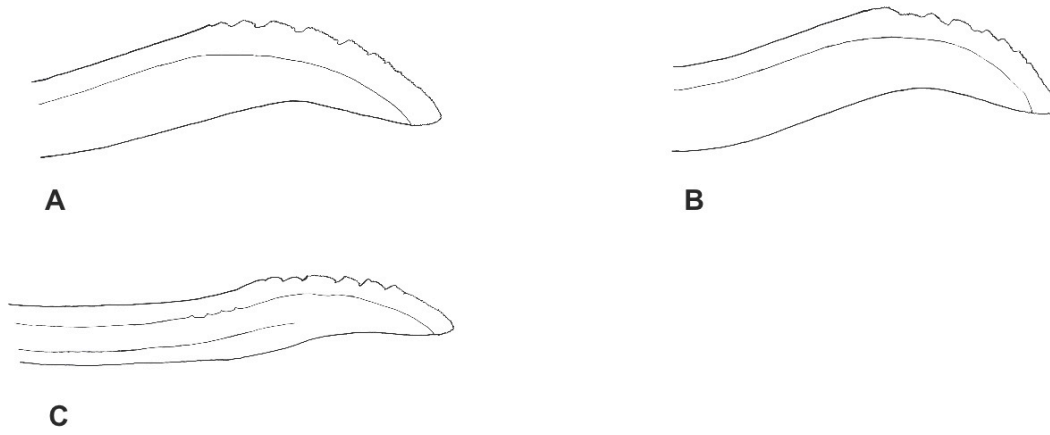


Figure 12. A-C, apex of valvifer (lateral view). A, *Kybos strigilifer*; B, *Kybos strobli*; C, *Kybos virgator*. Scale bar = 0.1 mm.

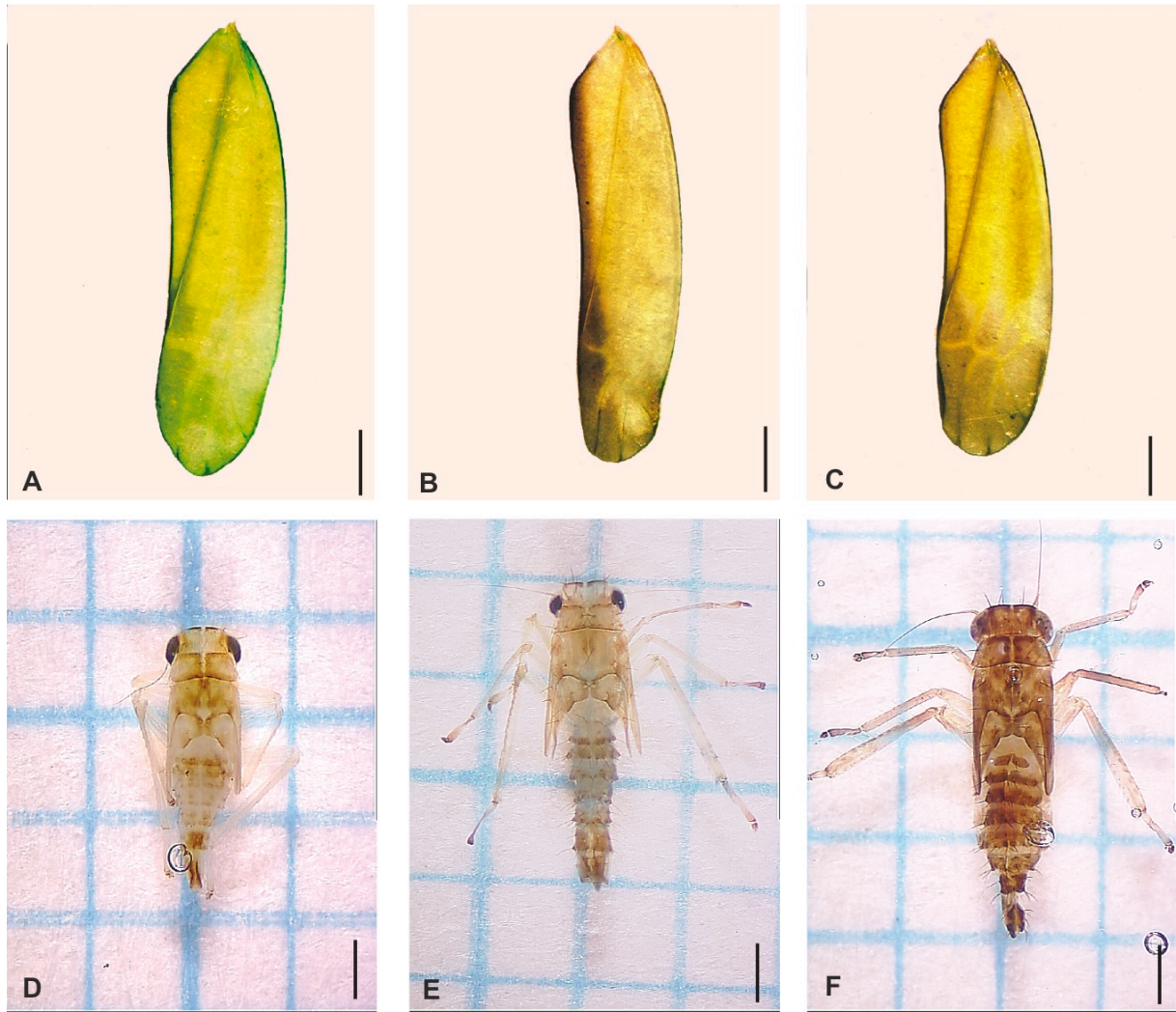


Figure 13. A-C, colour patterns of forewings of *Kybos* spp. A, *Kybos lindbergi*; B, *Kybos rufescens*; C, *Kybos strigilifer*; D-F, nymphs of *Kybos* spp. D, *Kybos limpidus*; E, *Kybos smaragdulus*; D, *Kybos virgator*. Scale bars = 0.5 mm.

3.1.4 Key to *Kybos* species

1	Male	2
	Female	20
2(1)	Colour of commisural border of fore wing concolorous	3
	Colour of commisural border of fore wing bearing a brown band	19
3(2)	Colour of face fuscous	sordidulus
	Colour of face greenish or yellowish	4
4(3)	Aedeagus without appendages	5
	Aedeagus with appendages	8
5(4)	Pygofer process in male broadend apically (Fig. 5L)	populi
	Pygofer process in male slender apically	6
6(5)	Aedeagal stem, in profile, thick. On <i>Populus</i> spp.	abstrusus
	Aedeagal stem, in profile, slender. On <i>Salix</i> spp.	7
7(6)	Aedeagus bearing conspicuous spines ventrally (Fig. 2E); phragma lobes of tergite 3 shorter than broad (Fig. 6G). On <i>Salix viminalis</i>	limpidus
	Aedeagus without conspicuous spines ventrally; phragma lobes of tergite 3 longer than broad (Fig. 6J).	mesasiaticus
8(4)	Aedeagus with parallel-sided appendages	9
	Aedeagus with divergent appendages	13
9(8)	Basis of appendages of aedeagus widely apart from each other	10
	Basis of appendages of aedeagus close to each other	12
10(9)	Process of anal collar long and slender (Fig. 4E). On <i>Salix eleagnos</i>	digitatus
	Process of anal collar short and stout (e.g. Fig. 4L). On <i>Alnus</i> spp. or <i>Betula</i> spp.	11
11(10)	Length of apodemes of sternite II as long as broad or longer. On <i>Betula</i> spp.	ludus
	Length of apodemes of sternite II shorter than broad. On <i>Alnus</i> spp.	smaragdula
12(9)	Aedeagal appendages broad and almost as long as stem (Fig. 2M)	austriacus
	Aedeagal appendages slender and shorter than stem (Fig. 2T)	lindbergi
13(8)	Process of anal collar very long and slender (Fig. 4P)	virgator
	Process of anal collar short and stout (e.g. Figs. 4A, 4C, 4O)	14
14(13)	Aedeagal appendages strongly curved in ventral view (Fig. 2P)	candelabricus
	Aedeagal appendages straight in ventral view (e.g. Figs. 2K, 2O, 3A)	15
15(14)	Aedeagal appendages, in ventral view, as long as stem (Fig. 2K)	aetnicola
	Aedeagal appendages, in ventral view, distinctly shorter than stem	16
16(15)	Length of apodemes of sternite II longer than broad (Figs. 6D, 7C)	17
	Length of apodemes of sternite II shorter than broad (e.g. Fig. 6K)	18
17(16)	Aedeagal appendages, in lateral view, ventral of main stem. On <i>Betula</i> spp.	calyculus
	Aedeagal appendages, in lateral view, parallel to main stem. On <i>Salix</i> spp.	strigilifer
18(16)	Aedeagal appendages, in lateral view, parallel to main stem, blunt apically (Fig. 2W)	mucronatus
	Aedeagal appendages, in lateral view, dorsal of main stem, pointed apically (Fig. 2G)	strobli
19(1)	Length of phragma lobes of tergite III shorter than broad (Fig. 6M)	rufescens
	Length of phragma lobes of tergite III longer than broad, clearly separated medially, diverging (Fig. 6C)	butleri
20(1)	Colour of commisural border of fore wing bearing a brown band	21
	Colour of commisural border of fore wing concolorous	22
21(20)	Outer margin of base of first valvifer distinctly angular (Fig. 8B)	butleri
	Outer margin of base of first valvifer weakly rounded (Fig. 8E)	rufescens
22(20)	Base of first valvifer, in ventral view, narrow, angular, straight distally, bearing a median transverse ridge (e.g. Fig. 8F)	23

Base of first valvifer, in ventral view, broad, irregularly rounded, often hook-shaped distally, lacking a median transverse ridge (e.g. Fig. 8A, 8D).....	26
23(22) Apex of median valvula bearing obliquely triangular teeth (e.g. Figs. 11H, 12 B). On <i>Alnus</i> spp.	24
Apex of median valvula bearing irregular, blunt teeth (e.g. Figs. 11C, 12C). On <i>Betula</i> spp. or <i>Salix</i> spp.	25
24(23) Apex of median valvula weakly curved	<i>mucronatus</i>
Apex of median valvula strongly curved (Figs. 11H, 12B).....	<i>smaragdula, strobli</i>
25(23) On <i>Betula</i> spp.	<i>austriacus, calyculus, lindbergi, ludus</i>
On <i>Salix</i> spp.	<i>candelabricus, digitatus, strigilifer, virgator</i>
26(22) Base of first valvifer, in ventral view, angular distally. Sternite VII without a prolonged lobe, evenly rounded (Fig. 8D).	27
Base of first valvifer, in ventral view, hook-shaped distally. Sternite VII with a prolonged lobe (Figs. 8A, 8C).....	28
27(26) On <i>Populus</i> spp.	<i>populi</i>
On <i>Salix</i> spp.	<i>sordidulus</i>
28(26) On <i>Populus nigra</i>	<i>abstrusus</i>
On <i>Salix</i> spp.	<i>limpidus, mesasiaticus</i>

3.1.5 Molecular evidence

There are no COI or 16S sequences available at GenBank for *Kybos* or other members of the Empoascini. The cytochrome c oxidase subunit I (COI) of the mitochondrial DNA is often used to investigate the relationships of closely related species. This essentially non-recombining genome, inherited only by the mother, leads to shorter coalescence times (Sperling, 2003). COI has been proposed as the gene for barcoding animals (Hebert *et al.*, 2003).

COI sequences were obtained for 46 specimens and 16S sequences for 51 specimens. For the analyses only these specimens were retained where both, COI and 16S sequences were obtained. We sequenced 769 bp for COI and 388 bp for 16S. The base frequencies over all are: A=0.33084, C=0.12960, G=0.13396, T=0.40560.

We aligned the raw COI and 16S sequences manually (excluding highly length-variable regions) and analysed the aligned sequences separately and together (Appendices 2-3).

Following species are composed of genetically homogeneous specimens with identical or almost identical COI sequences: *mucronatus* + *strigilifer/perplexus* + *virgator/volgensis*, *calyculus*, *digitatus*, *smaragdula* (except sample 1449 which is closer to the group of *mucronatus/strigilifer/virgator*). Of particular interest are the identical sequences of *K. virgator/volgensis* from the same locality. This corroborates the conclusions drawn from morphological evidence that the two are synonyms. *K. strigilifer/perplexus* from the same locality are identical, but are also identical with the sequences of *mucronatus* and *virgator*. With respect to the synonymy of *strigilifer* and *perplexus* the molecular data do neither

support nor contradict morphological evidence. In the case of *K. lindbergi* and *ludus* the sequences are similar but not identical. The molecular data is not conclusive but does not contradict the morphological evidence.

The situation is quite different in *K. butleri* and *K. rufescens*. The former consists of two well defined genotypes (Figs. 14-15), the latter shows an unusually large genetic variation (Fig. 16). In both species neither morphology nor host plant association suggest that there may be cryptic species involved. The two genotypes of *K. butleri* cannot be attributed to distribution as both samples contain specimens from different localities. It is of importance to note that the two genotypes do not consist of the same specimens regarding COI and 16S. The specimens 1415/1498, respectively 1305/1496/1497 represent the two genotypes in COI, whereas in 16S sequences 1305/1498, respectively 1415/1496/1497 form the two separated genotypes.

Generally the 16S sequences show less differences than COI and are therefore less informative for the taxonomy.

37

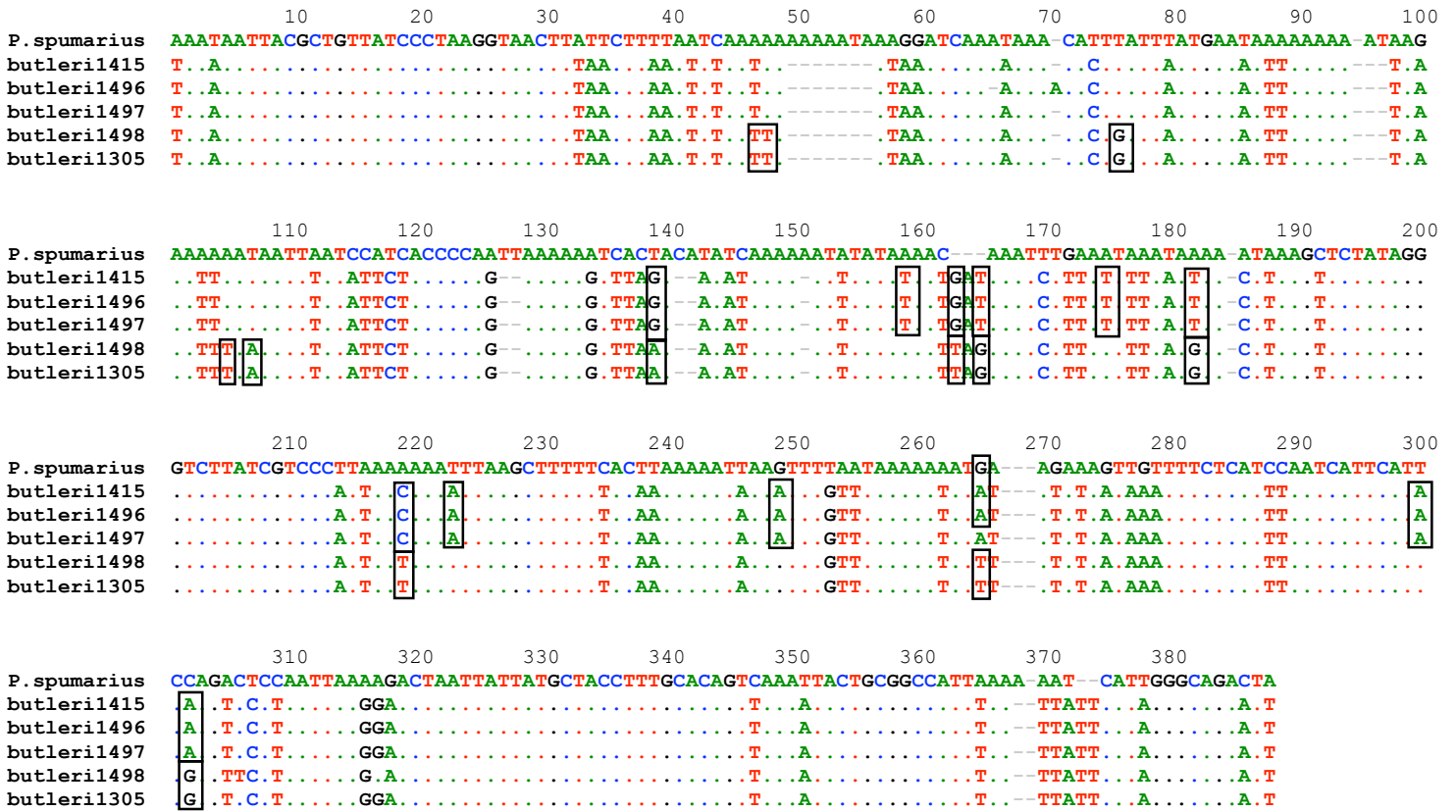


Figure 15. 16S sequences of *Kybos butleri*, positions of the two genotypes are marked with squares. Numbers indicate base pair positions.

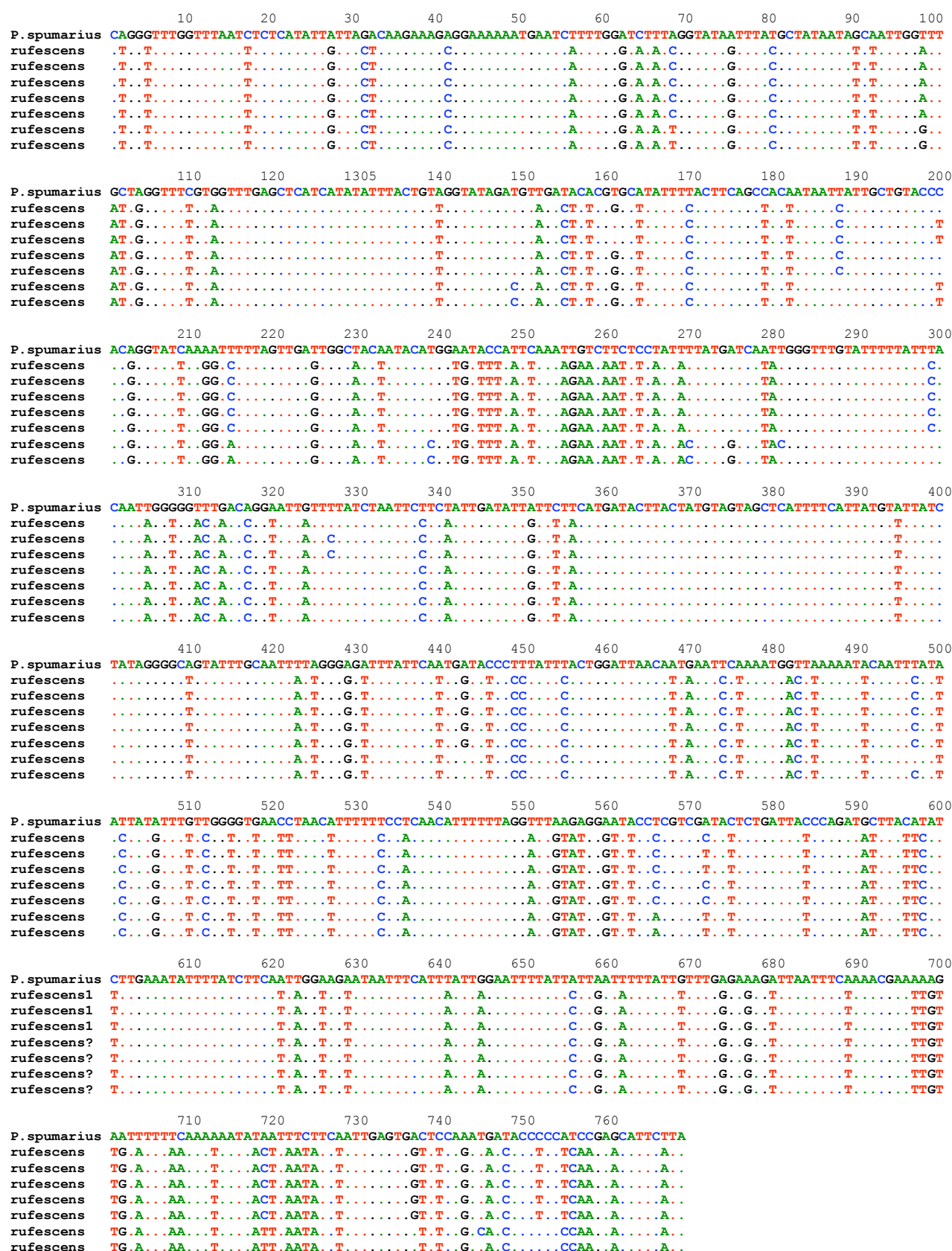


Figure 16. COI sequences of all examined *Kybos rufescens* individuals. Numbers indicate base pair positions.

3.1.6 Bioacoustics

We recorded male songs of the following species: *Kybos rufescens*, *K. strobli* and *K. lindbergi*. Unpublished songs of *Empoasca* s.str. were available from other studies made by the research group in Ljubljana (M. Virant-Doberlet, pers. comm.). Males were calling all the time whereas females responded only if they were virgins. Already mated females did not reply to the male calling songs.

The sonograms (Fig. 17) show the time pattern of the calls as well as the frequency range (spectrograms). The common male calling song of *K. lindbergi* has a main frequency of about 0.3 kHz which rises to around 1.0-1.5 kHz at the end of the bursts. The song consists of usually three short bursts, each not longer 1 s. The starting burst is always longer than the following ones. *K. strobli* possesses a monotonic call of about 5-6 s. At the beginning the frequency is low (0.2 kHz) and increases up to 2.0-2.5 kHz. In the sonogram of *K. rufescens* we see both, the common male calling song and the female response. The male song consists of two strong bursts with a main frequency of 2.0 kHz and a second somewhat weaker burst at 3.6 kHz. Each burst lasts just under 1 s. The female response is extremely weak and of low frequency (around 0.1 kHz). Also the time of the bursts are very short (0.1-0.25 s). Two of the female bursts are only visible on the spectrogram because they are simultaneous with the male calls. The sonogram depicts only a portion of a longer duet between the sexes.

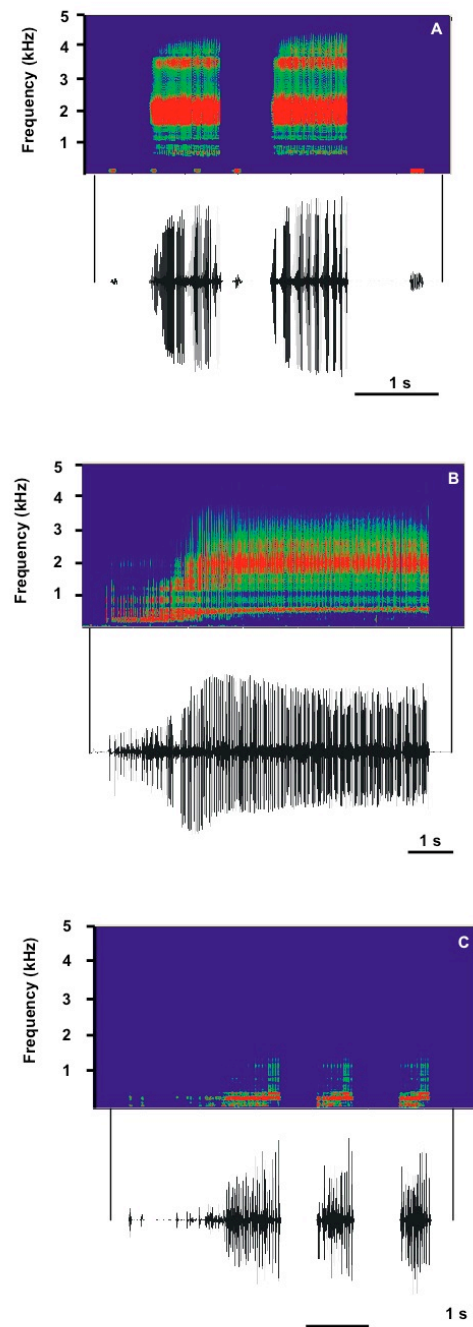


Figure 17. A-C, sono- and spectrograms of the common male calling song of *Kybos* spp. A, *K. rufescens*; B, *K. strobli*, C, *K. lindbergi*.

3.2 Phylogeny

3.2.1 Morphological analysis

A cladistic analysis was performed with 21 taxa (*Kyboasca* 1 species, *Empoasca* 1 species, *Kybos* 19 species) using 16 characters (Tables 1-2). Three characters relate to the wings, 6 to the aedeagus, 1 to the anal collar, 5 to the female genitalia and 1 to the nymph. All characters are binary except for characters 6, 10 and 15 which are multistate. A heuristic analysis was performed using NoNa (Goloboff, 2001), WinClada (Nixon, 2002) and PAUP*4.10 (Swofford, 1998). The analysis resulted in 48 most parsimonious trees with following parameters: length = 43, consistency index = 62, retention index = 80.

Fig. 18 shows the consensus tree (nelsen) with a strongly supported monophyletic *Kybos* (arrow). Within *Kybos* the general resolution is relatively weak. There are 6 groups: group 2 is strongly supported and includes the taxa with aedeagal appendages. Groups 1, 3 and 4 are weakly supported by a single synapomorphy each and groups 5 and 6 are supported only by homoplasies. Group 4 contains all the taxa associated with Betulaceae.

Of the 16 characters used in the analysis nine constitute synapomorphies (Fig. 18, dark circles). Four of the synapomorphies refer to female characters, three to wing characters and only two to aedeagal characters.

Table 1. Morphological characters used for cladistic analysis (Table 2, Fig. 18).

1. Second apical cell in fore wing stalked (0); not stalked (1);
2. Commisural border of fore wing concolorous (0); bearing a brown band (1);
3. Colour of corioclaval suture of fore wing black or fuscous (0); concolorous with forewing (1);
4. Aedeagus without appedages (0); with appendages (1);
5. Aedeagus with divergent appedages (0); with parallel sided appedages (1);
6. Appedages of aedeagus (in lateral view) parallel to main stem (0); dorsally of main stem (1); ventrally of main stem (2);
7. Length of aedeagal appedages as long as stem (0); shorter than stem (1);
8. Basis of aedeagal appedages widely apart from each other (0); close to each other (1);
9. Aedeagal appedages apically pointed (0); blunt (1);
10. Process of anal collar very short and pointed (0); long and slender (1); short and stout (2);
11. Female sternit 7 with a prolonged lobe (0); without a prolonged lobe (1);
12. Ventral edge of median valvula apically evenly curved (0); apically strongly curved (1);
13. Structure of teeth of valvula regularly triangular (0); irregularly blunt (1);
14. Base of first valvifer with a median ridge (0); without a median ridge (1);
15. Distal edge of base of first valvifer straight (0); rounded 'hook-shaped' (2);
16. Length of larval tibia 3 about half of the body length (0); less than half of the body length (1).

Table 2. Matrix of the morphological characters used for the cladistic analysis (Fig. 18). For characters see Table 1.

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
<i>bipunctata</i>	1	0	1	0	-	-	-	-	-	0	1	0	0	1	0	?
<i>vitis</i>	1	0	1	0	-	-	-	-	-	1	1	0	0	1	1	?
<i>abstrusus</i>	0	0	1	0	-	-	-	-	-	1	0	0	1	1	2	?
<i>aetnicola</i>	0	0	0	1	0	2	0	1	0	2	0	1	1	0	0	0
<i>austriacus</i>	0	0	0	1	1	0	0	1	0	2	0	1	1	0	0	?
<i>butleri</i>	0	1	1	0	-	-	-	-	-	1	0	0	1	1	2	?
<i>calyculus</i>	0	0	0	1	0	2	1	1	0	2	0	1	1	0	0	?
<i>candelabricus</i>	0	0	0	1	0	0	0	0	0	2	0	0	1	0	1	?
<i>digitatus</i>	0	0	0	1	1	1	1	0	1	1	0	0	1	0	1	?
<i>limpidus</i>	0	0	1	0	-	-	-	-	-	1	0	0	1	1	2	1
<i>lindbergi</i>	0	0	0	1	1	0	1	1	0	2	0	1	1	0	0	0
<i>ludus</i>	0	0	0	1	1	0	1	0	0	2	0	1	1	0	0	0
<i>mesasiaticus</i>	0	0	1	0	-	-	-	-	-	1	0	0	1	1	2	?
<i>mucronatus</i>	0	0	0	1	0	2	1	1	1	2	0	0	1	0	0	?
<i>populi</i>	0	0	1	0	-	-	-	-	-	1	1	0	1	1	2	1
<i>rufescens</i>	0	1	1	0	-	-	-	-	-	1	0	0	1	1	2	?
<i>smaragdula</i>	0	0	0	1	1	1,2	1	0	0	2	0	1	0	0	0	1
<i>sordidulus</i>	0	0	1	0	-	-	-	-	-	1	1	0	1	1	2	?
<i>strigilifer</i>	0	0	0	1	0	0	1	1	0	2	0	1	1	0	1	?
<i>strobli</i>	0	0	0	1	0	1	1	1	0	2	0	1	0	0	0	?
<i>virgator</i>	0	0	0	1	0	0	1	1	0	1	0	0	1	0	1	?

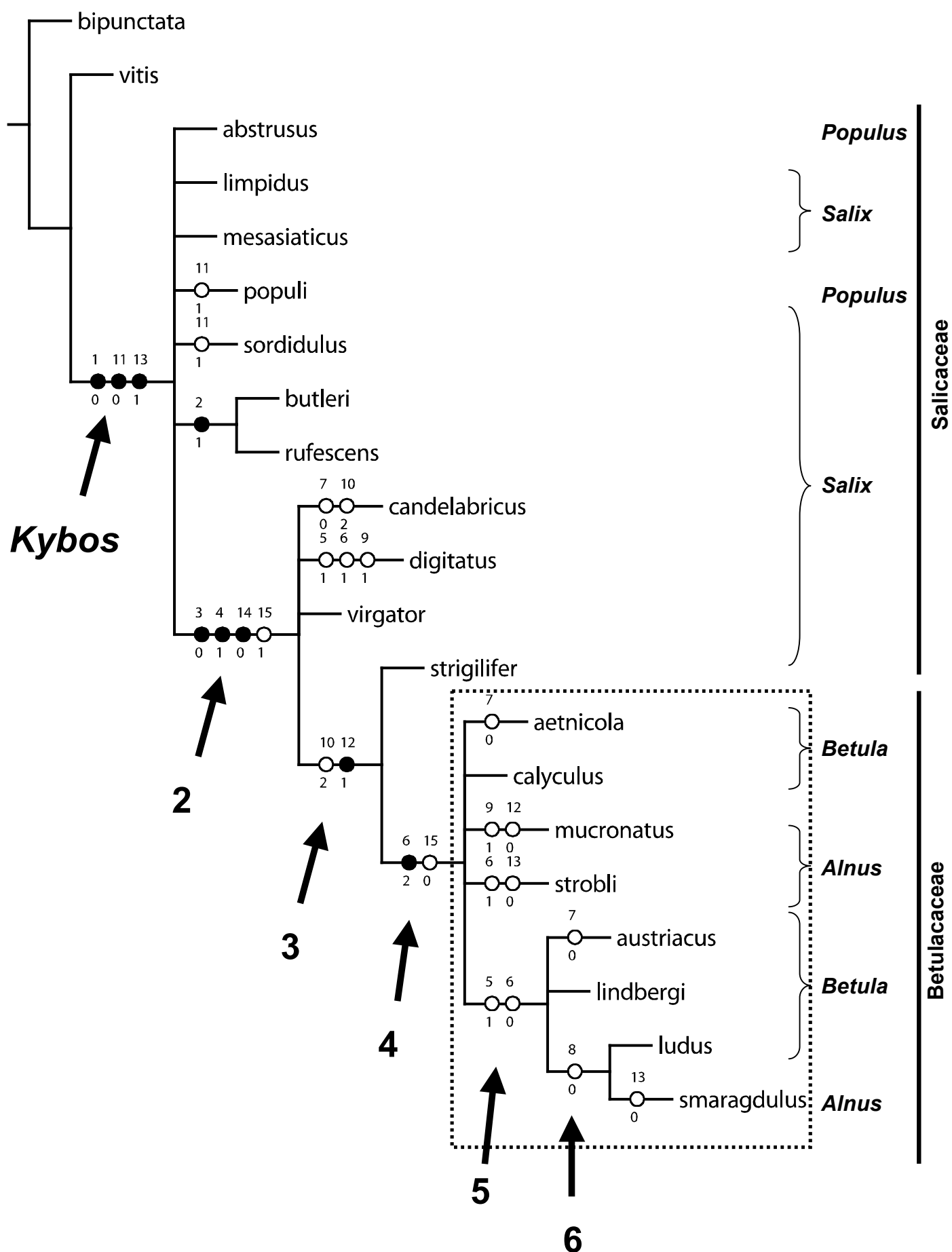


Figure 18. Cladogram based on morphological characters compared to host plant information

3.2.2 Molecular analyses

Within *Kybos* species 479 of the 769 bp of COI are constant and 178 characters are parsimony-informative. Gaps were treated as "missing" characters. The resulting tree has the following parameters: length = 548, consistency index = 0.6971, retention index = 0.8061, homoplasy index = 0.3029. In the 16S sequences 226 of the 382 characters are constant, 72 bp are parsimony-informative. Gaps were treated as "missing" characters. The obtained tree has following parameters: length = 245, consistency index = 0.841, retention index = 0.843, homoplasy index = 0.159.

Fig. 19 shows the tree obtained from COI, fig. 20 the resulting tree from 16S. The tree from COI is better resolved than the one from 16S. *Kybos* forms in both trees a monophyletic group. In the COI we can recognize following well supported groups: *calyculus*, *digitatus*, *lindbergi/ludus*, *smaragdula* and the Central Asiatic group including *dentifera*, *mesasiaticus*, *soosi*. *K. butleri* is polyphyletic and appears at two different places and does not consist of the same groupings in COI and 16S.

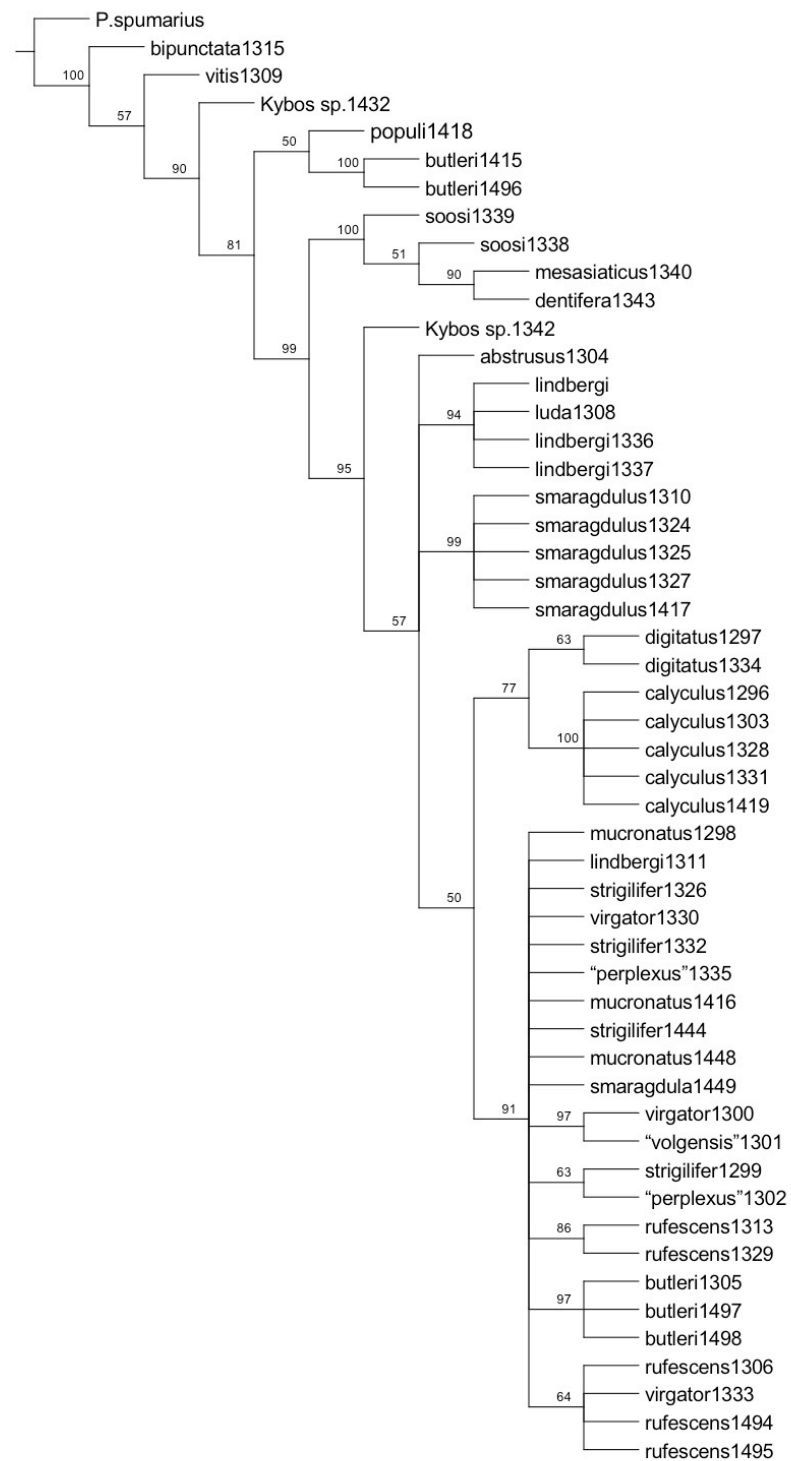


Figure 19. Consensus tree obtained from COI sequences. Numbers on branches indicate bootstrap values.

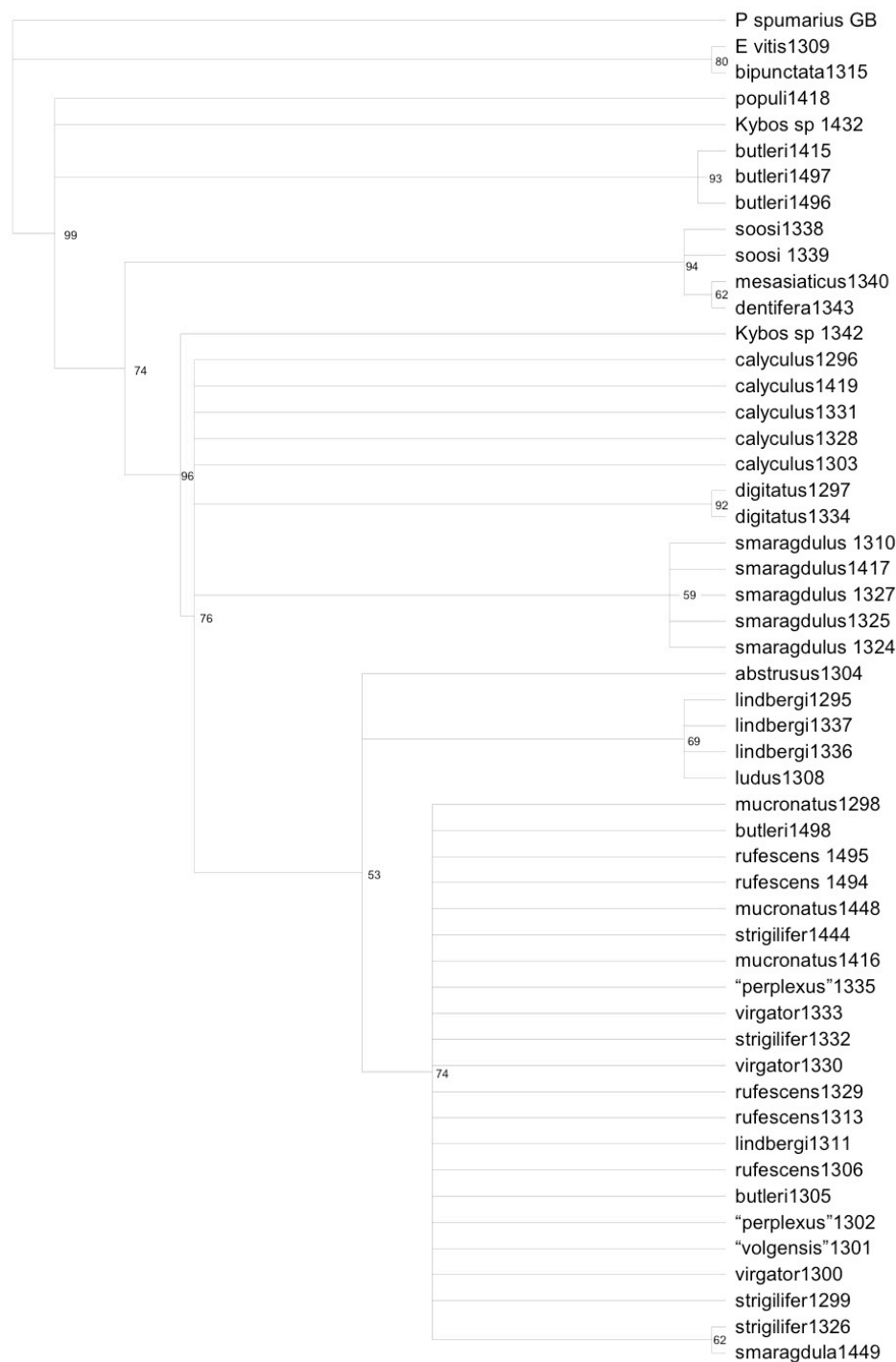


Figure 20. Consensus tree obtained from 16S sequences. Numbers on branches indicate bootstrap values.

3.3 Distribution and host associations

Most Central European *Kybos* species are widely distributed and occur in high abundance where their host plants grow. They generally occur also on a wide altitudinal range (e.g. *K. rufescens* and *K. lindbergi* in Switzerland from Basel at 280 m and Bever at 1700 m). Only a few species are restricted. *K. aetnicola* is endemic to higher elevations of Mt. Etna, Sicily, *K. austriacus* occurs in higher elevations the Austrian and Swiss Alps but is always very rare. Also *K. strobli* is restricted to higher elevations in the Alps but has a wider distribution and is relatively common.

K. ludus and *K. lindbergi* form in Europe a vicariant species pair: *K. ludus* in the West and Southwest, as well as *K. lindbergi* in the East and Northeast respectively (Fig. 21). The two species occur together in a small area in Germany and Switzerland where morphological intermediates can be found relatively frequently. This area is a possible hybrid zone.

Unlike most Central European species *K. calyculus* and *K. digitatus* are only rarely collected and little is known about their biology.

All species outside Europe and most European species are associated with *Salix* and *Populus* species. Some European species are associated with *Alnus* or *Betula* spp. Detailed information on host ranges is lacking for most species. From the available field data it appears that species are restricted to a single host genus where they are narrowly oligophagous on a few closely related species. Only a few species are monophagous on a single host species: *K. digitatus* on *S. elaeagnos*, *K. rufescens* on *S. purpurea*, *K. abstrusus* on *P. nigra*, *K. aetnicola* on *B. aetnensis*. For some species confirmed host information is scarce (*Kybos mucronatus* only on *A. glutinosa* and *K. strobli* on *A. incana*?).

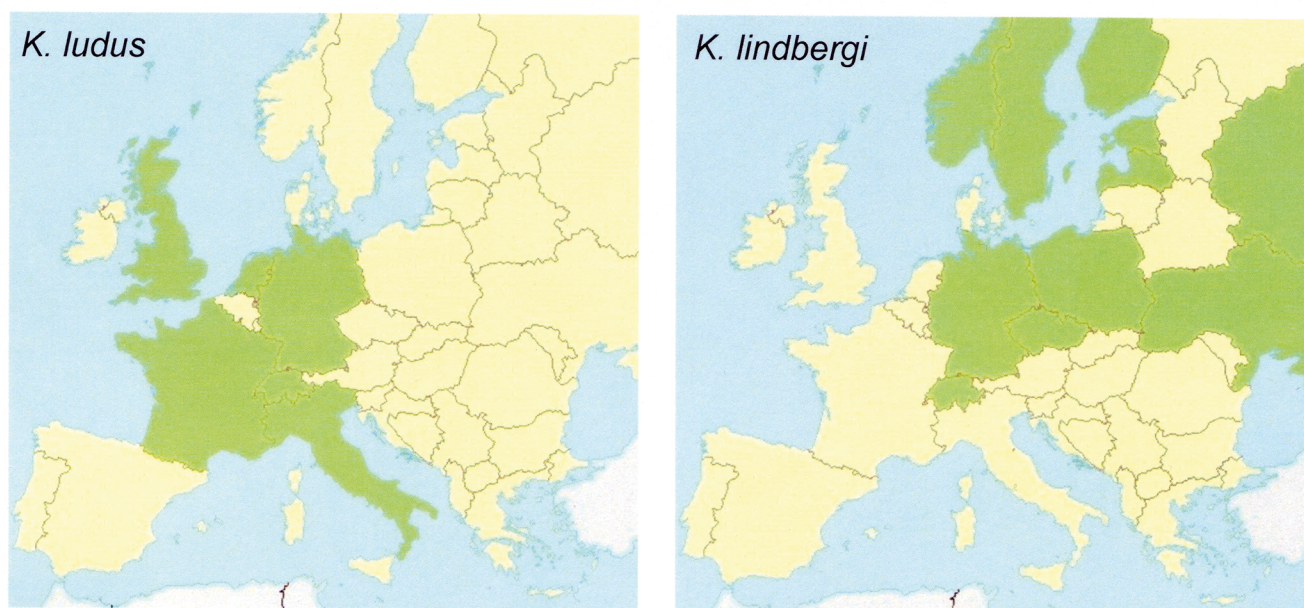


Figure 21. Distribution of *Kybos ludus* and *K. lindbergi* in Europe (after Fauna Europaea, revised).

4 Discussion

4.1 Taxonomy

As in many other cicadellid genera, the morphological differences between *Kybos* species are generally small. Species are mostly diagnosed by the aedeagus and the apodemes, both structures of the male. Additional diagnostic characters previously used are the anal collar and pygofer of the male, fore wing pattern and colour, sternite VII of female and host plant association. This leads to the unsatisfactory situation that in many cases females and nymphs cannot be identified and that identification without host information can be tricky. In addition there is the problem with the apodemes, that they reach their final extension only a few days after the final adult moult. Günthart (1977) documented this variation in other typhlocybina taxa (several *Flammigeroidia* species, *Empoasca decipiens* and *Eupteryx melissae*). The aedeagal shape is considered to be a good species diagnoser but there are not enough studies investigating the intraspecific variability of this character. In fact, this may be one of the reasons why currently also the identification of some males can be difficult. At the current state it seems probable that some nominal species have been described on the basis of teneral males with undeveloped apodemes, and others on too narrowly interpreted aedeagal variations. We expected that a thorough revision of *Kybos* species using morphology would reduce the number of currently recognised Central European species. We also expected that additional diagnostic characters in females and nymphs may be found.

Despite detailed investigations our study did not yield additional morphological characters in the males for species diagnosis but we confirm that the two structures (aedeagus and apodemes) are, in principal, the major species diagnosers.

Our assumption that previously teneral specimens with undeveloped apodemes were described as distinct nominal species was confirmed in the following example. *K. volgensis* differs from *K. virgator* only in the shorter apodemes as well as some minor differences in the shape of the aedeagus. The difference of the aedeagal appendages does not exceed the variability found within a single population of *K. virgator*. Here we suggest that *K. volgensis* is a junior synonym of *K. virgator*. Depending on the collecting date a large number of immature adults can be found which may lead to the erroneous conclusion that the shape of the apodemes is fully developed. This suggestion can be easily verified by additional sampling at the same site a few days later.

The problem of intraspecific aedeagal variation is more difficult to resolve. It is necessary to examine a large number of specimens and carry out breeding experiments. Such breeding experiments were made by Günthart (1974). She showed that the variation of the

aedeagus within *K. strigilifer* is marked but she did not draw any taxonomic conclusions. The present study documents that the characters diagnosing *K. perplexus* (and *paraltaicus*) fall within the variation encountered in *K. strigilifer*. *K. perplexus* is therefore synonymised with *K. strigilifer*. Looking outside the Central European fauna the large number of species known from a few specimens only is striking. Our hypothesis is that a number of these taxa will turn out to be synonyms of other taxa.

Other characters such as the wing coloration and the anal collar and pygofer of males bear only limited taxonomic information on species level. They are mostly useful to define species groups. We observed more or less the same with the female genitalia. Here the most important diagnostic character is the shape of the ovipositor base. With a few exceptions (*K. butleri*, *K. populi*) it is not possible to diagnose species with female sexual characters. Nymphal characters are even less informative.

The bioacoustic studies suggest that the songs are highly diagnostic. Probably one of the main purposes of these calls is to attract and find mates. With this hypothesis we would expect that closely related sympatric species differ strongly in their songs. The sound is produced by the so called apodemes. These are specialised internal, often striated areas of cuticle located at the two or three basal abdominal tergites and sternites respectively. Differences in the shape of the apodemes are responsible for different sounds (but not necessarily vice versa). The songs of the three recorded species are drastically different but more species should be examined. Comparing *Kybos* songs with songs of representatives of *Empoasca* s.str. show that there are significant differences in frequency modulation.

Regarding the molecular data we observed that the COI sequences are more informative than those from 16S mtDNA. But the "barcoding gene" COI is only partially informative within the genus *Kybos* with respect to species diagnosis. Some species such as *K. calyculus* and *K. digitatus* are homogenous, many others, however, are quite variable. Remarkable are the two haplotypes of *K. butleri*. They are not related to geographically separated populations because specimens from a particular locality represented a mixture of both haplotypes. Failure of COI to diagnose species was also found in other groups (Sperling 2003; Meier *et al.* 2006).

Kybos is well-characterised by the wing venation, male and female genital structures. Here we add bioacoustic and molecular support for its monophyly. *Kybos* has been treated as genus (mostly by European workers) or as subgenus of *Empoasca* (mostly North American authors). Here we follow the European tradition. *Empoasca*, on the other hand, is in its current state probably paraphyletic.

4.2 Phylogeny

Both the morphological and molecular trees are poorly resolved. But in all trees *Kybos* forms a monophyletic group. In the cladistic analysis based on morphology the female genital structures and wing characters are the most informative ones. Male characters are, in contrast to the taxonomy, much less important for the tree structure. Six groups are recognised of which only one is strongly supported (Fig. 18). Group 2 includes all species which bear aedeagal appendages. The other groups are weakly supported. Interesting is the fact, that group 4 consists of all taxa living on Betulaceae. Our groups are similar to the groups established by Dworakowska (1976) but less resolved. Dworakowska proposed the following groupings based on morphology and host plant information (the species examined here are in bold):

smaragdula-group: *aetnicola*, *austriacus*, *calyculus*, *digitatus*, *lindbergi*, *ludus*, *smaragdulus* and *strobli*

mucronatus-group: *auricillatus*, *candelabricus*, *mucronatus* (incl. *mucronatus verbae*), *strigilifer* (incl. *perplexus*) and *virgator* (incl. *volgensis*)

limpidus-group: *cornutus*, *dlabolai* and *limpidus*

butleri-group: *abstrusus*, *butleri*, *chadchalicus*, *iliensis*, *koreana*, *mesasiaticus*, *mitjaevi*, *niveicolor niveicolor*, *niveicolor japonicus*, *populi*, *pyramidalis*, *rubrovenosus*, *rufescens* (incl. *rufescens rufescens* and *rufescens matsumurai*), *soosi* and possibly *oshanini*

sordidulus belongs to the Nearctic *trifasciatus*-group (established by Ross, 1963)

The topologies of the two molecular cladograms (Figs. 19-20) are similar but with significant differences (e.g. grouping of *K. butleri*). The 16S tree is less resolved than that of COI. Apart from the monophyly of *Kybos* there is no congruence between the morphological and molecular trees. All trees show, that *Kybos* is monophyletic.

We think that the phylogeny obtained from morphological characters is more reliable than the molecular phylogeny. The cladistic analysis based on morphology uses different character systems (wing coloration, male genitalia, female genitalia and nymphal characters). The congruence of these different character systems provides a more stable hypothesis than the trees obtained from molecular data. Predictions are only possible for the morphology, but not for the COI and 16S sequences.

4.3 Biogeography

Both *Kybos* and *Kyboasca* are restricted to the Holarctic Region whereas *Empoasca*, in its present probably paraphyletic definition, has a world-wide distribution. Of the 114 currently known *Kybos* species (Appendix 4), 42 are restricted to the Palearctic, 76 to the

Nearctic and only 4 occur in both regions (*K. butleri*, *K. ludus*, *K. smaragdulus* and *K. sordidulus*). The first three have almost certainly been introduced from the West Palaearctic into the Nearctic. Introductions are documented also for other biogeographic regions. *K. lindbergi*, *K. ludus* and *K. smaragdulus* have been introduced with their host plants into Australia and New Zealand (Dumbleton 1964; Knight 1976; Fletcher & Knight 1998). *K. sordidulus* may be genuinely Holarctic though further research is needed. More species are currently known from the western part of the Nearctic than from the eastern part. As the available information on Nearctic species is scarce it is not possible to decide whether this pattern is real or just an artifact. A similar situation is found in the Palaearctic Region. The knowledge on the West Palaearctic *Kybos* fauna is much more complete than that of the East Palaearctic Region.

In Europe most species are widely distributed but two species are restricted to higher elevations in the Alps (*K. austriacus*, *K. strobli*) and *K. aetnicola* has been recorded only from Sicily. Incidentally the three species belong to the highly derived clade of Betulaceae feeders but they do not form a monophyletic group.

According to Asche & Hoch (2004) (Fig. 21) *K. ludus* occurs mainly in Western Europe, whereas *K. lindbergi* lives in Northern and Eastern Europe. Both species feed on *Betula* and differ only in subtle details of the aedeagus. In Germany and Switzerland both species are sympatric and intermediate forms are frequent. This hybrid zone was already mentioned by Dworakowska (1976). Although our investigations based on morphology and molecular techniques did not clarify the problem of these two closely related species, we suggest to treat the two taxa as distinct species which are geographically separated. Only in a relatively small area in Central Europe we can find hybrids between *K. lindbergi* and *K. ludus*.

4.4 Host plants

Species of *Kybos* and of its presumed sister group *Kyboasca* are generally monophagous on various genera of woody Dicotyledones, the former on Salicaceae and Betulaceae (Appendix 4), the latter on Fabaceae, Rosaceae, Ulmaceae etc. Species of *Empoasca* s.str., in contrast, are polyphagous on a variety of herbaceous and woody dicotyledonous families.

Apart from three West Nearctic species (*K. cascada*, *K. fontana* and *K. incida*) which have been reported from both *Populus* and *Salix* spp. all members of *Kybos* are restricted to a single host genus. The species associated with a particular genus, however, do not form a monophyletic group. Most *Kybos* spp. are associated with Salicaceae except for nine West Palaearctic species which live on Betulaceae (six on *Betula*, three on *Alnus*). It is interesting

to note that the Betulaceae feeders form a monophyletic group in the morphological cladistic analysis (Fig. 18) suggesting a single host shift from Salicaceae to Betulaceae. This host shift happened only in the West Palearctic.

5 Conclusions

The taxonomy of *Kybos* was traditionally based mostly on the morphology of the male genitalia, in particular the aedeagus and the apodemes. Our study on the Central European members confirms that these characters are the prime diagnostic structures. However, intraspecific variation of the aedeagus has previously not been sufficiently examined, a notable exception being the work of Günthart (1974). In some cases individuals with morphological extremes have been attributed to independent species creating taxonomic chaos, e.g. *K. perplexus* as morphological extreme of *K. strigilifer*. Some caution is also required with respect to the apodemes which are only fully developed a few days after eclosion. In the past the apodemes of teneral males have been misjudged and provided the base for the erection of species, e.g. *K. volgensis* as teneral form of *K. virgator*. Other male genital characters such as the pygofer, the stylus and the anal collar, as well as wing coloration, can be used for separating species groups but are not diagnostic at species level. Unfortunately this is also true for the female genitalia and even more so for nymphal characters. We expect that a similar situation can be found in other genera of Typhlocybinae.

As with morphology alone many individuals (mostly females) cannot be identified other character systems are required. Molecular methods have been successfully applied to this sort of problem, in particular sequence data of the genes COI, 16S, wingless, NADH, 18S and others. Good results with this method led to the idea that a single gene fragment may suffice to diagnose each organismal species. This approach, known as “barcoding of life” uses the COI gene (Hebert *et al.* 2003; Tautz *et al.* 2003). After initial euphoria several papers have been published recently demonstrating that COI shows at least in some species considerable variation (Sperling 2003; Hickerson *et al.* 2006; Meyer *et al.* 2006). Our study confirms these observations and adds some more examples to the list of morphologically well-defined species which are supported neither by COI nor 16S, e.g. *K. butleri* and *K. rufescens*. The taxonomic situation within *Kybos* remains unaltered despite our molecular study, i.e. females can often not be attributed to a species with certainty.

As for the taxonomy neither the morphological nor the molecular cladistic analyses provided sufficient information for constructing a fully resolved phylogeny of *Kybos*. Both approaches support its monophyly but otherwise the results from these data sets are quite different. The morphological tree is better supported and is congruent, to a certain extent, with host plant associations. In the absence of a fully resolved tree it is impossible to investigate detailed evolutionary patterns in the diversification of the genus *Kybos*, i.e. the contribution of coevolution with the hosts or host shifts versus geographical vicariance or dispersal as forces

for speciation. It is generally assumed that coevolution is the major force but studies have shown that geographic vicariance is more important at least in some groups (Burckhardt & Basset 2000; Percy 2003; Burckhardt & Ouvrard 2007). In *Kybos* there are no detectable patterns of coevolution or biogeographic vicariance but there is a monophyletic group, restricted to the West Palaearctic, associated with Betulaceae (contrasting the remainder of species on Salicaceae). While most species are widely distributed a few are endemic to small areas at higher elevations (Alps, Mt. Etna).

For resolving the remaining taxonomic problems of Central European *Kybos* species, in particular closely related sympatric taxa, breeding experiments and additional bioacoustic studies are required. For a better resolution of the phylogeny additional taxa, including Nearctic species, should be added and sequences of other genes should be investigated. Additional targeted field work should provide further details on the distribution of individual *Kybos* species as well as host associations.

6 Acknowledgments

This thesis was prepared under the scientific supervision of PD Dr. Daniel Burckhardt (NHMB, Basel, Switzerland) and Prof. Dr. Peter Nagel (Institute for Biogeography, Basel, Switzerland); for all their help and stimulating entomological discussions I am very grateful. Material for the thesis was provided by following colleagues: Pavel Lauterer (MZMB, Brno, Czech Republic), Igor Malenovský (MZMB, Brno, Czech Republic), Petr Kment (NMP, Prague, Czech Republic), Chris Dietrich (Center for Biodiversity, Illinois, USA), Heidi Günthart (Dielsdorf, Switzerland), Herbert Nickel (University of Göttingen, Germany), Denise Wyniger (NHMB, Switzerland and AMNH, New York, USA), Randall Schuh (AMNH, New York, USA), Simone Fattorini (INER, Rome, Italy), Holger Löcker (Graz, Austria) and Mike Wilson (NMW, Cardiff, UK). The bioacoustic facilities have been made available by Andrej Čokl and Meta Virant-Doberlet from the National Institute of Biology (Ljubljana, Slovenia) which is gratefully acknowledged. For the molecular work I want to thank Jakob Damgaard (Institute Of Biology, Copenhagen, Denmark), Alexander Szallies (Institute for Biogeography, Basel, Switzerland) and Georg Armbruster (Institute for Conservation Biology, Basel, Switzerland). I thank the late Roland Molenda (Institute for Biogeography, Basel, Switzerland) cordially to introduce me into the field of molecular zoogeography; it is a bereavement that he passed away so young. Daniel Mathys and Marcel Düggelein (ZMB – Zentrum für Mikroskopie, Basel, Switzerland) helped with the SEM pictures for which I am very grateful. Special thanks go to Pavel Lauterer, Heidi Günthart, Jakob Damgaard, Georg Armbruster, Roland Molenda, Alexander Szallies, Irena Dworakowska, Daniel Burckhardt, Herbert Nickel, Reinhard Remane, Igor Malenovský, Meta Virant-Doberlet, Valerio Mazzoni and Mike Wilson for their stimulating discussions on Hemiptera. For the wonderful time at the Museum and the Institute for Biogeography in Basel I thank following persons: Clarah Andriamalala, Wendy Altherr, Georg Armbruster, Ruth Bächli, Kathleen Beese, Angelo Bolzern, Michel Brancucci, Kevin Brown, Armin Coray, Michele Glasstetter, Beni Gnos, Ambros Hänggi, Sybille Hock, René Hoess, Doris Hölling, Gerhard Hotz, Andreas Kaupp, Lilith Köppen, Thibault Lachat, Henryk Luka, Christian A. Meyer, Ursula Meyer, Roland Molenda, Ursula Monzeglio, Renate Müller, Ralf Peveling, Kirsten Schütz, Eva Sprecher, Edi Stöckli, Alexander Szallies, Stefanie von Fumetti, Bea Watson, Urs Wüest, Denise Wyniger, Marc Zehntner, Samuel Zschokke, Isabelle Zürcher. SYNTHESYS (Synthesis of systematic resources) funded my visit for the molecular research at the Institute of Biology (Copenhagen) which is gratefully acknowledged. Finally I want to thank all my family and friends who continuously supported me during this period.

7 References

- Asche M, Hoch H. 2004. Fauna Europaea: Cicadomorpha. Fauna Europaea version 1.1, <http://www.faunaeur.org> (accession date: 10.07.2006)
- Blocker HD, Triplehorn BW. 1985. External Morphology of Leafhoppers. In: Nault LR, Rodriguez JG, eds. *The leafhoppers and planthoppers*. New York: Wiley-Interscience, 41-60.
- Bourgoin T, Campbell BC. 2002. Inferring a Phylogeny for Hemiptera: Falling into the 'Autapomorphic Trap'. *Denisia* **04**: 67-82.
- Burckhardt DH, Basset Y. 2000. The jumping plant-lice (Hemiptera, Psylloidea) associated with *Schinus* (Anacardiaceae): systematics, biogeography and host plant relationships. *Journal of Natural History* **34**: 57-155.
- Burckhardt D, Ouvrard D. 2007. The taxonomy, biogeography and host plant relationships of jumping plant-lice (Hemiptera: Psyllidae) associated with creosote bushes (*Larrea* spp., Zygophyllaceae). *Systematic Entomology* **32**: 136-155.
- Carver M, Gross GF, Woodward TE. 1991. Hemiptera. In: CSIRO, ed. *The Insects of Australia*. 2nd ed. Melbourne: Melbourne University Press. 429-509.
- Cerutti N. 1939. Les Typhlocybidae du Valais. *Bulletin de la Murithienne* **56**: 81-95.
- Claridge MF. 1985. Acoustic signals in the Homoptera: Behaviour, taxonomy and Evolution. *Annual Review of Entomology* **30**: 297-317.
- Claridge MF. 2006. Insect Sounds and Communication. In: Drosopoulos S, Claridge MF, eds. *Insect Sounds and Communication: physiology, behaviour, ecology and evolution*. Boca Raton: CRC Taylor & Francis, 3-10.
- Clary DO, Wolstenhome DR. 1985. The mitochondrial DNA molecule of *Drosophila yakuba*: Nucleotide sequence, gene organization, and genetic code. *Journal of Molecular Evolution* **22**: 252-271.
- Cryan JR. 2005. Molecular phylogeny of Cicadomorpha (Insecta: Hemiptera: Cicadoidea, Cercopoidea and Membracoidea): adding evidence to the controversy. *Systematic Entomology* **30**: 563-574.
- Dallwitz MJ. 1980. A general system for coding taxonomic descriptions. *Taxon* **29**: 41-46.
- Dallwitz MJ, Paine TA, Zurcher EJ. 1999. User's Guide to the DELTA Editor. <http://biodiversity.uno.edu/delta>.
- Davidson RH, DeLong DM. 1938. Studies of the genus *Empoasca* (Homoptera, Cicadellidae). Part V. Twelve new species of *Empoasca* from the United States. *Ohio Journal of Science* **38**: 90-96.
- Davis RB. 1975. Classification of Selected Higher Categories of Auchenorrhynchous Homoptera (Cicadellidae and Aetalionidae). *U.S. Department of Agriculture Technical Bulletin* **1494**: 1-52.
- della Giustina W. 1989. Homopteres Cicadellidae. Volume 3. Compléments aux ouvrages d'Henri Ribaut. *Faune De France* **73**: 1-350.
- DeLong DM. 1931. A revision of the American species of *Empoasca* known to occur north of Mexico. *U.S. Department of Agriculture Technical Bulletin* **231**: 1-59.
- DeLong DM. 1971. The bionomics of leafhoppers. *Annual Review of Entomology* **16**: 179-210.
- Dietrich CH. 2002. Evolution of Cicadomorpha (Insecta, Hemiptera). *Denisia* **04**: 155-170.
- Dietrich CH. 2005. Keys to the families of Cicadomorpha and subfamilies and tribes of Cicadellidae (Hemiptera: Auchenorrhyncha). *Florida Entomologist* **88**: 502-517.
- Dlabola J. 1958. Zikaden-Ausbeute vom Kaukasus. (Homoptera Auchenorrhyncha). *Acta Entomologica Musei Nationalis Pragae* **32**: 317-352.
- Dolling WR. 1991. *The Hemiptera*. London: Oxford University Press.

- Drosopoulos S. 2006.** Acoustic Communication, Mating Behaviour and Hybridisation of Planthoppers. In: Drosopoulos S, Claridge MF, eds. *Insect Sounds and Communication: physiology, behaviour, ecology and evolution*. Boca Raton: CRC Taylor & Francis, 239-253.
- Dumbleton LJ. 1964.** New records of Hemiptera-Homoptera and a key to the leafhoppers (Cicadellidae-Typhlocybinæ) in New Zealand. *New Zealand journal of science* **7**: 571-578.
- Dworakowska I. 1973.** On Some Palaearctic Species of the Genus *Kybos* Fieb. (Auchenorrhyncha, Cicadellidae, Typhlocybinæ). *Bulletin de l' Académie Polonaise des Sciences* **21**: 235-244.
- Dworakowska I. 1976.** *Kybos* Fieb., subgenus of *Empoasca* Walsh (Auchenorrhyncha, Cicadellidae, Typhlocybinæ) in Palaearctic. *Acta Zoologica Cracoviensia* **21**: 387-463.
- Edwards J. 1908.** On some British Homoptera hitherto undescribed or unrecorded. *Entomologist's Monthly Magazine* **19**: 80-87.
- Fallén CF. 1806.** Försök till de Svenska Cicad-arternas uppställning och beskrifning. *Nya Handlingar / Kongliga Svenska Vetenskaps-Akademiens* **27**: 6-43.
- Felsenstein J. 1985.** Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* **39**: 783-791.
- Fieber FX. 1866.** Neue Gattungen und Arten in Homopteren (Cicadina Bur.). *Verhandlungen der Zoologisch-Botanischen Gesellschaft in Wien* **16**: 497-516.
- Fletcher MJ, Knight WJ. 1998.** Conference Abstract. New Australian Records for Exotic Leafhoppers. 29th Annual General Meeting and Scientific Conference of the Australian Entomological Society, Brisbane 1998: 41.
- Futuyma DJ, Mitter C. 1996.** Insect-plant interactions: The evolution of component communities. *Philosophical Transactions of the Royal Society London, Series B* **351**: 1361-1366.
- Goloboff P. 1999.** NONA (NO NAME) ver. 2. Published by the author. Tucumán, Argentina.
- Grimaldi D, Engel MS. 2005.** *Evolution of Insects*. Cambridge: Cambridge University Press.
- Günthart H. 1974.** Beitrag zur Kenntnis der Kleinzikaden (Typhlocybinæ, Hom., Auch.) der Schweiz, 1. Ergänzung. *Mitteilungen der Schweizerischen Entomologischen Gesellschaft* **47**: 15-27.
- Günthart H. 1977.** Einfluss des Insektenalters auf Bestimmungsmerkmale. Biotaxonomische und rasterelektronenmikroskopische Untersuchungen bei Kleinzikaden (Hom. Auchenorrhyncha, Cicadellidae). *Mitteilungen der Schweizerischen Entomologischen Gesellschaft* **50**: 189-201.
- Hamilton KGA. 1983.** Introduced and native leafhoppers common to the Old and New Worlds (Rhynchota: Homoptera: Cicadellidae). *Canadian Entomologist* **115**: 473-511.
- Hebert PD, Cywinska A, Ball SL, deWaard JR. 2003.** Biological identifications through DNA barcodes. *Proceedings of the Royal Entomological Society, Series B* **270**: 313-321.
- Hickerson MJ, Meyer CP, Craig M. 2006.** DNA barcoding will often fail to discover new animal species over broad parameter space. *Systematic Biology* **55**: 729-739.
- Hodkinson ID, Casson D. 1991.** A lesser predilection for bugs: Hemiptera (Insecta) diversity in tropical rain forest. *Biological Journal of the Linnean Society* **43**: 101-109.
- Horváth G. 1897.** Homoptera nova ex Hungarica. *Természetrázi füzetek* **20**: 620-648.
- Jaenike J. 1990.** Host specialization in phytophagous insects. *Annual Review of Entomology* **21**: 243-273.
- Knight WJ. 1976.** Typhlocybinæ of New Zealand (Homoptera: Cicadellidae). *New Zealand journal of zoology* **3**: 71-87.

- Kristensen NP. 1991.** Phylogeny of Extant Hexapods. In: CSIRO, ed. *The Insects of Australia*. 2nd ed. Melbourne: Melbourne University Press, 125-140.
- Le Quesne WJ, Payne KR. 1981.** Cicadellidae (Typhlocybinae) with a checklist of the British Auchenorrhyncha (Hemiptera, Homoptera). *Handbooks for the Identification of British Insects II (2c)*: 1-95.
- Lindberg H. 1952.** *Empoasca borealis* n. sp. und *Boreotettix* (n. gen.) *serricauda* (Kontk.) (Homoptera, Cicadidae) aus Nordfinnland. *Notulae entomologicae* **32**: 144-147.
- Linnavuori R. 1949.** Hemipterologisches aus Finnland. *Annales Entomologica Fennica* **15**: 145-156.
- Linnavuori R. 1951.** Hemipteral observations. *Annales Entomologica Fennica* **17**: 51-64.
- Linnavuori R. 1953.** On some new or interesting leafhopper species of the family Cicadellidae. *Annales entomologici Fennici* **19**: 56-63.
- Medrano JF, Asen E, Sparrow L. 1990.** DNA extraction from nucleated red blood cells. *Biotechniques* **8**: 43.
- Meier R, Shiyang K, Vaidya G, K. L. Ng P. 2006.** DNA Barcoding and Taxonomy in Diptera: A Tale of High Intraspecific Variability and Low Identification Success. *Systematic Biology* **55**: 715-728.
- Melichar L. 1896.** Einige neue Homoptera-Arten und Varietäten. *Verhandlungen der Zoologisch-Botanischen Gesellschaft in Wien* **46**: 176-180.
- Metcalf ZP. 1968.** *General Catalogue of the Homoptera. Fascicle VI. Cicadelloidea. Part 17. Cicadellidae*. Washington: U.S. Department of Agriculture, Agriculture Research Service.
- Mitjaev ID. 1968.** Keskol'ko Novych Vidov Zikad (Homoptera, Cicadellidae) iz Jugo-Sapadnogo Altaja i Sajsanskoj Kotloviny - Several new species of leafhoppers (Homoptera, Cicadellidae) from South-Western Altai and Zaisan Hollow. *Zoologiceskij zurnal* **47**: 635-637.
- Nast J. 1972.** *Palaeartic Auchenorrhyncha (Homoptera): An annotated check list*. Warszawa: Polish Scientific Publ.
- Nei M, Kumar S. 2000.** *Molecular Evolution and Phylogenetics*. New York: Oxford University Press.
- Nickel H. 2003.** *The leafhoppers and planthoppers of Germany (Hemiptera, Auchenorrhyncha): patterns and strategies in a highly diverse group of phytophagous insects*. Sofia and Moscow: Pensoft.
- Nielson MW. 1985.** Leafhopper systematics. In: Nault LR, Rodriguez JG, eds. *The leafhoppers and planthoppers*. New York: Wiley-Interscience, 11-39.
- Nixon KC. 2002.** WinClada ver. 1.00.08. Published by the author. Ithaca, USA.
- Orosz A. 1996.** Data to the Knowledge of the Cicadomorpha Species (Homoptera: Auchenorrhyncha) in the Bükk National Park. *The Fauna of the Bükk National Park*: 151-157.
- Ossiannilsson F. 1941.** A New Swedish Species of *Empoasca* (Hom.). *Empoasca strigilifera* n. sp. *Entomologisk Tidskrift* **62**: 198-199.
- Ossiannilsson F. 1942.** Contributions to the knowledge of Swedish Cicadina: With description of a new species. *Opuscula entomologica* **6**: 113-114.
- Ossiannilsson F. 1946.** On the sound-production and the sound-producing organ in Swedish Homoptera Auchenorrhyncha. *Opuscula Entomologica* **11**: 82-84.
- Ossiannilsson F. 1949.** Insect Drummers, A study on the morphology and function of the sound-producing organ of Swedish Homoptera Auchenorrhyncha. *Opuscula Entomologica Suppl. X*: 1-145.
- Ossiannilsson F. 1955.** Några för Sverige nya stritar (Homoptera, Auchenorrhyncha): Med en synonymisk anmärkning - A few leafhoppers (Homoptera, Auchenorrhyncha) new to Sweden with a synonymic note. *Entomologisk tidskrift* **76**: 131-133.

- Ossiannilsson F. 1978.** The Auchenorrhyncha (Homoptera) of Fennoscandia and Denmark. Part 1: Introduction, infraorder Fulgoromorpha. *Fauna Entmologica Scandinavica* **7**: 1-222.
- Ossiannilsson F. 1981.** The Auchenorrhyncha (Homoptera) of Fennoscandia and Denmark. Part 2: The Families Cicadidae, Cercopidae, Membracidae, and Cicadellidae (excl. Deltocephalinae). *Fauna Entmologica Scandinavica* **7**: 223-593.
- Percy D. 2003.** Radiation, diversity and host-plant interactions among island and continental legume-feeding psyllids. *Evolution* **57**: 2540-2556.
- Price PW. 1997.** *Insect Ecology*. 3rd edition. New York: John Wiley.
- Ribaut H. 1933.** Sur quelques Espèces du genre *Empoasca* (Homoptera-Typhlocybidae). *Bulletin de la Société d'Histoire Naturelle de Toulouse* **65**: 150-161.
- Ribaut H. 1936.** Homoptères Auchenorhynques (I. Typhlocybidae). *Faune de France* **31**: 1-231.
- Ribaut H. 1952.** Homoptères Auchenorhynques. II (Jassidae). *Faune de France* **57**: 1-474.
- Ross HH. 1963.** An Evolutionary Outline of the Leafhopper Genus *Empoasca* Subgenus *Kybos*, with a Key to the Nearctic Fauna (Hemiptera, Cicadellidae). *Annals of the Entomological Society of America* **56**: 202-223.
- Shaw KC, Vargo A, Carlson OV. 1974.** Sounds and associated behavior of some species of *Empoasca* (Homoptera: Cicadellidae). *Journal of the Kansas Entomological Society* **47**: 284-307.
- Sorensen JT, Campbell BC, Gill RJ, Steffen-Campbell JD. 1995.** Non-monophyly of Auchenorrhyncha ("Homoptera"), based upon 18S rDNA phylogeny: eco-evolutionary and cladistic implications within pre-heteropterodea Hemiptera (s.l.) and a proposal for new monophyletic suborders. *Pan-Pacific Entomologist* **71**: 31-60.
- Sperling F. 2003.** DNA barcoding. Deus et machina [opinion page]. *Newsletter of the Biological Survey of Canada (Terrestrial Arthropods)* **22**: 50-53.
- Swofford DL. 1998.** *PAUP*. Phylogenetic Analysis Using Parsimony (* and Other Methods)*, Version 4. Sunderland: Sinauer Associates.
- Tautz D, Arctander P, Minelli A, Thomas RH, Vogler AP. 2003.** A plea for DNA taxonomy. *Trends in Ecology and Evolution* **18**: 70-74.
- Thompson JD, Higgins DG, Gibson TJ. 1994.** CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positions-specific gap penalties and weight matrix choice. *Nucleic Acids Research* **22**: 4673-4680.
- Vilbaste J. 1961.** Novye Zikadovye (Homoptera: Cicadina) iz Okrestnostej Astraschani - Neue Zikaden (Homoptera, Cicadina) aus der Umgebung von Astrachan. *Eesti NSV Teaduste Akadeemia toimetised, Bioloogiline seeria* **10**: 315-331.
- Wagner W. 1935.** Die Zikaden der Nordmark und Nordwest-Deutschlands. *Verhandlungen des Vereins für naturwissenschaftliche Heimatforschung zu Hamburg* **24**: 1-44.
- Wagner W. 1949.** Drei neue Typhlocybiden aus Steiermark. *Zentralblatt für das Gesamtgebiet der Entomologie* **3**: 43-45.
- Wagner W. 1955.** Neue mitteleuropäische Zikaden und Blattflöhe. *Entomologische Mitteilungen aus dem Zoologischen Staatsinstitut und Zoologischen Museum Hamburg* **6**: 160-194.
- Wagner W. 1959.** Über neue und schon bekannte Zikadenarten aus Italien (Hemiptera-Homoptera). *Fragmenta Entomologica* **III**: 67-86.
- Walsh BD. 1862.** Fire blight: Two new foes of the apple and pear. *Prairie farmer* **10**: 147-149.
- Wheeler QD. 2005.** Letter to the editor - Losing the plot: DNA "barcodes" and taxonomy. *Cladistics* **21**: 405-407.

- Yoshizawa K, Saigusa T. 2001.** Phylogenetic analysis of paraneopteran orders (Insecta: Neoptera) based on forewing base structure, with comments on monophyly of Auchenorrhyncha (Hemiptera). *Systematic Entomology* **26**: 1-13.
- Zachvatkin AA. 1953a.** Biologo - sistematitscheskie zametki tsikadinakh sred ne-russkoy fauny - Biological and systematic notes on Cicadina fauna of central Russia. In: Smirnov ES (ed.). *Sbornik nauchnykh rabot*. Moscow: Izdatel'stvo, 205-209.
- Zachvatkin AA. 1953b.** K faune Eupterygidae (Homoptera, Cicadina) Sredney Azii (Empoascinae) - Faunistic notes on Eupterygidae (Homoptera, Cicadina) from Central Asia. Empoascinae. In: Smirnov ES (ed.). *Sbornik nauchnykh rabot*. Moscow: Izdatel'stvo, 237-245.

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos abstrusus</i>	1♂	Bulgaria	19.07.1971	P. Lauterer	MZMB	
<i>Kybos abstrusus</i>	1♂	Czech Republic, Mor. Budejovice	23.06.1981	P. Lauterer	MZMB	
<i>Kybos abstrusus</i>	1♀	Finland, Turku	28.08.1948	R. Linnavuori	AMNH	
<i>Kybos abstrusus</i>	1♂	France, Strasbourg	13.09.1993	I. Malenovský	MZMB	
<i>Kybos abstrusus</i>	1♂	Slovakia, Palúzdka	15.07.1963	P. Lauterer	MZMB	
<i>Kybos abstrusus</i>	9♂ 5♀	Slovakia, Palúdzká	15.07.1963	P. Lauterer	MZMB	
<i>Kybos abstrusus</i>	8♂ 5♀	Slovakia, Palúdzká	15.07.1963	P. Lauterer	MZMB	
<i>Kybos abstrusus</i>	1♂	Slovakia, Oslany	10.07.1977	P. Lauterer	MZMB	
<i>Kybos aenicola</i>	2♂ 6♀ 1 nymph	Italy, Mt. Etna	30.08.1948	Hartig	INER	type series
<i>Kybos austriacus</i>	1♂	Slovakia, Betliar	11.07.1979	P. Lauterer	MZMB	
<i>Kybos austriacus</i>	1♂	Switzerland, Zernež	26.08.1977	H. Günthart	Coll. Günthart	
<i>Kybos austriacus</i>	1♂	Switzerland, Mte. Brè	03.08.1983	L. Rezbanyai	NHMB	
<i>Kybos austriacus</i>	1♂	Switzerland, Zernež	18.08.1987	H. Günthart	Coll. Günthart	
<i>Kybos austriacus</i>	2♂	Switzerland, Zernež	21.08.1987	H. Günthart	Coll. Günthart	
<i>Kybos austriacus</i>	1♀	Switzerland, Zernež	21.08.1987	H. Günthart	Coll. Günthart	
<i>Kybos butleri</i>	3♂ 5♀	Austria, Oberrakitsch	21.09.1996	P. Lauterer	MZMB	
<i>Kybos butleri</i>	1♂	Bulgaria	20.07.1971	P. Lauterer	MZMB	
<i>Kybos butleri</i>	1♂	Bulgaria	27.07.1972	P. Lauterer	MZMB	
<i>Kybos butleri</i>	1♂	Czech Republic, Sakvice	27.08.1973	P. Lauterer	MZMB	
<i>Kybos butleri</i>	5♂	Czech Republic, Moravia mer.	19.06.1980	P. Lauterer	MZMB	
<i>Kybos butleri</i>	1♂	Czech Republic, Chriby	21.08.1980	P. Lauterer	MZMB	
<i>Kybos butleri</i>	1♂	Czech Republic, Moravia centr.	21.08.1980	P. Lauterer	MZMB	
<i>Kybos butleri</i>	3♂	Czech Republic, Mor. Budejovice	23.06.1981	P. Lauterer	MZMB	
<i>Kybos butleri</i>	2♂ 1♀	Czech Republic, Bílé Karpaty	09.09.1998	I. Malenovský	MZMB	
<i>Kybos butleri</i>	3♀	Czech Republic, Bílé Karpaty	09.09.2000	I. Malenovský	MZMB	
<i>Kybos butleri</i>	1♂	Germany, Weil am Rhein	27.07.2000	R. Mühlethaler	NHMB	
<i>Kybos butleri</i>	1♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	
<i>Kybos butleri</i>	1♂	Switzerland, Basel	22.06.2000	R. Mühlethaler	NHMB	
<i>Kybos calyculus</i>	1♂	Czech Republic, Charvatská	31.07.1974	P. Lauterer	MZMB	
<i>Kybos calyculus</i>	3♂	Czech Republic, Brno-Slatina	21.06.2003	R. Mühlethaler	NHMB	
<i>Kybos candelabricus</i>	2♂ 3♀	Turkey, Topasulag	16.06.1970	J. Diabola	NMP	
<i>Kybos candelabricus</i>	1♂ 5♀	Turkey, Ankara	08.-10.8.1963	R. Linnavuori	NMP	
<i>Kybos digitatus</i>	2♂	France, Rougon	26.08.1996	I. Malenovský	MZMB	
<i>Kybos digitatus</i>	1♂	Germany, Weil am Rhein	27.07.2000	R. Mühlethaler	NHMB	
<i>Kybos digitatus</i>	1♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler	NHMB	
<i>Kybos digitatus</i>	9♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler	NHMB	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos digitatus</i>	1 ♀	Germany, Weil am Rhein	19.08.2002	R. Mühlethaler	NHMB	
<i>Kybos digitatus</i>	1 ♀	Germany, Weil am Rhein	11.06.2003	R. Mühlethaler	NHMB	
<i>Kybos digitatus</i>	2 ♀	Germany, Weil am Rhein	23.07.2004	R. Mühlethaler	NHMB	
<i>Kybos digitatus</i>	5 ♂ 3 ♀	Germany, Weil am Rhein	29.07.2004	R. Mühlethaler	NHMB	
<i>Kybos digitatus</i>	1 ♂	Switzerland, Basel	19.08.2001	R. Mühlethaler	NHMB	
<i>Kybos limpidus</i>	1 ♂	Bulgaria	27.07.1972	P. Lauterer	MZMB	
<i>Kybos limpidus</i>	1 ♂ 3 ♀	Czech Republic, Sakvice	27.08.1973	P. Lauterer	MZMB	
<i>Kybos limpidus</i>	3 ♂ 4 ♀	Germany, Berlin-Hermsdorf	10.08.2002	R. Mühlethaler	NHMB	
<i>Kybos limpidus</i>	4 nymphs	Poland, Warsaw	24.08.1973	I. Dworakowska	AMNH	
<i>Kybos limpidus</i>	1 ♀	Poland, Warsaw	29.08.1973	I. Dworakowska	Coll. Günthart	
<i>Kybos limpidus</i>	2 ♂ 5 ♀	Poland, Warsaw	29.08.1973	I. Dworakowska	AMNH	
<i>Kybos lindbergi</i>	1 ♂	Bulgaria	27.07.1972	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Czech Republic, Brno-Staró	20.09.1965	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	5 ♂	Czech Republic, Jeseník-Vel.	20.09.1970	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Czech Republic, Moravia mer.	14.08.1975	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Czech Republic, Moravia occ.	16.06.1977	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	3 ♂ 4 ♀	Czech Republic, Moravia occ.	05.09.1978	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	3 ♀	Czech Republic, Ridelov	05.09.1978	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	4 ♂ 8 ♀	Czech Republic, Moravia mer.	13.09.1978	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Czech Republic, Moravia occ.	30.08.1979	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Czech Republic, Moravia occ.	30.08.1979	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Czech Republic, Moravia centr.	17.06.1981	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	5 ♂	Czech Republic, Moravia or.	24.06.1982	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	2 ♂ 2 ♀	Czech Republic, Brno-Mokrá	18.06.1994	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	8 ♀ 1 nymph	Czech Republic, Hodonín-Pánsv	17.08.1995	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	3 ♀	Czech Republic, Bílé Karpáty	14.09.1999	I. Malenovský	MZMB	
<i>Kybos lindbergi</i>	20 ♂	Czech Republic, Brno-Slatina	21.06.2003	R. Mühlethaler	NHMB	
<i>Kybos lindbergi</i>	2 ♂ 1 ♀	Finland, Turku	07.07.1974	H. Günthart	Coll. Günthart	
<i>Kybos lindbergi</i>	1 ♂ 1 ♀	Poland, Kosobudy	12.07.1966	I. Dworakowska	AMNH	
<i>Kybos lindbergi</i>	5 ♂ 15 ♀	Slovakia	11.07.1979	P. Lauterer	MZMB	
<i>Kybos lindbergi</i>	1 ♂	Switzerland, Riehen	26.05.2000	D. Wyniger	NHMB	
<i>Kybos lindbergi</i>	1 ♂	Switzerland, Dielsdorf	19.07.2002	R. Mühlethaler	NHMB	
<i>Kybos lindbergi</i>	1 ♂	Switzerland, Basel	20.07.2002	R. Mühlethaler	NHMB	
<i>Kybos ludus</i>	1 ♂	USA, Wooster, Ohio	1917	D.M. DeLong	OSU	holotype
<i>Kybos ludus</i>	1 ♂	Bulgaria	27.07.1972	P. Lauterer	MZMB	
<i>Kybos ludus</i>	2 ♂	Switzerland, Uster	25.07.1977	H. Günthart	Coll. Günthart	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos ludus</i>	1♂	Switzerland, Zernež	26.08.1977	H. Günthart	Coll. Günthart	
<i>Kybos ludus</i>	1♂	Switzerland, Mte. Brè	03.08.1983	L. Rezbanyai	NHMB	
<i>Kybos ludus</i>	1♂	Switzerland, Basel	20.07.2002	R. Mühlethaler	NHMB	
<i>Kybos mesasiaticus</i>	1♀	Afghanistan, Bala Murghab	02.07.1964	J. Diabola	NMP	
<i>Kybos mesasiaticus</i>	1♂	Afghanistan, Bala Murghab	10.-13.07.1964	J. Diabola	NMP	
<i>Kybos mesasiaticus</i>	2♂ 7♀	Mongolia, Songino	03.09.1966	J. Diabola	NMP	
<i>Kybos mucronatus</i>	1♂	Bulgaria	13.08.1972	P. Lauterer	MZMB	
<i>Kybos mucronatus</i>	1♂	Bulgaria	16.08.1972	P. Lauterer	MZMB	
<i>Kybos mucronatus</i>	1♂	Czech Republic, Bohemia mer.	04.08.1973	P. Lauterer	MZMB	
<i>Kybos mucronatus</i>	1♂	Czech Republic, Moravia occ.	02.08.1984	P. Lauterer	MZMB	
<i>Kybos mucronatus</i>	1♂	France, Offendorf	10.07.1995	I. Malenovský	MZMB	
<i>Kybos mucronatus</i>	1♂	Poland, Karwia	25.08.1958	R. Bielawski	AMNH	"verbae"
<i>Kybos mucronatus</i>	1♂	Poland, Kazimierz	14.07.1967	I. Dworakowska	AMNH	"verbae"
<i>Kybos mucronatus</i>	2♂	Switzerland, Zernež	04.08.1973	H. Günthart	Coll. Günthart	
<i>Kybos mucronatus</i>	1♀	Switzerland, Zernež	04.08.1975	H. Günthart	Coll. Günthart	
<i>Kybos mucronatus</i>	1♂	Switzerland, Zernež	19.07.1990	H. Günthart	Coll. Günthart	On <i>Salix</i>
<i>Kybos mucronatus</i>	1♂	Switzerland, Zernež	19.07.1990	H. Günthart	Coll. Günthart	
<i>Kybos mucronatus</i>	1♂	Switzerland, Bever	09.09.2002	R. Mühlethaler	NHMB	"verbae"
<i>Kybos mucronatus</i>	2♂	Switzerland, Zuoz	?	H. Günthart	Coll. Günthart	"verbae"
<i>Kybos populi</i>	2♂	Bulgaria	20.07.1971	P. Lauterer	MZMB	
<i>Kybos populi</i>	1♂	Bulgaria, Harmanli	20.07.1971	P. Lauterer	MZMB	
<i>Kybos populi</i>	1♂	Czech Republic, Bohemia mer.	08.08.1973	P. Lauterer	MZMB	
<i>Kybos populi</i>	2♂ 2♀	Czech Republic, Moravia occ.	14.06.1977	P. Lauterer	MZMB	
<i>Kybos populi</i>	1♂	Czech Republic, Moravia occ.	14.06.1977	P. Lauterer	MZMB	
<i>Kybos populi</i>	2♀	Czech Republic, Nemcicky	14.06.1977	P. Lauterer	MZMB	
<i>Kybos populi</i>	1♂	Czech Republic, Moravia occ.	30.08.1979	P. Lauterer	MZMB	
<i>Kybos populi</i>	1♂	Czech Republic, Moravia centr.	10.08.1983	P. Lauterer	MZMB	
<i>Kybos populi</i>	5♀	Czech Republic, Brno-Mokrá	18.06.1994	P. Lauterer	MZMB	
<i>Kybos populi</i>	2♂	Czech Republic, Mikulov	27.07.2001	P. Lauterer	MZMB	
<i>Kybos populi</i>	9♂ 7♀ 1 nymph	Czech Republic, Mikulov	27.07.2001	P. Lauterer	MZMB	
<i>Kybos populi</i>	6♀	Czech Republic, Bílé Karpaty	20.09.2001	I. Malenovský & P. Kment	MZMB	
<i>Kybos populi</i>	1♀	Czech Republic, Brno	21.06.2003	R. Mühlethaler	NHMB	
<i>Kybos populi</i>	1♀	France, Strasbourg	22.07.1993	I. Malenovský	MZMB	
<i>Kybos populi</i>	2♂ 1♀	France, Strasbourg	05.08.1993	I. Malenovský	MZMB	
<i>Kybos populi</i>	1♀	France, Strasbourg	01.09.1993	I. Malenovský	MZMB	
<i>Kybos populi</i>	2♂	France, Strasbourg	13.09.1993	I. Malenovský	MZMB	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos populi</i>	3 ♀	France, Strasbourg	13.09.1993	I. Malenovský	MZMB	
<i>Kybos populi</i>	1 ♀	France, Strasbourg	15.09.1994	I. Malenovský	MZMB	
<i>Kybos populi</i>	1 ♀	France, Strasbourg	18.09.1994	I. Malenovský	MZMB	
<i>Kybos populi</i>	4 ♀	Slovakia, Roháče-Tatliakova	14.08.1959	P. Lauterer	MZMB	
<i>Kybos populi</i>	14 ♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	
<i>Kybos populi</i>	1 ♂	Switzerland, Birsfelden	30.08.1972	H. Günthart	Coll. Günthart	
<i>Kybos populi</i>	1 ♂	Switzerland, Morel	28.08.1986	H. Günthart	Coll. Günthart	
<i>Kybos populi</i>	1 ♂	Switzerland, Basel	19.08.2001	R. Mühlethaler	NHMB	
<i>Kybos populi</i>	1 ♂	Switzerland, Samedan	06.09.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	2 ♂	Bulgaria	19.07.1971	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Bulgaria	20.07.1971	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	3 ♂	Czech Republic, Bohemia	10.08.1973	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Czech Republic, Moravia mer.	19.06.1980	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Czech Republic, Moravia centr.	25.08.1980	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	8 ♂	Czech Republic, Moravia mer.	03.06.1981	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	16 ♂ 5 ♀	Czech Republic, Mor. Budejovice	23.06.1981	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Czech Republic, Moravia or.	23.06.1983	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Czech Republic, Moravia or.	20.08.1984	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Czech Republic, Bílé Karpaty	17.06.2000	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♂	Finland, Turku	21.07.1947	R. Linnavuori	AMNH	
<i>Kybos rufescens</i>	1 ♂	Finland, Turku	18.07.1948	R. Linnavuori	AMNH	
<i>Kybos rufescens</i>	1 ♂	France, Strasbourg	07.09.2000	I. Malenovský	MZMB	
<i>Kybos rufescens</i>	2 ♂	Germany, Weil am Rhein	27.07.2000	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♂	Germany, Weil am Rhein	05.10.2000	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	130 ♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler & P. Lauterer	NHMB	
<i>Kybos rufescens</i>	1 ♂	Germany, Weil am Rhein	23.07.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	4 ♂	Germany, Weil am Rhein	19.08.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	4 ♂	Germany, Weil am Rhein	03.06.2003	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	3 ♂	Germany, Weil am Rhein	11.06.2003	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	2 ♂ 7 ♀	Greece	04.06.1995	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	1 ♂	Greece	12.06.1995	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	2 ♂ 2 ♀	Poland, Komic	16.06.1966	I. Dworakowska	AMNH	
<i>Kybos rufescens</i>	2 ♂ 2 ♀	Polonia	23.06.1971	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	4 ♂ 5 ♀	Slovakia, Palúdzka	15.07.1963	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	7 ♂	Slovakia, Vazec u Strby	16.07.1963	P. Lauterer	MZMB	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos rufescens</i>	4♀	Slovakia, Vazec	16.07.1963	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	4♂ 13♀	Slovakia, Vel. Fatra-Blatnica	10.07.1976	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	7♂ 4♀	Slovakia	10.07.1976	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	4♂	Slovakia	11.07.1976	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	6♂ 26♀	Slovakia	07.07.1979	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	9♂	Slovakia	15.07.1979	P. Lauterer	MZMB	
<i>Kybos rufescens</i>	9♂	Switzerland, Basel	13.07.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1♂	Switzerland, Basel	19.08.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	2♂	Switzerland, Samedan	04.09.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1♂	Switzerland, Samedan	09.09.2002	R. Mühlethaler	NHMB	
<i>Kybos rufescens</i>	2♂ 36♀	Switzerland, Kaisten	21.06.2003	D. Wyniger	NHMB	
<i>Kybos smaragdula</i>	1♂	Bulgaria	08.08.1972	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Bohemia mer.	04.08.1973	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Bohemia mer.	08.08.1973	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	2♂	Czech Republic, Moravia occ.	22.06.1976	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Bohemia mer.	01.10.1980	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	7♂	Czech Republic, Mor. Budejovice	23.06.1981	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	3♂	Czech Republic, Moravia or.	24.06.1982	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Brno-Staró	20.07.1982	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Moravia occ.	02.08.1984	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Moravia occ.	15.08.1984	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♀	Czech Republic, Doubravice-Klemov	07.07.1987	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	5♀	Czech Republic, Brnenec	25.06.1999	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	10♀	Czech Republic, Brno-Utechov	26.06.1999	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Bílé Karpaty	18.06.2000	I. Malenovský	MZMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Bílé Karpaty	09.09.2000	I. Malenovský	MZMB	
<i>Kybos smaragdula</i>	1♂	Germany, Oberlausnitz	29.08.1998	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	3♂	Polonia	23.06.1971	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	4♂ 6♀	Romania	29.07.1971	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	4♂	Romania, Munti Bucegi	29.07.1971	P. Lauterer	MZMB	
<i>Kybos smaragdula</i>	1♀	Switzerland, Birsfelden	06.09.1971	H. Günthart	Coll. Günthart	
<i>Kybos smaragdula</i>	1♂	Switzerland, Basel	24.05.2000	R. Mühlethaler	NHMB	On Betula
<i>Kybos smaragdula</i>	1♂	Switzerland, Riehen	14.08.2000	D. Wyniger	NHMB	
<i>Kybos smaragdula</i>	2♀	Switzerland, Wildenstein	27.06.2002	R. Mühlethaler	NHMB	
<i>Kybos sordidulus</i>	1♂	Finland, Turku	28.07.1946	R. Linnavuori	AMNH	
<i>Kybos sordidulus</i>	2♂	Russia, Moscow	01.06.1963	J. Diabola	NMP	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos strigilifer</i>	1♂	Czech Republic, Moravia occ.	14.06.1977	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Moravia occ.	30.08.1979	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Moravia occ.	02.07.1980	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Mor. Budejovice	23.06.1981	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Moravia occ.	14.06.1983	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Moravia occ.	02.08.1984	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	5♀	Czech Republic, Vranov u Brno	06.06.1993	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♀	Czech Republic, Silesia; Uhelná	19.06.1993	H. Günthart	Coll. Günthart	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Silesia; Uhelná	21.06.1993	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Bílé Karpaty	09.09.1998	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Bílé Karpaty	17.06.2000	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	1♂	Czech Republic, Brno-Reckovice	22.06.2003	R. Mühlethaler	NHMB	
<i>Kybos strigilifer</i>	3♂	Finland, Turku	27.06.1948	R. Linnavuori	AMNH	
<i>Kybos strigilifer</i>	1♂	France, Strasbourg	15.09.1994	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	2♂	France, La Wantzenau	10.07.1995	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	1♂	France, Breitenbach	07.07.1996	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	1♂	France, Strasbourg	06.09.1996	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	2♂	France, Strasbourg	07.09.1996	I. Malenovský	MZMB	
<i>Kybos strigilifer</i>	1♂	Germany, Weil am Rhein	27.07.2000	R. Mühlethaler	NHMB	
<i>Kybos strigilifer</i>	1♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler	NHMB	
<i>Kybos strigilifer</i>	1♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler	NHMB	
<i>Kybos strigilifer</i>	1♂	Germany, Weil am Rhein	09.07.2003	R. Mühlethaler	NHMB	
<i>Kybos strigilifer</i>	1♂	Germany, Weil am Rhein	29.07.2004	R. Mühlethaler	NHMB	
<i>Kybos strigilifer</i>	1♂ 1♀	Poland, Tomaszon	09.09.1967	I. Dworakowska	MZMB	
<i>Kybos strigilifer</i>	7♂ 3♀	Slovakia, Belanské Tatry	07.08.1967	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Slovakia	11.07.1976	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	1♂	Slovakia	15.07.1979	P. Lauterer	MZMB	
<i>Kybos strigilifer</i>	2♂	Slovenia, Tolmin	24.07.2001	H. Löcker	Coll. H. Löcker	
<i>Kybos strigilifer</i>	1♀	Switzerland, Meilen	13.09.1972	H. Günthart	Coll. Günthart	
<i>Kybos strigilifer</i>	1♂	Switzerland, Basel	25.08.2000	R. Mühlethaler	NHMB	
<i>Kybos strobli</i>	1♂	Czech Republic, Pouzdrany-luzní	10.06.1974	P. Lauterer	MZMB	
<i>Kybos strobli</i>	1♂	Czech Republic, Pouzdrany-luzní	10.06.1974	P. Lauterer	MZMB	
<i>Kybos strobli</i>	3♂	Czech Republic, Horní Nemci	10.09.1998	I. Malenovský	MZMB	
<i>Kybos strobli</i>	1♀	Poland, Pieminy Mts.	05.07.1973	I. Dworakowska	MZMB	
<i>Kybos strobli</i>	1♂ 1♀	Poland, Pieniny Mts.	05.07.1975	I. Dworakowska	AMNH	
<i>Kybos strobli</i>	10♂	Romania	29.07.1971	P. Lauterer	MZMB	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos strobli</i>	9♂	Romania, Munti Bucegi	29.07.1971	P. Lauterer	MZMB	
<i>Kybos strobli</i>	1♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	
<i>Kybos strobli</i>	4♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	
<i>Kybos strobli</i>	1♂	Switzerland, Genève	10.09.1919	?	MHNG	
<i>Kybos strobli</i>	2♂	Switzerland, Untervaz	16.06.1974	H. Günthart	Coll. Günthart	
<i>Kybos strobli</i>	1♂	Switzerland, Sta. Maria	02.07.1982	H. Günthart	Coll. Günthart	
<i>Kybos strobli</i>	1♂	Switzerland, Zernezh	03.07.1982	H. Günthart	Coll. Günthart	
<i>Kybos strobli</i>	1♀	Switzerland, Zuoz	1987?	H. Günthart	Coll. Günthart	
<i>Kybos virgator</i>	10♂	Bulgaria	19.07.1971	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	25♂	Bulgaria	20.07.1971	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Bulgaria	27.07.1972	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Bulgaria	08.08.1972	P. Lauterer	MZMB	
<i>Kybos virgator</i>	3♂ 6♀	Bulgaria	09.08.1972	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Bulgaria	13.08.1972	P. Lauterer	MZMB	
<i>Kybos virgator</i>	1♂	Bulgaria	13.08.1972	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Bulgaria	16.08.1972	P. Lauterer	MZMB	
<i>Kybos virgator</i>	1♂ 3♀	Bulgaria	19.08.1972	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	3♂	Czech Republic, Brno-Holásky	02.08.1957	P. Lauterer	MZMB	
<i>Kybos virgator</i>	3♂	Czech Republic, Brno-Holásky	02.08.1957	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	2♂ 2♀	Czech Republic, Belkvice u Olomouce	20.08.1961	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	2♂	Czech Republic, Moravia mer.	06.06.1973	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia mer.	14.08.1973	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia mer.	08.10.1973	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Pasohlávky	06.06.1974	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	2♂	Czech Republic, Pouzdrany-luzní	10.06.1974	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia mer.	19.06.1975	P. Lauterer	MZMB	
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia occ.	22.06.1976	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia mer.	24.06.1976	P. Lauterer	MZMB	
<i>Kybos virgator</i>	2♂	Czech Republic, Moravia mer.	24.06.1976	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia occ.	16.06.1977	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia mer.	03.06.1981	P. Lauterer	MZMB	
<i>Kybos virgator</i>	1♂	Czech Republic, Mor. Budejovice	23.06.1981	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	1♂ 1♀	Czech Republic, Moravia mer.	06.10.1981	P. Lauterer	MZMB	
<i>Kybos virgator</i>	1♂	Czech Republic, Moravia or.	23.06.1983	P. Lauterer	MZMB	
<i>Kybos virgator</i>	2♂	Czech Republic, Ketkovice	30.05.1993	I. Malenovský	MZMB	
<i>Kybos virgator</i>	1♂ 1♀	Czech Republic, Ketkovice	30.05.1993	I. Malenovský	MZMB	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos virgator</i>	2♀	Czech Republic, Zblovce	12.06.1993	I. Malenovský	MZMB	
<i>Kybos virgator</i>	2♂	Czech Republic, Horní Nemci	10.09.1998	I. Malenovský	MZMB	
<i>Kybos virgator</i>	1♂	Czech Republic, Bílé Karpaty	18.06.2000	I. Malenovský	MZMB	
<i>Kybos virgator</i>	2♂	Czech Republic, Bílé Karpaty	09.09.2000	I. Malenovský	MZMB	
<i>Kybos virgator</i>	3♂	Finland, Turku	20.07.1948	R. Linnavuori	AMNH	
<i>Kybos virgator</i>	1♂	France, Strasbourg	05.08.1993	I. Malenovský	MZMB	
<i>Kybos virgator</i>	1♂	France, Strasbourg	01.09.1993	I. Malenovský	MZMB	
<i>Kybos virgator</i>	2♂	France, Strasbourg	17.07.1994	I. Malenovský	MZMB	
<i>Kybos virgator</i>	6♂	Germany, Weil am Rhein	31.05.2002	R. Mühlethaler	NHMB	
<i>Kybos virgator</i>	1♂	Germany, Weil am Rhein	19.08.2002	R. Mühlethaler	NHMB	
<i>Kybos virgator</i>	7♂ 11♀	Greece	05.05.1995	P. Lauterer	MZMB	
<i>Kybos virgator</i>	2♂	Poland, Ruda	16.08.1974	I. Dworakowska	AMNH	
<i>Kybos virgator</i>	4♂	Polonia	23.06.1971	P. Lauterer	MZMB	
<i>Kybos virgator</i>	1♂	Slovakia	04.07.1976	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	2♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	aberration?
<i>Kybos virgator</i>	6♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	26♂	Slovakia, Slovenský Kras	09.07.1976	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	3♂	Slovakia, Oslany	10.07.1977	P. Lauterer	MZMB	"volgensis"
<i>Kybos virgator</i>	4♂	Slovenia, Hudicevec	25.06.2001	H. Löcker	Coll. H. Löcker	
<i>Kybos virgator</i>	1♂	Slovenia, Hudicevec	11.09.2001	H. Löcker	Coll. H. Löcker	
<i>Kybos virgator</i>	1♂	Switzerland, Riehen	20.06.2000	D. Wyniger	NHMB	
<i>Kybos virgator</i>	1♂	Switzerland, Riehen	20.06.2000	D. Wyniger	NHMB	"volgensis"
<i>Kybos virgator</i>	1♂	Switzerland, Basel	22.06.2000	R. Mühlethaler	NHMB	
<i>Kybos virgator</i>	2♂	Switzerland, Basel	22.06.2000	R. Mühlethaler	NHMB	"volgensis"
<i>Kybos virgator</i>	3♂	Switzerland, Basel	27.07.2000	R. Mühlethaler	NHMB	
<i>Kybos virgator</i>	1♂	Switzerland, Basel	27.07.2000	R. Mühlethaler	NHMB	"volgensis"
<i>Kybos virgator</i>	1♂	Switzerland, Basel	25.08.2000	R. Mühlethaler	NHMB	"volgensis"
<i>Kybos virgator</i>	4♂	Switzerland, Basel	21.05.2002	R. Mühlethaler	NHMB	
<i>Kybos virgator</i>	1♂	Switzerland, Basel	13.07.2002	R. Mühlethaler	NHMB	"volgensis"
<i>Kybos virgator</i>	1♂	Slovenia, Tolmin	24.07.2001	H. Löcker	Coll. H. Löcker	

DNA Samples

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos abstrusus</i>	1 ♀	Switzerland, Martigny	17.10.2005	Mühlethaler	NHMB	
<i>Kybos calyculus</i>	1 ♀	Czech Republic, Telnice	14.06.2005	Malenovský	NHMB	
<i>Kybos butleri</i>	1 ♂	United Kingdom, Cardiff	01.08.2005	Wilson	NHMB	
<i>Kybos butleri</i>	1 ♂	Czech Republic, Brno	24.09.2005	Lauterer	NHMB	
<i>Kybos butleri</i>	1 ♂	Czech Republic, Brno	24.09.2005	Lauterer	NHMB	
<i>Kybos butleri</i>	1 ♂	Czech Republic, Brno	24.09.2005	Lauterer	NHMB	
<i>Kybos butleri</i>	1 ♂	United Kingdom, Cardiff	01.08.2005	Wilson	NHMB	
<i>Kybos butleri</i>	1 ♂	United Kingdom, Cardiff	01.08.2005	Wilson	NHMB	only 16S
<i>Kybos calyculus</i>	1 ♂	Switzerland, Bruderholz	03.07.2005	Mühlethaler	NHMB	
<i>Kybos calyculus</i>	1 ♂	Czech Republic, Telnice	14.07.2005	Malenovský	NHMB	
<i>Kybos calyculus</i>	1 ♂	Czech Republic, Telnice	14.07.2005	Malenovský	NHMB	
<i>Kybos dentifera</i>	1 ♂	Kazachstan		Dietrich	NHMB	
<i>Kybos digitatus</i>	1 ♂	Germany, Weil am Rhein	30.08.2005	Mühlethaler	NHMB	
<i>Kybos digitatus</i>	1 ♂	Germany, Weil am Rhein	01.06.2005	Mühlethaler	NHMB	
<i>Kybos iliensis</i>	1 ♂	Kazachstan		Dietrich	NHMB	
<i>Kybos lindbergi</i>	1 ♂	Switzerland, Bruderholz	03.07.2005	Mühlethaler	NHMB	
<i>Kybos smaragdula</i>	1 ♂	Czech Republic, Telnice	14.07.2005	Malenovský	NHMB	
<i>Kybos lindbergi</i>	1 ♂	Switzerland, Samedan	03.09.2005	Mühlethaler	NHMB	
<i>Kybos smaragdula</i>	1 ♂	Czech Republic, Telnice	14.07.2005	Malenovský	NHMB	
<i>Kybos smaragdula</i>	1 ♂	Czech Republic, Brno	18.06.2005	Lauterer	NHMB	
<i>Kybos calyculus</i>	1 ♀	Luxembourg, Michelau	27.08.2005	Malenovský	NHMB	
<i>Kybos ludus</i>	1 ♂	USA, Berkeley	09.08.2005	Mühlethaler	NHMB	
<i>Kybos mesasiaticus</i>	1 ♂	Kazachstan		Dietrich	NHMB	"verbae"
<i>Kybos mucronatus</i>	1 ♀	Switzerland, Samedan	03.09.2005	Mühlethaler	NHMB	
<i>Kybos mucronatus</i>	1 ♂	Germany, Füssen	04.08.2003	Nickel	NHMB	
<i>Kybos mucronatus verbae</i>	1 ♀	Switzerland, Samedan	03.09.2005	Mühlethaler	NHMB	
<i>Kybos perplexus</i>	1 ♂	Switzerland, Grellingen	28.05.2005	Mühlethaler	NHMB	
<i>Kybos perplexus</i>	1 ♂	Germany, Weil am Rhein	01.06.2005	Mühlethaler	NHMB	
<i>Kybos populi</i>	1 ♂	Czech Republic, Telnice	14.06.2005	Malenovský	NHMB	
<i>Kybos rufescens</i>	1 ♂	Germany, Weil am Rhein	30.08.2005	Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♂	Switzerland, La Punt	13.09.2002	Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♀	Switzerland, Samedan	03.09.2005	Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♀	Switzerland, Martigny	17.10.2005	Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♂	Slovenia, Ljubljana	01.07.2004	Mühlethaler	NHMB	
<i>Kybos rufescens</i>	1 ♂	Germany, Weil am Rhein	01.06.2005	Mühlethaler	NHMB	

Species	Individuals	Locality	Collecting date	Collector	Depository	Notes
<i>Kybos rufescens</i>	1♂	Germany, Weil am Rhein	01.06.2005	Mühlethaler	NHMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Dvory	26.06.2005	Malenovský	NHMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Dvory	26.06.2005	Lauterer	NHMB	
<i>Kybos smaragdula</i>	1♂	Germany, Füssen	26.06.2003	Nickel	NHMB	only 16S
<i>Kybos smaragdula</i>	1♂	Germany, Füssen	19.08.2003	Nickel	NHMB	
<i>Kybos soosi</i>	1♂	Kazachstan		Dietrich	NHMB	
<i>Kybos soosi</i>	1♀	Kazachstan		Dietrich	NHMB	
<i>Kybos</i> sp.	1♀	USA, Muir Woods	14.08.2005	Mühlethaler	NHMB	
<i>Kybos</i> sp.	1♀	Kazachstan,		Dietrich	NHMB	
<i>Kybos strigilifer</i>	1♂	Switzerland, Grellingen	28.05.2005	Mühlethaler	NHMB	
<i>Kybos smaragdula</i>	1♂	Czech Republic, Bile Karpaty	24.07.2005	Malenovský	NHMB	
<i>Kybos strigilifer</i>	1♀	Luxembourg, Michelau	27.08.2005	Malenovský	NHMB	
<i>Kybos strigilifer</i>	1♂	Germany, Füssen	19.08.2003	Nickel	NHMB	
<i>Kybos strobli</i>	1♂	Germany, Füssen	04.09.2003	Nickel	NHMB	only 16S
<i>Kybos virgator</i>	1♂	Switzerland, Grellingen	28.05.2005	Mühlethaler	NHMB	
<i>Kybos virgator</i>	1♀	Switzerland, Samedan	03.09.2005	Mühlethaler	NHMB	
<i>Kybos virgator</i>	1♂	Germany, Weil am Rhein	30.08.2005	Mühlethaler	NHMB	
<i>Kybos virgator</i>	1♂	Switzerland, Grellingen	28.05.2005	Mühlethaler	NHMB	"volgensis"
<i>Kybos virgator</i>	1♂	Germany, Füssen	26.06.2003	Nickel	NHMB	"volgensis", only 16S
<i>Empoasca vitis</i>	1♀	Germany, Weil am Rhein	23.09.2005	Mühlethaler	NHMB	
<i>Kyboasca bipunctata</i>	1♂	Slovenia, Ljubljana	01.07.2004	Mühlethaler	NHMB	

Appendix 1. Material examined for morphological and molecular studies. Abbreviations: AMNH = American Museum of Natural History, USA; INER = Istituto Nazionale di Entomologia Roma, Italy; NHMB = Naturhistorisches Museum Basel, Switzerland; NMP = Národní muzeum v Praze, Czech Republic; MZMB = Moravské zemské muzeum v Brně, Czech Republic; OSU = Ohio State University, USA.

[illegible]

[illegible]

	410	420	430	440	450	460	470	480	490	500
P. spumariu	TATAGGGCAGTATTTGCAATTTTAGGAGATTTTATTCCAATGATACCCCTTTATTTACTGGATTAACTAAATGAAATTCCTCAAAAATGGTTAAAAATACAAATTTATATA									
bipunctata	A...T									

	510	520	530	540	550	560	570	580	590	600
P. spumariu	ATTAATTTTC	TTGGG	TGAACCTTAACATTTTTT	CCCTCAACATTTTTT	TTAGCTTTTAAAGAGGAATACCTCGT	CGATATCTGATTTACCCAGATGCTTACATAT				
bipunctata	T.A.	T.TT.	T.	C.	G.	TTT.A.	T.T.	C.T.	T.	CT.
E.vitis130	T.	A.T.	T.		C.TTAT.	TTT.A.	T.T.	C.	T.	TTC.
abstrusus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
butleri130	C.G.	T.T.	T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
butleri141	C.	T.C.	A.TT.	T.C.	G.GTAT.	T.T.	T.T.	T.	T.	TTC.
butleri149	C.	T.C.	A.TT.	T.C.	G.GTAT.	T.T.	T.T.	T.	T.	TTC.
butleri149	C.G.	T.T.	T.TT.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
butleri149	C.G.	T.T.	T.TT.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
calyculus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
calyculus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
calyculus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
calyculus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
calyculus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
calyculus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
dentiferal	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
digitatus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
digitatus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	C.A.T.	TTC.
Kybos sp.1	C.	T.	A.TT.	C.	G.	TAT.	GT.T.C.	T.T.	T.	TTC.
Kybos sp.1	C.G.	T.C.	T.	C.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
lindbergil	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.G.	T.T.	T.	T.	TTC.
lindbergil	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
ludus1308	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
mesasiatic	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
mucronatus	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
mucronatus	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
mucronatus	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
perplexus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
perplexus1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
populi1418	C.	T.A.	A.TT.	C.	C.	A.TAT.	GT.T.C.	T.T.	T.	TTC.
rufescens1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
rufescens1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
rufescens1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
rufescens?	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
rufescens?	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
rufescens?	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
smaragdulu	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
smaragdulu	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
smaragdulu	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
smaragdulu	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.A.	T.T.	T.	T.	TTC.
soosi1338	C.G.	T.G.	T.T.	C.A.	A.TAT.	GT.T.A.	T.T.	T.	T.	TTC.
soosi?1339	C.G.	T.C.	T.T.	C.A.	A.TAT.	GT.T.A.	T.T.	T.	T.	TTC.
strigilife	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
strigilife	C.G.	T.C.	T.T.	C.A.	A.TAT.	GT.T.C.	T.T.	T.	T.	TTC.
strigilife	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
virgator13	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
virgator13	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.
volgensis1	C.G.	T.C.	T.T.	C.A.	A.GTAT.	GT.T.C.	T.T.	T.	T.	TTC.

[illegible]

[illegible]

[illegible]

[illegible]

Species	West Palearctic	East Palearctic	West Nearctic	East Nearctic	Salix	Populus	Alnus	Betula
<i>abstrusus</i> (Linnavuori, 1949)	x					x		
<i>acodens</i> DeLong, 1931			?	?				
<i>adunca</i> DeLong, 1931			?	?				
<i>aetnicola</i> Wagner, 1959	x							x
<i>alberta</i> (Ross, 1963)			x		x			
<i>albilacustris</i> Tishechkin, 2002	x				x			
<i>albolinea</i> Gillette, 1898				x	x			
<i>alexanderiae</i> (Ross, 1963)			x		x			
<i>altaicus</i> Mitjaev, 1963		x						
<i>amiciis</i> (Ross, 1963)			x					
<i>andresia</i> (Ross, 1963)			x	x	x			
<i>angustifoliae</i> (Ross, 1963)			x			x		
<i>annella</i> Hartzell, 1923			x		x			
<i>apata</i> DeLong & Davidson, 1936			?	?				
<i>arizona</i> (Ross, 1963)			x			x		
<i>aureoviridis</i> Uhler, 1877			x		x			
<i>auricillatus</i> Diabola, 1963		x						x
<i>austriacus</i> (Wagner, 1949)	x							x
<i>butleri</i> (Edwards, 1908)	x		(x)		x			
<i>caesari</i> (Hamilton 1972)			x					
<i>calyculus</i> (Cerutti, 1939)	x							x
<i>candelabricus</i> Diabola, 1958	x				x			
<i>carsona</i> DeLong & Davidson, 1936			x			x		
<i>cascada</i> (Ross, 1963)			x		x	x		
<i>chadchalicus</i> Diabola, 1967		x			x			
<i>chapingo</i> (Ross, 1963)			x		x			
<i>chromata</i> Davidson & DeLong, 1942			?	?				
<i>clinata</i> (Ross, 1963)			x	x	x			
<i>clypeata</i> Gillette & Baker, 1895				x				
<i>columbiana</i> (Hamilton, 1972)			x					
<i>confusa</i> DeLong & Davidson, 1936			?	?				
<i>consecta</i> (Ross, 1963)			x		x			
<i>copula</i> DeLong, 1931			x	x		x		
<i>cornutus</i> Dworakowska, 1968		x			x			
<i>coronata</i> (Hamilton, 1972)			x	x		x		
<i>crystola</i> (Ross, 1963)			x	x	x			
<i>denticula</i> Gillette, 1898			?	?				
<i>dentifera</i> (Logvinenko, 1980)		x			x			
<i>digita</i> (DeLong, 1931)			?	?				
<i>digitatus</i> (Ribaut, 1936)					x			
<i>dissimularis</i> (Hamilton, 1972)	x		x			x		
<i>dlabolat</i> Dworakowska, 1973		x						
<i>dworakowskii</i> Mitjaev, 1980		x			x			
<i>empusa</i> (Hamilton, 1972)			x					
<i>excava</i> Davidson & DeLong, 1938			?	?				
<i>exiguae</i> (Ross, 1963)			x		x			
<i>fontana</i> (Ross, 1963)			x		x	x		
<i>gelbata</i> DeLong & Davidson, 1936			x			x		
<i>gleditsia</i> DeLong & Davidson, 1936			?	?				
<i>gribisa</i> (Ross, 1963)			x			x		
<i>grossata</i> DeLong & Davidson, 1936			?	?				

Species	West Palaearctic	East Palaearctic	West Nearctic	East Nearctic	Salix	Populus	Alnus	Betula
<i>hartzelli</i> Baker, 1925			x					
<i>huangheicus</i> (Kuoh, 1981)		x						
<i>humilis</i> (Ross, 1963)			x		x			
<i>iliensis</i> Mitjaev, 1971	x				x			
<i>improcera</i> (Ross, 1963)			x					
<i>incida</i> DeLong, 1931			x		x	x		
<i>isotoma</i> (Logvinenko, 1980)		x			x			
<i>ivanovi</i> (Logvinenko, 1980)	x							
<i>facinta</i> DeLong & Davidson, 1936								
<i>koreanus</i> Matsumura, 1931		x				x		
<i>laurifoliae</i> Mitjaev, 1999		x				x		
<i>limpidus</i> (Wagner, 1955)	x				x			
<i>lindbergi</i> (Limavuori, 1951)	x							x
<i>livingstonii</i> Gillette, 1898			x					
<i>lucidae</i> (Ross, 1963)			x		x			
<i>ludus</i> (Davidson & DeLong, 1938)	x		(x)	(x)				x
<i>mesasiaticus</i> (Zachvatkin, 1953)	x	x			x			
<i>mesolinea</i> Davidson & DeLong, 1939			?	?				
<i>mitjaevi</i> Dworakowska, 1976	x				x			
<i>micronatus</i> (Ribaut, 1933)	x						x	
<i>niveicolor</i> Zachvatkin, 1953	x							
<i>obrudens</i> DeLong, 1932			?	?		x		
<i>obtusa</i> Walsh, 1862			?	?				
<i>occidua</i> (Ross, 1963)			x		x			
<i>osborni</i> Hartzell, 1923			?	?				
<i>oshamini</i> (Zachvatkin, 1953)	x				x			
<i>ovalis</i> (Ross, 1963)			?	?				
<i>paraltaica</i> (Orosz, 1996)			?	?				
<i>patula</i> DeLong, 1931			x		x			
<i>pectinata</i> DeLong, 1931			?	?				
<i>petaluma</i> (Ross, 1963)			x		x			
<i>petiolaridis</i> (Ross, 1963)			x	x?	x			
<i>populi</i> (Edwards, 1908)	x					x		
<i>portola</i> (Ross, 1963)			x			x		
<i>pyramidalis</i> Mitjaev, 1971	x					x		
<i>reddina</i> (Ross, 1963)			x		x			
<i>rossi</i> (Hamilton, 1972)			x			x		
<i>rubrafacia</i> DeLong, 1932			?	?				
<i>rubra</i> DeLong & Davidson, 1936			x			x		
<i>rubrovenosus</i> Dworakowska, 1973	x							
<i>rufescens</i> Melichar, 1896					x			
<i>salicis</i> Wheeler, 1937	x			x				
<i>saluta</i> DeLong, 1931								
<i>smaragdulus</i> (Fallén, 1806)	x	(x) ? x	(x)	(x)			x	
<i>soosi</i> Dworakowska, 1976		x						
<i>sordidulus</i> (Ossiannilsson, 1941)	x	x	x	x	x			
<i>sprita</i> (Ross, 1963)			x			x		
<i>stepposus</i> Vilbaste, 1980		x						
<i>strangula</i> Davidson & DeLong, 1939			x					
<i>strigilifer</i> (Ossiannilsson, 1941)	x				x			
<i>strobli</i> (Wagner, 1949)	x						x	

Species	West Palaearctic	East Palaearctic	West Nearctic	East Nearctic	Salix	Populus	Alnus	Betula
<i>sublactea</i> Van Duzee, 1917			?	?				
<i>tereholli</i> Vilbaste, 1980		x						
<i>tigris</i> (Hamilton, 1972)			x			x		
<i>topoli</i> Zachvatkin, 1953	x	x						
<i>transversa</i> Van Duzee, 1917								
<i>trifasciata</i> Gillette, 1898			x			x		
<i>ishagotaica</i> Vilbaste, 1980		x						
<i>unica</i> (Provancher, 1890)			?	?				
<i>velutina</i> (Ross, 1963)			x		x			
<i>virgator</i> (Ribaut, 1933)	x				x			
<i>yukonensis</i> (Ross, 1963)			x		x			
<i>ziona</i> DeLong & Davidson, 1936			?	?				

Roland Mühlethaler

- 06.04.1974 Geboren in Basel-Stadt. Schweizer Bürger.
- 1994 Humanistisches Gymnasiums Basel; Maturität Typus B.
- 1994-2001 Studium an der Universität Basel in den Fächern Geographie; Natur-, Landschafts- und Umweltschutz; Zoologie und Meteorologie.
- 1999 Halbjähriges Berufspraktikum am Forschungsinstitut für Biologischen Landbau (FiBL) in Frick (Schweiz) im Fachbereich Biologischer Pflanzenschutz.
- 2000-2001 Diplomarbeit "Untersuchungen zur Zikadenfauna der Lebensraumtypen von Basel-Stadt (Schweiz)" am Institut für Natur-, Landschafts- und Umweltschutz (NLU) – Biogeographie unter Leitung von Prof. Dr. P. Nagel.
Diplom in Geographie an der Uni Basel.
- 2002-2007 Dissertation unter der Leitung von Prof. Dr. P. Nagel (NLU – Biogeographie) und PD Dr. Daniel Burckhardt (Naturhistorisches Museum Basel) über die Taxonomie, Phylogenie und Biogeographie der mitteleuropäischen *Kybos*-Arten (Insecta, Hemiptera, Cicadellidae).